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농학박사학위논문

*Fusarium graminearum*의
cytochrome P450 효소들의 기능연구

**Functional characterization of cytochrome
P450 monooxygenases in the cereal head
blight fungus *Fusarium graminearum***

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신 지 영

**Functional characterization of cytochrome
P450 monooxygenases in the cereal head
blight fungus *Fusarium graminearum***

A dissertation submitted in partial
fulfillment of the requirement for
the degree of

DOCTOR OF PHILOSOPHY

to the Faculty of
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at

SEOUL NATIONAL UNIVERSITY

By

Jiyoung Shin

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지도교수 이 인 원

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농생명공학부 식물미생물학전공
신 지 영

신 지 영의 박사학위논문을 인준함
2017년 6월

위 원 장

박 은 우



부 위원장

이 인 원



위 원

김 국 형



위 원

채 순 기



위 원

반 용 선



A THESIS FOR THE DEGREE OF DOCTOR OF PHILOSOPHY

**Functional characterization of cytochrome
P450 monooxygenases in the cereal head
blight fungus *Fusarium graminearum***

UNDER THE DIRECTION OF
DR. YIN-WON LEE

SUBMITTED TO THE FACULTY OF THE GRADUATE SCHOOL
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BY
Jiyoung Shin

MAJOR IN PLANT MICROBIOLOGY
DEPARTMENT OF AGRICULTURAL BIOTECHNOLOGY

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APPROVED AS A QUALIFIED THESIS OF JIYOUNG SHIN
FOR THE DEGREE OF DOCTOR OF PHILOSOPHY
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Abstract

Functional characterization of cytochrome P450 monooxygenases in the cereal head blight fungus *Fusarium graminearum*

Jiyoung Shin

Major in Plant Microbiology

Department of Agricultural Biotechnology

The Graduate School

Seoul National University

Fusarium graminearum is a prominent plant pathogenic fungus causing *Fusarium* head blight in major cereal crops worldwide. To understand the molecular mechanisms underlying fungal development and virulence, large collections of *F. graminearum* mutants have been constructed. Cytochrome P450 monooxygenases (P450s) are widely distributed in organisms and are involved in a diverse array of molecular/metabolic processes; however, no systematic functional analysis of P450s has been attempted in filamentous fungi. In this study, we constructed a genome-wide deletion mutant set covering 102 P450s and analyzed

these mutants for changes in 38 phenotypic categories, including fungal development, stress responses, and responses to several xenobiotics, to build a comprehensive phenotypic dataset. Most P450 mutants showing defective phenotypes were impaired in a single phenotypic trait, demonstrating that my mutant library is a good genetic resource for further fungal genetic studies. In particular, we identified novel P450s specifically involved in virulence (5) and both asexual (1) and sexual development (2). Most P450s seem to play redundant roles in the degradation of xenobiotics in *F. graminearum*. This study is the first phenome-based functional analysis of P450s, and it provides a valuable genetic resource for further basic and applied biological research in filamentous fungi and other plant pathogens.

Keywords: *Fusarium graminearum*, Cytochrome P450 monooxygenase, Mutant library, Sexual development, Virulence, Azole fungicide sensitivity, Xenobiotic induction

Student Number: 2012-23383

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INTRODUCTION

Systematic gene knockout mutant libraries and the corresponding phenotypic datasets provide powerful tools for studying the molecular mechanisms underlying various biological processes and for identifying molecular targets that can be utilized for medical or agricultural applications. Genome-wide sets of gene deletions in model yeasts have given rise to a new era in the genetics of eukaryotic cells (Winzeler et al., 1999; Kim et al., 2010). Despite decades of progress in the genetics of filamentous fungi, their significance to public health, food security, and ecosystems has been increasing worldwide (Fisher et al., 2012). However, no whole-gene deletion mutant library is available for filamentous fungi, and genome-wide functional genetic analyses have mainly focused on signaling mediators, such as transcriptional regulators (Colot et al., 2006; Son et al., 2011b; Jung et al., 2015), kinases (Wang et al., 2011; De Souza et al., 2013; Lee et al., 2016), and phosphatases (Yun et al., 2015).

Cytochrome P450 enzymes (P450s) are proteins belonging to a superfamily containing heme as a cofactor and are found in most organisms, including fungi (Nelson et al., 1996). P450s commonly function as monooxygenases that transfer molecular oxygen to substrates and mediate diverse chemical reactions (e.g., hydroxylation, dealkylation, epoxidation, deamination, desulfuration, dehalogenation, sulfoxidation, and N-oxide reduction) (Sono et al., 1996; Mansuy, 1998). Therefore, P450 catalysis participates in various important cellular/metabolic processes, including the biosynthesis of hormones and cell

structure, secondary metabolism, fatty acid metabolism, and the degradation of xenobiotics (toxins, carcinogens, and pollutants) (Werck-Reichhart and Feyereisen, 2000).

Approximately 0.3% of genes in the animal kingdom encode P450s (Ingelman-Sundberg, 2005). The human genome contains 57 P450-encoding genes, which are well-known for their drug-metabolizing activities. The rice and *Arabidopsis thaliana* genomes contain 437 (0.8%) and 276 (0.8%) genes encoding putative P450s (Park et al., 2008). Some of these enzymes are known to be involved in the biosynthesis of various secondary metabolites that function as chemical defenses against microbial pathogens and herbivores (Bolwell et al., 1994; Anzenbacher and Anzenbacherova, 2001).

There are considerable disparities in P450 gene numbers among fungal species, depending on their lifestyle. Only three (0.06%), ten (0.16%), and six (0.09%) P450s have been identified in the budding yeast *Saccharomyces cerevisiae* and the opportunistic fungal pathogens *Candida albicans* and *Cryptococcus neoformans*, respectively (McCullough et al., 1996; Sabiiti and May, 2012). However, plant pathogenic fungi tend to harbor more P450 genes than other fungal species. The rice blast fungus *Magnaporthe oryzae* exhibits 107 P450s (0.97%), and *Cryphonectria parasitica*, which causes chestnut blight, displays 121 P450s (1.08%). Several fungal P450s have been shown to play critical roles in primary and secondary metabolism and the degradation of xenobiotics (Črešnar and Petrič, 2011). However, the involvement of P450s in fungal development and other

functions, such as xenobiotic detoxification, has rarely been elucidated, and there has been no systematic approach to the investigation of P450 genes in filamentous fungi, including plant pathogens.

The ascomycete fungus *Fusarium graminearum* is an economically important plant pathogen that causes head blight in major cereal crops, such as wheat, barley, and rice (Leslie and Summerell, 2006). In addition to yield losses, this fungus contaminates grains with mycotoxins (deoxynivalenol and zearalenone), which pose a serious threat to human and animal health (Desjardins and Proctor, 2007). The genome of *F. graminearum* contains 119 P450 genes (0.9%), although only eight P450s have been functionally characterized in this fungus. Three P450s (*TR11*, *TR14*, and *TR111*) are trichothecene biosynthetic genes (Brown et al., 2001; Kimura et al., 2007), and *Fg08079* and *CLM2* are required for the biosynthesis of butenolide (Harris et al., 2007) and culmorin (Bahadoor et al., 2016), respectively. Three other genes (*CYP51A*, *CYP51B*, and *CYP51C*) are paralogs of *CYP51*, which encodes an ergosterol biosynthetic enzyme (Liu et al., 2011; Fan et al., 2013).

In this study, I constructed a genome-wide P450 gene deletion mutant library for *F. graminearum*. Each mutant was analyzed in 23 phenotypic categories and under 15 xenobiotic induction conditions. My genetic resources and findings contribute to determining the functions of P450s in various cellular processes in filamentous fungi and in other lower eukaryotes.

MATERIALS AND METHODS

I. Fungal strains and media

The *F. graminearum* wild-type strain Z-3639 was used for generating deletion mutants (Bowden and Leslie, 1999). The wild-type and all transgenic strains were stored as mycelia and conidia in a 20% glycerol solution at -70 °C. The culture media were prepared and used according to the *Fusarium* laboratory manual (Leslie and Summerell, 2006).

II. DNA extraction Southern blotting and PCR

Genomic DNA was isolated from freeze-dried mycelium powder as previously described (Leslie and Summerell, 2006). Standard techniques were used for restriction endonuclease digestion, agarose gel electrophoresis, and Southern blot analysis (Sambrook and Russell, 2001). The primers employed in this study were synthesized by an oligonucleotide synthesis facility (Neo Probe, Daejeon, Korea; Supporting information Table S6). General PCR was performed following the manufacturer's instructions (Takara Bio Inc., Otsu, Japan).

Table 1. List and phenotypic summary of P450s in *F. graminearum*.

Locus ID	Gene name	Class	Best hit of Nelson's	Mycelia growth			Pigmentation	Sexual development				Conidiation	Toxin production		Virulence	Stress responses											Xenobiotics
				CM	PDA	MM		NP	PM	AF	AD		ZEA	DON		Osmotic	ROS	Fungicide	Cell wall	pH4	pH11	Heat	Cold	UV	Azole fungicides		
FGSG_00127.3	<i>Fg00127</i>	B-class P450	CYP645A1	4 ^a	4 ^b	4 ^c	4 ^d	4 ^e	4 ^f	4 ^g	4 ^h	4 ⁱ	4 ^j	4 ^j	4 ^k	4 ^l	4 ^l	4 ^l	4 ^l	4 ^l	4 ^l	4 ^l	4 ^l	4 ^l	4 ^l	4 ^l	4 ^m
FGSG_11585.3	<i>Fg11585</i>	B-class P450	CYP55A8	N/A ^o	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
FGSG_10960.3	<i>Fg10960</i>	B-class P450	N/M ^p	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_12284.3	<i>Fg12284</i>	Cytochrome P450	CYP587B1	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
FGSG_03542.3	<i>Fg03542</i>	Cytochrome P450	CYP526B1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_14006.3	<i>Fg14006</i>	Cytochrome P450	CYP570C2	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
FGSG_13739.3	<i>Fg13739</i>	Cytochrome P450	CYP631A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_12371.3	<i>Fg12371</i>	Cytochrome P450	CYP639A1	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
FGSG_01583.3	<i>Fg01583</i>	Cytochrome P450	CYP642A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	5 _{cc} ^q	3	4	
FGSG_13796.3	<i>Fg13796</i>	Cytochrome P450	CYP644A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_07859.3	<i>Fg07859</i>	Cytochrome P450	N/M	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_05334.3	<i>Fg05334</i>	Cytochrome P450	N/M	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_02668.3	<i>Fg02668</i>	Cytochrome P450	N/M	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	3	4	4
FGSG_09033.3	<i>Fg09033</i>	Cytochrome P450	N/M	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
FGSG_14007.3	<i>Fg14007</i>	Cytochrome P450	N/M	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_00863.3	<i>Fg00863</i>	Cytochrome P450	N/M	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_08377.3	<i>Fg08377</i>	E-class P450, group I	CYP544A3	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4

FGSG_12283.3	<i>Fg12283</i>	E-class P450, group I	CYP587B1	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
FGSG_02366.3	<i>Fg02366</i>	E-class P450, group I	CYP59B1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_03860.3	<i>Fg03860</i>	E-class P450, group I	CYP526B1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_09916.3	<i>Fg09916</i>	E-class P450, group I	CYP534C1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_01972.3	<i>Fg01972</i>	E-class P450, group I	CYP505A7	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	2
FGSG_07596.3	<i>Fg07596</i>	E-class P450, group I	CYP505C2	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_11303.3	<i>Fg11303</i>	E-class P450, group I	CYP547B1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_01048.3	<i>Fg01048</i>	E-class P450, group I	CYP617A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	3	4
FGSG_10461.3	<i>Fg10461</i>	E-class P450, group I	CYP617C1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_07977.3	<i>Fg07977</i>	E-class P450, group I	CYP5080A4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_02458.3	<i>Fg02458</i>	E-class P450, group I	CYP528A2	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_09351.3	<i>Fg09351</i>	E-class P450, group I	CYP531D1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_01745.3	<i>Fg01745</i>	E-class P450, group I	CYP532A3	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	3	4
FGSG_12737.3	<i>Fg12737</i>	E-class P450, group I	CYP537A2	4	4	4	4	4	4	4	4	4	4	4	2	4	4	4	4	4	4	4	4	4	4	4
FGSG_11655.3	<i>Fg11655</i>	E-class P450, group I	CYP548B1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_01739.3	<i>Fg01739</i>	E-class P450, group I	CYP551B1	3	3	3	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	3	4
FGSG_11038.3	<i>Fg11038</i>	E-class P450, group I	CYP552B3	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_02344.3	<i>Fg02344</i>	E-class P450, group I	CYP552C2	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_10695.3	<i>Fg10695</i>	E-class P450, group I	CYP567B1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_02419.3	<i>Fg02419</i>	E-class P450, group I	CYP570C1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	3	4

FGSG_02929.3	<i>Fg02929</i>	E-class P450_group I	CYP570C1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	3	4
FGSG_11498.3	<i>Fg11498</i>	E-class P450_group I	CYP570D1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_12534.3	<i>Fg12534</i>	E-class P450_group I	CYP573A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_13176.3	<i>Fg13176</i>	E-class P450_group I	CYP577A2	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_07804.3	<i>Fg07804</i>	E-class P450_group I	CYP627A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_03086.3	<i>Fg03086</i>	E-class P450_group I	CYP628A1	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
FGSG_11424.3	<i>Fg11424</i>	E-class P450_group I	CYP629A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_00012.3	<i>Fg00012</i>	E-class P450_group I	CYP630A1	4	4	4	4	4	4	4	4	4	4	2	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_08412.3	<i>Fg08412</i>	E-class P450_group I	CYP630B1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_03264.3	<i>Fg03264</i>	E-class P450_group I	CYP632A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_02367.3	<i>Fg02367</i>	E-class P450_group I	CYP643A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_01722.3	<i>Fg01722</i>	E-class P450_group I	CYP643A2	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	3	4
FGSG_12599.3	<i>Fg12599</i>	E-class P450_group I	CYP643B1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_03535.3	<i>TRI4</i>	E-class P450_group I	CYP58A3	4	4	4	4	4	4	4	4	4	0	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_03741.3	<i>Fg03741</i>	E-class P450_group I	CYP504A5	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_06447.3	<i>Fg06447</i>	E-class P450_group I	CYP504B3	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
FGSG_10571.3	<i>Fg10571</i>	E-class P450_group I	CYP5065A1	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
FGSG_11389.3	<i>Fg11389</i>	E-class P450_group I	CYP530A2	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_09086.3	<i>Fg09086</i>	E-class P450_group I	CYP619A1	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
FGSG_12568.3	<i>Fg12568</i>	E-class P450_group I	CYP620A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4

FGSG_03700.3	<i>Fg03700</i>	E-class P450, group I	CYP620B1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_00007.3	<i>CLM2</i>	E-class P450, group I	CYP620C1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_05113.3	<i>Fg05113</i>	E-class P450, group I	CYP621A1	3	3	3	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	3	4
FGSG_08809.3	<i>Fg08809</i>	E-class P450, group I	CYP580A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_02113.3	<i>Fg02113</i>	E-class P450, group I	CYP614A1	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
FGSG_08183.3	<i>Fg08183</i>	E-class P450, group I	CYP615A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_09671.3	<i>Fg09671</i>	E-class P450, group I	CYP616A1	4	4	4	4	1	1	0	0	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_02371.3	<i>Fg02371</i>	E-class P450, group I	CYP624A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_10991.3	<i>Fg10991</i>	E-class P450, group I	CYP625A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_11465.3	<i>Fg11465</i>	E-class P450, group I	CYP626A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_10629.3	<i>Fg10629</i>	E-class P450, group IV	CYP526F1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_02138.3	<i>Fg02138</i>	E-class P450, group IV	CYP540B2	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_03548.3	<i>Fg03548</i>	E-class P450, group IV	CYP540B3	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_02118.3	<i>Fg02118</i>	E-class P450, group IV	CYP617B1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_12342.3	<i>Fg12342</i>	E-class P450, group IV	CYP618A1	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
FGSG_01767.3	<i>Fg01767</i>	E-class P450, group IV	CYP527E1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	3	4
FGSG_08186.3	<i>Fg08186</i>	E-class P450, group IV	CYP552C3	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_04590.3	<i>Fg04590</i>	E-class P450, group IV	CYP566B1	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
FGSG_01959.3	<i>Fg01959</i>	E-class P450, group IV	CYP61A1	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
FGSG_03686.3	<i>Fg03686</i>	E-class P450, group IV	CYP61Ax	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4

FGSG_05960.3	<i>Fg05960</i>	E-class P450, group IV	CYP550B1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4, cc	4	4, cc	4
FGSG_03796.3	<i>Fg03796</i>	E-class P450, group IV	CYP633A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_08207.3	<i>Fg08207</i>	E-class P450, group IV	CYP633B1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_08191.3	<i>Fg08191</i>	E-class P450, group IV	CYP635A1	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
FGSG_03498.3	<i>Fg03498</i>	E-class P450, group IV	CYP637A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	3	4
FGSG_02114.3	<i>Fg02114</i>	E-class P450, group IV	CYP638A1	3	3	3	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	2	4
FGSG_12372.3	<i>Fg12372</i>	E-class P450, group IV	CYP639A1	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
FGSG_10910.3	<i>Fg10910</i>	E-class P450, group IV	CYP641A1	3	3	3	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_01000.3	<i>CYP51B</i>	E-class P450, group IV	CYP51F1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_04092.3	<i>CYP51A</i>	E-class P450, group IV	CYP51F2	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	1	4
FGSG_11024.3	<i>CYP51C</i>	E-class P450, group IV	CYP51F3	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_11536.3	<i>Fg11536</i>	E-class P450, group IV	CYP54C1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_01868.3	<i>Fg01868</i>	E-class P450, group IV	CYP602B1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_11282.3	<i>Fg11282</i>	E-class P450, group IV	CYP606B1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_08187.3	<i>Fg08187</i>	E-class P450, group IV	CYP622A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_02117.3	<i>Fg02117</i>	E-class P450, group IV	CYP623A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_09195.3	<i>Fg09195</i>	E-class P450, group IV	CYP623B1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_00071.3	<i>TRI1</i>	E-class P450, group IV	CYP68C2	4	4	4	4	4	4	4	4	4	0	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_02672.3	<i>Fg02672</i>	E-class P450, group IV	CYP68J1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	3	4
FGSG_04717.3	<i>Fg04717</i>	E-class P450, group IV	CYP68J2	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4

FGSG_11002.3	<i>Fg11002</i>	E-class P450, group IV	CYP68K1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_01786.3	<i>Fg01786</i>	E-class P450, group IV	CYP506A2	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_02872.3	<i>Fg02872</i>	E-class P450, group IV	CYP634A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_02111.3	<i>Fg02111</i>	E-class P450, group IV	CYP636A1	4	4	4	4	4	4	4	4	4	2	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_11108.3	<i>Fg11108</i>	E-class P450, group IV	CYP640A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_13797.3	<i>Fg13797</i>	E-class P450, group IV	CYP644A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_01284.3	<i>Fg01284</i>	P450, CYP52	CYP539A3	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_08005.3	<i>Fg08005</i>	P450, CYP52	CYP539E1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_08320.3	<i>Fg08320</i>	P450, CYP52	CYP584D1	4	4	4	4	5	2	0	0	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_13459.3	<i>Fg13459</i>	Pisatin demethylase- like	CYP5078A2	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_03260.3	<i>Fg03260</i>	Pisatin demethylase- like	CYP5080A1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_08079.3	<i>Fg08079</i>	Pisatin demethylase- like	CYP53A6	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	3	4
FGSG_13445.3	<i>Fg13445</i>	Pisatin demethylase- like	CYP53A7	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_10451.3	<i>Fg10451</i>	Pisatin demethylase- like	CYP53A8	3	3	3	4	4	4	4	4	4	2	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_02982.3	<i>Fg02982</i>	Pisatin demethylase- like	CYP531C1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_06068.3	<i>Fg06068</i>	Pisatin demethylase- like	CYP548A3	4	4	4	4	4	4	4	4	3	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_01740.3	<i>Fg01740</i>	Pisatin demethylase- like	CYP561D1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	3	4
FGSG_05806.3	<i>Fg05806</i>	Pisatin demethylase- like	CYP62B1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_03540.3	<i>TR111</i>	Pisatin demethylase- like	CYP65A2	4	4	4	4	4	4	4	4	4	0	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_07765.3	<i>Fg07765</i>	Pisatin demethylase- like	CYP65Q1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4

FGSG_03191.3	<i>Fg03191</i>	Pisatin demethylase-like	CYP65R1	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
FGSG_08023.3	<i>Fg08023</i>	Pisatin demethylase-like	CYP65S1	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A

^a Percentage of average radial growth on potato complete media (CM) of deletion mutants compared to wild-type strain was scored (5, more than wild-type strain; 4, 91-100%; 3, 61-90%; 2, 31-60 %; 1, 0-30 %; 0, 0 %).

^b Percentage of average radial growth on potato dextrose agar media (PDA) of deletion mutants compared to wild-type strain was scored (5, more than wild-type strain; 4, 91-100%; 3, 61-90%; 2, 31-60 %; 1, 0-30 %; 0, 0 %).

^c Percentage of average radial growth on minimal media (MM) of deletion mutants compared to wild-type strain was scored (5, more than wild-type strain; 4, 91-100%; 3, 61-90%; 2, 31-60 %; 1, 0-30 %; 0, 0 %).

^d Pigmentation representing aurofusarin accumulation after 5 days of growth on PDA was scored by comparing with wild-type strain (5, more than wild-type strain; 4, similar to wild-type strain; 2, less than wild-type strain; 0, no pigmentation).

^e Number of perithecia (NP) was scored from “0” to “5” (1, less than 25% of the wild-type strain; 2, 25-50% less; 3, 50-75% less; 4, 75-100% less; 5, more than wild-type strain; 0, no perithecia).

^f Perithecia maturation (PM) was determined by size compared to wild-type strain (4, normal; 3, 1-2 day delayed; 2, 3-7 days delayed; 1, more than 7 days delayed maturation).

^g Ascospore formation (AF) represents the existence and morphology of ascospores (4, normal ascospores; 2, abnormal shape of ascospore; 0, no observable ascospores).

^h Ascospore discharge (AD) was determined by discharged number of ascospores compared to wild-type strain (4, normal; 2, reduced; 0, no discharge).

ⁱ Conidiation represents conidia production of mutants compared to wild-type strain (4, similar number to wild-type strain; 3, approximately 10-fold reduction; 2, approximately 100-fold reduction; 1, approximately 1000-fold reduction; 0, no visible conidia production).

^j Toxin production (ZEA, zearalenone; DON, deoxynivalenol) was quantified compared to wild-type strain (6, overproduction; 4, similar to wild-type strain; 2, reduced production; 0, no detectable production).

^k Virulence on wheat heads was scored (4, normal; 0, markedly reduced virulence compared to wild-type strain).

^l Radial growth of mutants on each stress-inducing media was evaluated compared to growth on mock complete media (CM) (2, susceptible; 4, normal; 6, resistant).

^m Radial growth of mutants on each xenobiotics media was evaluated compared to growth on mock complete media (CM) (2, susceptible; 4, normal; 6, resistant).

^o N/A: not applicable.

^p N/M: not matched.

^q Color change of pigment (cc)

III. Generation of P450 gene deletion mutants

DNA cassettes for targeted gene deletion were generated using the double-joint (DJ) PCR and split marker recombination methods (Catlett et al., 2003; Yu et al., 2004). The *GEN* was amplified from pII99 (Namiki et al., 2001), and the 5' and 3' regions of the target gene were amplified from Z-3639. Three amplicons (5' region, *GEN*, and 3' region) were fused via DJ PCR using previously described conditions (Yu et al., 2004), and fungal transformation was performed as previously described (Son et al., 2011a). At least three independent transformants were isolated for each gene deletion via co-dominant PCR screening. To exclude ectopic mutants, (P450 name)-5F and Gen-with 5F primers were used for PCR screening. To confirm a single copy integration of the mutant allele, Southern hybridization was performed (Supporting information Fig. S1).

IV. Asexual and sexual development

In most cases, two independent mutants for each gene deletion were used for phenotyping with three replications. Colony morphology and radial growth were assayed with complete medium (CM), potato dextrose agar (PDA) and minimal medium (MM) at 25 °C for three and five days after inoculation. The pigmentation of the bottom of the PDA plates (representing aurofusarin accumulation) after being cultured for five days was compared with the wild-type strain, and the intensity of the red color was scored (0-6) with the naked eye.

Conidium production was measured by counting the number of conidia with a

hemocytometer (Superior, Marienfeld, Germany) after culturing a culture plug of CM in 5 ml of carboxymethylcellulose medium (CMC) for five days at 25 °C on a rotary shaker (150 rpm). For the observation of conidium morphology, the conidia produced by each strain on yeast malt agar (YMA) were harvested, and differential interference contrast (DIC) images were obtained using a DE/Axio Imager A1 microscope (Carl Zeiss, Oberkochen, Germany).

For sexual development, each mutant was incubated on carrot agar at 25 °C for 5 days (Leslie and Summerell, 2006). Mycelia grown on carrot agar were mock fertilized with 700 µl of a 2.5% Tween 60 solution to induce sexual reproduction and were then incubated under a near-UV light (wavelength: 365 nm, HKiv Import & Export Co., Ltd., Xiamen, China) at 25 °C. Seven days after sexual induction, the maturation of perithecia was determined by measuring their size.

V. Virulence test and mycotoxin analysis

For the virulence test, the point inoculation method was employed, as previously described (Lee et al., 2009). The conidia of each strain were harvested from CMC and adjusted to 1×10^5 conidia/ml. Thereafter, 10 µl of each conidial suspension was injected into a center spikelet of a wheat head (cultivar Eunpamil) at mid-anthesis. The wheat plants were subsequently incubated in a humid chamber for 3 days and grown in a greenhouse for an additional 18 days. The first virulence test was performed

using two independent strains for each gene deletion and five wheat heads for each strain. Mutants showing typical FHB symptoms were excluded from the second virulence test. Infected wheat heads of reduced virulence ($p < 0.05$, t test) were imaged, and spikelets with head blight symptoms of ten wheat heads were counted in the second virulence test.

To determine the effect of P450 gene deletion on mycotoxin production, fungal strains were grown on 30 g of rice substrate for three weeks at 25 °C in the dark. Rice cultures were harvested, and mycotoxins were extracted as previously described (Seo et al., 1996). A portion of each extract was analyzed using TLC to estimate the amounts of DON and zearalenone. An ELISA test kit (Veratox® , Neogen Corp., Lansing, MI, USA) was applied for exact quantifications of DON and zearalenone in accordance with the manufacturer's instructions. The experiments were performed using two independent strains with three biological replicates for each gene deletion.

VI. Stress and azole fungicide sensitivity screening

A mycelial plug (5 mm in diameter) was collected from the periphery of a 3-day-old colony and placed on a CM plate supplemented with various stress agents as previously described (Son et al., 2011b). The tested stress conditions included oxidative stress (5 mM hydrogen peroxide and 0.1 mM menadione), osmotic stress (1 M NaCl, 1 M KCl, 1.5 M sorbitol, and 6 mM FeSO₄), pH stress (pH = 4 and pH

= 11), cell wall stress (60 mg/L Congo Red and 5 mg/L sodium dodecyl sulfate), inhibition of DNA synthesis (8.6 mg/L fungicide iprodione), inhibition of the mitogen activated protein kinase pathway (0.023 mg/L fungicide fludioxonil), inhibition of meiosis and intracellular transportation (0.65 mg/L fungicide benomyl), UV stress (365 nm; HKiv Import & Export Co.), and treatment with azole fungicides (0.025 mg/L tebuconazole, 0.05 mg/L difenoconazole, 0.1 mg/L epoxiconazole, 0.05 mg/L propiconazole, and 0.01 mg/L prochloraz) (Liu et al., 2011).

VII. Assay for xenobiotic induction

A total of 15 xenobiotics, including aliphatic (176 mM n-dodecane, 22 mM 1-dodecanol, and 10 mM levulinic acid), alicyclic (48.2 mM cyclohexanone), aromatic (15 mM methyl benzoate, 0.14 mM 4-nitrophenol, 0.53 mM phenoxyacetic acid, and 0.72 mM resorcinol), polyaromatic (0.32 mM benzo(a)pyrene, 0.15 mM 3-methyl cholanthrene, 0.14 mM 1-naphthol, 0.62 mM naphthalene, and 0.62 mM phenanthrene), and alkyl-substituted aromatic (0.06 mM sodium dodecylbenzenesulfonate and 0.1 mM 4-octylphenol) compounds, were tested for P450 transcriptional induction in liquid CM cultures. Xenobiotic chemicals of analytical grade were used, which were purchased from different commercial sources. CM cultures grown for 24 h at 25 °C on a rotary shaker (150 rpm) were spiked with individual chemicals, and mycelia were harvested after 24 h (Doddapaneni et al., 2005). The details of the tested xenobiotics and the molar

concentrations used are given in Supporting information Table S4. A parallel control culture treated with an equal volume of the solvent [dimethyl sulfoxide (DMSO) for solids, water for liquids] was also included.

VIII. Quantitative real time (qRT)-PCR

The total RNAs of the wild-type and P450 mutant strains were extracted from mycelia using the Easy-Spin Total RNA Extraction kit (iNtRON Biotech, Seongnam, Korea). First-strand cDNA was synthesized with the SuperScript III First-Strand Synthesis System (Invitrogen, Carlsbad, CA, USA). qRT-PCR was performed using iQ SYBR Green Master Mix (Bio-Rad, Hercules, CA, USA) and the 7500 real-time PCR system (Applied Biosystems, Foster City, CA, USA). For normalization, the endogenous housekeeping gene cyclophilin (*CYP1*; FGSG_07439.3) was used (Lin et al., 2011). The threshold cycle (ΔC_T) value of gene expression was subtracted from the ΔC_T value of each sample to obtain a $\Delta\Delta C_T$ value. The transcript level relative to the calibrator was expressed as $2^{-\Delta\Delta C_T}$ (Livak and Schmittgen, 2001). The qRT-PCR assays were repeated three times with two biological replicates.

RESULTS

I. Characterization and deletion of putative P450 genes in *F. graminearum*

To identify *F. graminearum* P450 genes, I employed the previously constructed Fungal Cytochrome P450 Database (Park et al., 2008). The *F. graminearum* genome contains 119 putative P450 genes, which cover 0.9% of the total genes that are similar to other plant pathogenic fungi (Supporting information Table S1). These P450s were classified into six classes based on InterPro terms (Fig. 1A). Approximately 70% of the P450s (87) were included in the “E-class P450” category, and 13 P450s were grouped into the “Pisatin demethylase” category. The P450 enzymes were also grouped into families based on amino acid sequence similarity (Supporting information Table S1).

As little is known about the genetic requirements for P450s in fungal development, I analyzed the profiles of transcript levels during sexual and asexual development (Fig. 2). RNA-seq results were obtained from previous studies (Sikhakolli et al., 2012; Son et al., 2013; Son et al., 2016) and visualized using ClustVis (Metsalu and Vilo, 2015). Based on their expression patterns, P450s were clustered into sexually (A group) and asexually (B group) induced gene groups. Intriguingly, group A genes were specifically induced during sexual reproduction but repressed during asexual reproduction, while the opposite was true for group B genes, demonstrating that different P450-mediated molecular processes are

involved in fungal reproduction processes in *F. graminearum*.

To examine the biological functions of the P450s, I attempted to generate deletion mutants for all of the P450 genes. Using the split-marker homologous recombination method, I successfully replaced 102 of the P450 genes with a geneticin resistance gene cassette (*GEN*). The exact genetic manipulations of the selected the geneticin resistance mutants were confirmed via co-dominant PCR screening, followed by Southern hybridization with a *GEN* probe (Supporting information Fig. S1). I failed to delete 17 genes in more than three independent trials, possibly due to lethality. At least two independent deletion mutants of each P450 gene were characterized for their phenotypes in terms of mycelial growth, pigmentation, conidiation, sexual development, virulence on wheat heads, mycotoxin production, responses to various stresses, fungicides (including azole fungicides), and several types of xenobiotic induction (Supporting information Table S1).

II. Effects of P450 mutation on vegetative growth and conidiogenesis

I compared the vegetative growth and pigmentation of each P450 mutant in nutrient-rich media (CM, complete medium; PDA, potato dextrose agar medium) and nutrient-starved medium (MM, minimal medium). Most of the P450 mutants displayed similar phenotypes for mycelial growth under the tested conditions, with the exception of five mutants (Supporting information Table S1 and Fig. 3). The

growth rates of these five mutants ($\Delta Fg02114$, $\Delta Fg01739$, $\Delta Fg05113$, $\Delta Fg10910$, and $\Delta Fg10451$) were slightly reduced compared with the wild-type strain (Fig. 3); however, they showed no difference in hyphal morphology.

Only one mutant was severely defective in asexual reproduction. The conidial production of the $\Delta Fg06068$ strains in carboxymethyl cellulose medium (CMC) was significantly reduced compared with the wild-type strain (Fig. 4A). Moreover, red pigment production during conidiation was mostly abolished in $\Delta Fg06068$, whereas the mutant normally accumulated red pigments during vegetative growth on PDA, demonstrating that the deletion of *Fg06068* led to malfunctions of several important cellular processes during conidiation (Supporting information Table S1 and Fig. 4B).

To determine how deletion of the *Fg06068* gene affects conidiation, microscopic observations were performed. The *F. graminearum* wild-type strain initially produced phialides from the hyphae, and mature phialides continuously produced conidia (Fig. 4C). Although some phialides were observable in $\Delta Fg06068$, the number of phialides was markedly reduced compared with the wild type, and most phialide cells were collapsed during conidiogenesis (Fig. 4C). However, the conidium morphologies of the $\Delta Fg06068$ and wild-type strains were indistinguishable.

To characterize the mechanistic roles of *Fg06068*, I analyzed the transcript levels of *Fg06068* and representative conidiation marker genes during conidiation.

The transcription of *Fg06068* was markedly induced after conidium induction in the wild-type strain (Fig. 4D). When I compared the transcript levels of representative conidiation-related genes between the wild-type and $\Delta Fg06068$ mutant strains (Ohara et al., 2004; Lysøe et al., 2011; Zheng et al., 2012), the transcript levels of all six genes (*STUA*, *HTF1*, *REN1*, *FLBC*, *ABAA*, and *WETA*) were observed to be significantly decreased in the $\Delta Fg06068$ mutants compared with the wild-type strain (Fig. 4D). Considering these findings together, I concluded that asexually induced *Fg06068* is required for phialide formation and function in *F. graminearum* and possibly participates in the regulation of the genetic pathway of conidiogenesis.

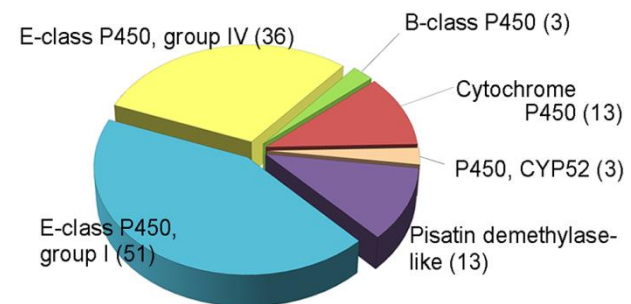
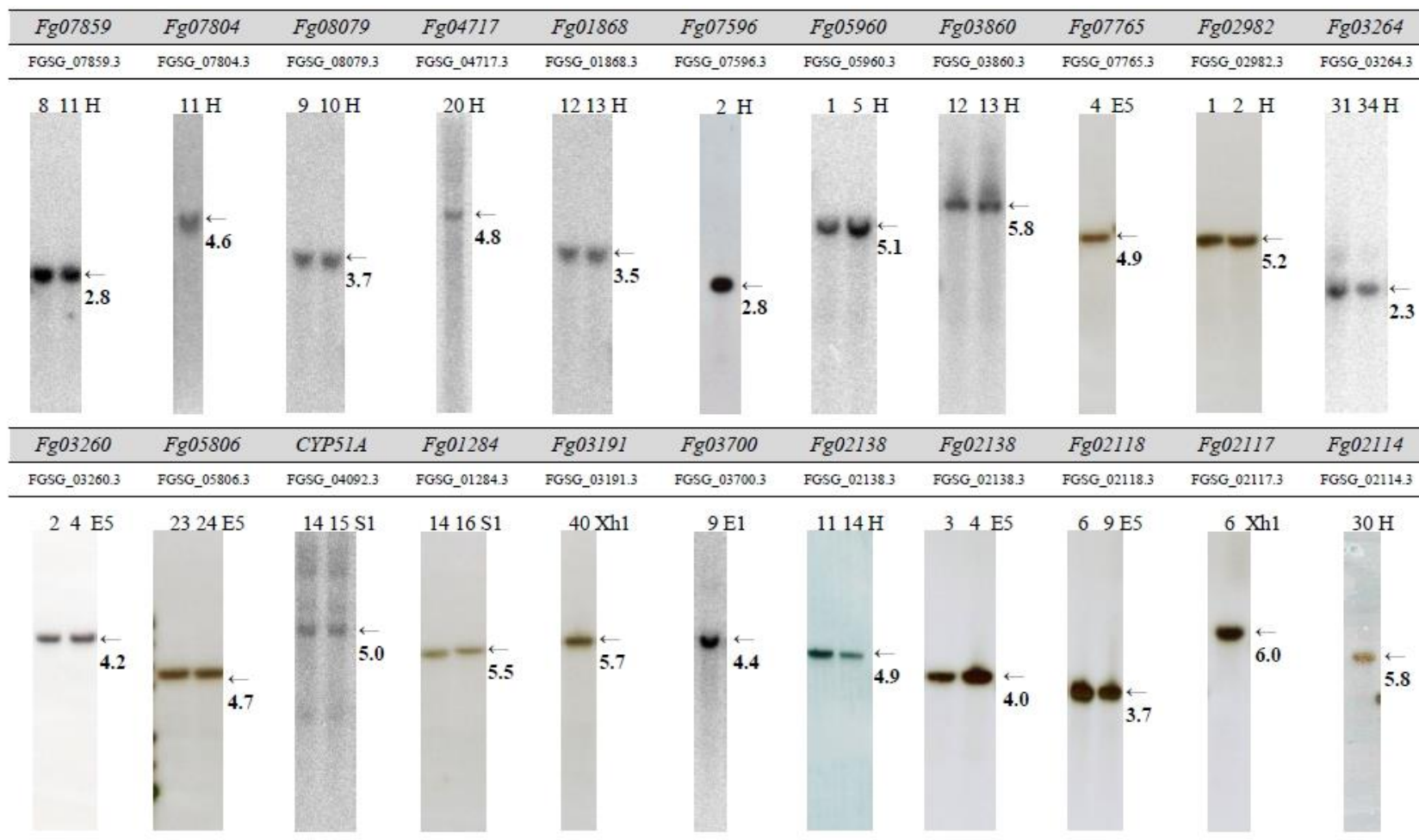
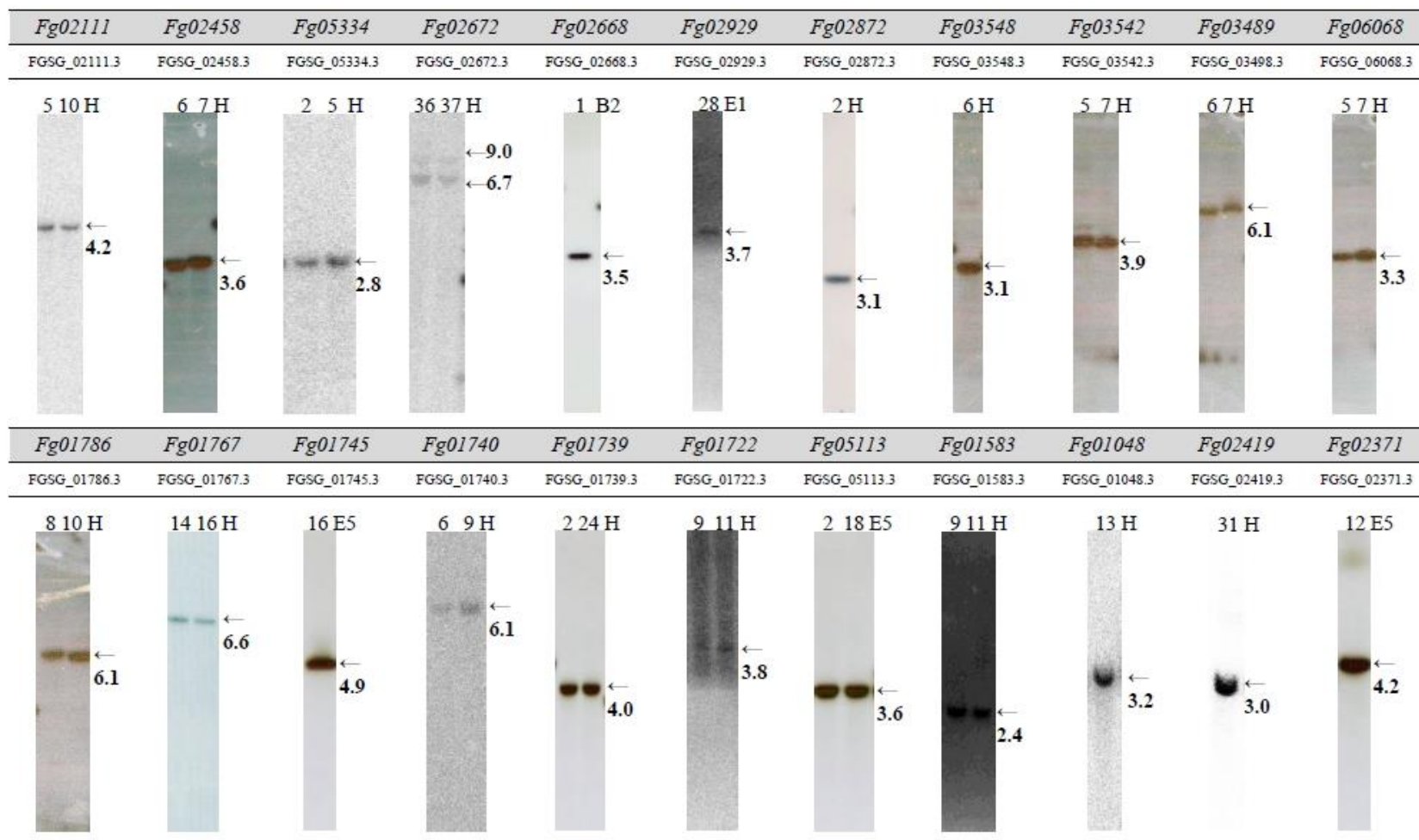
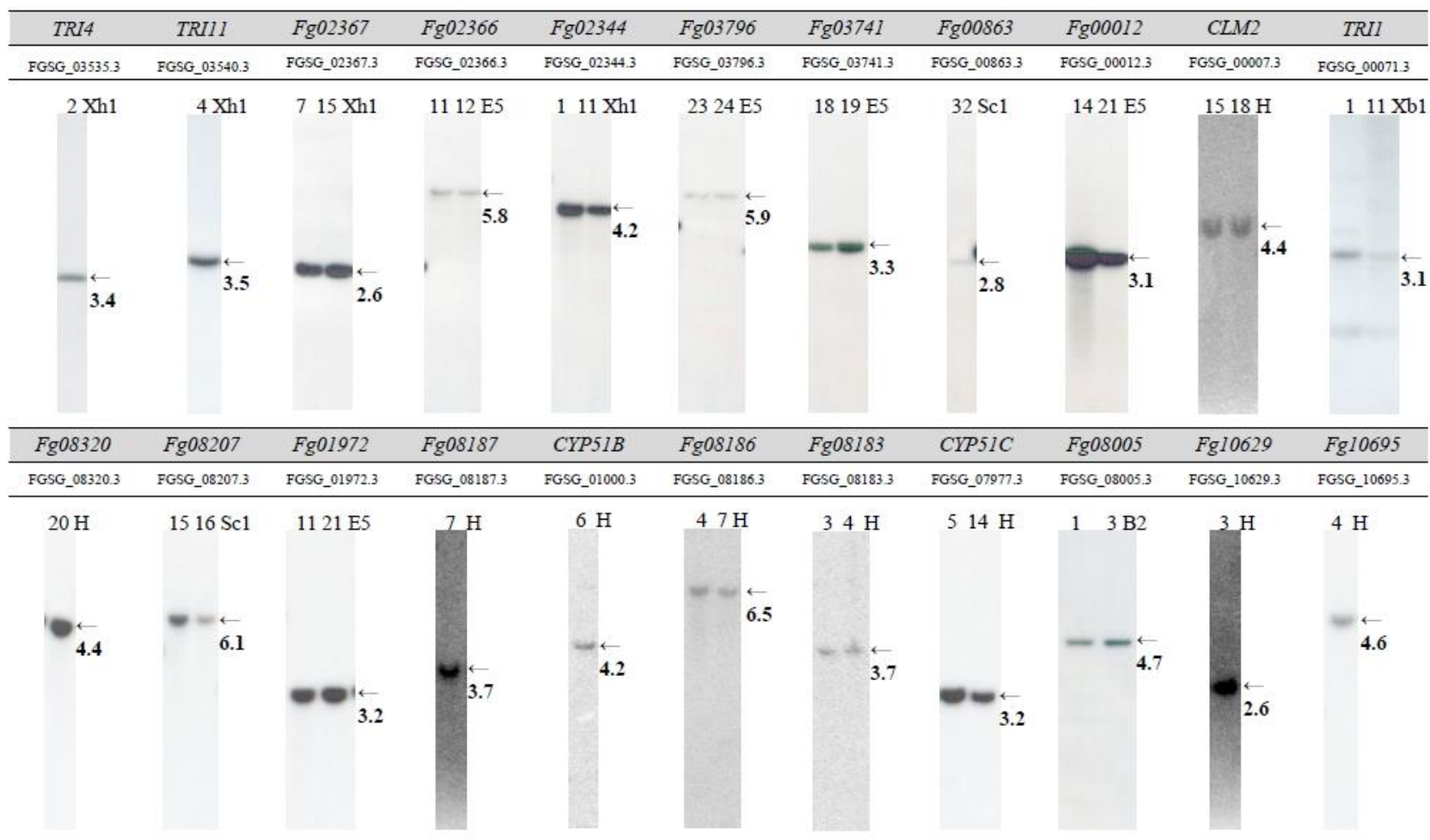
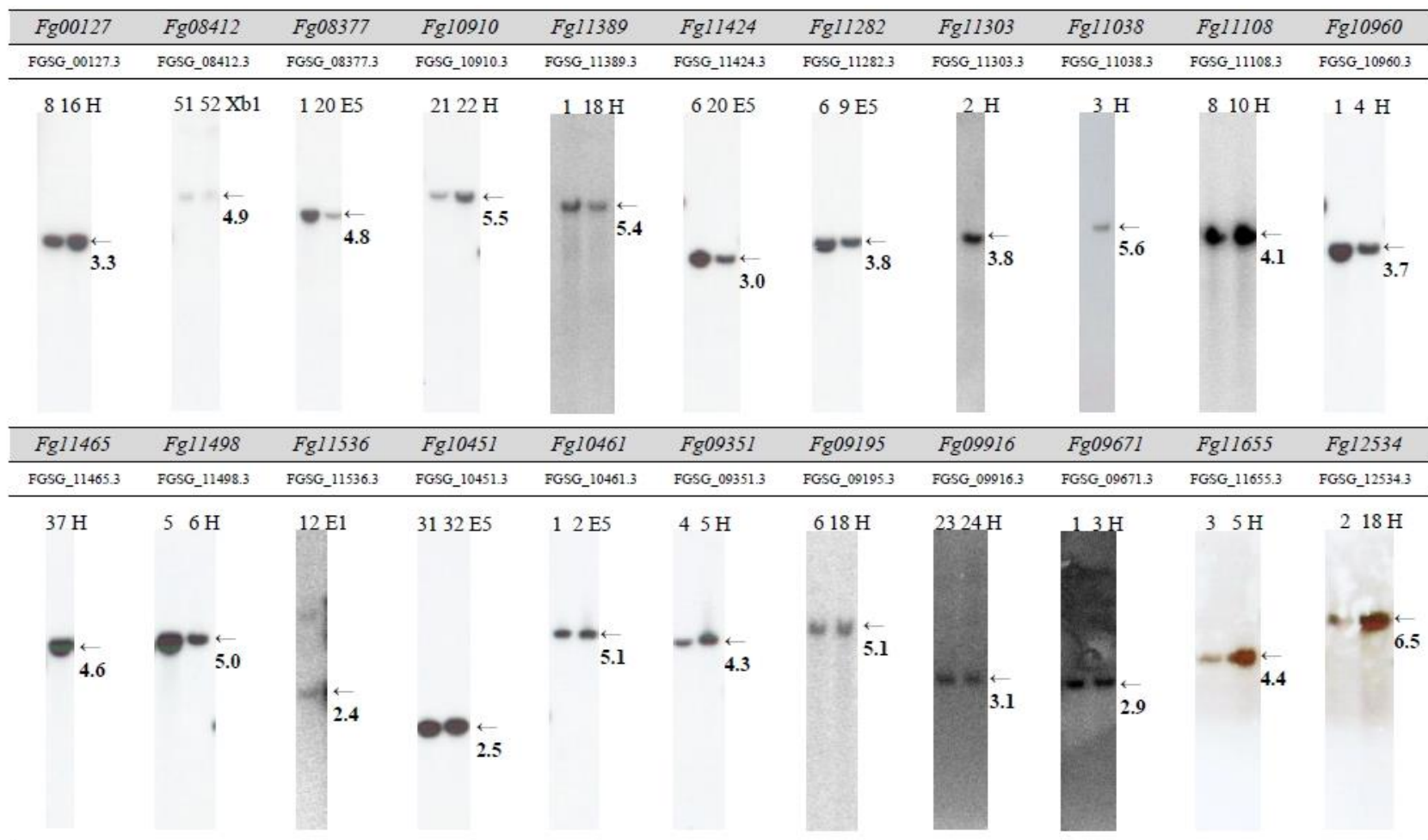


FIG 1. Classification of putative P450s in *F. graminearum*. The total P450s were categorized into six classes based on InterPro terms.









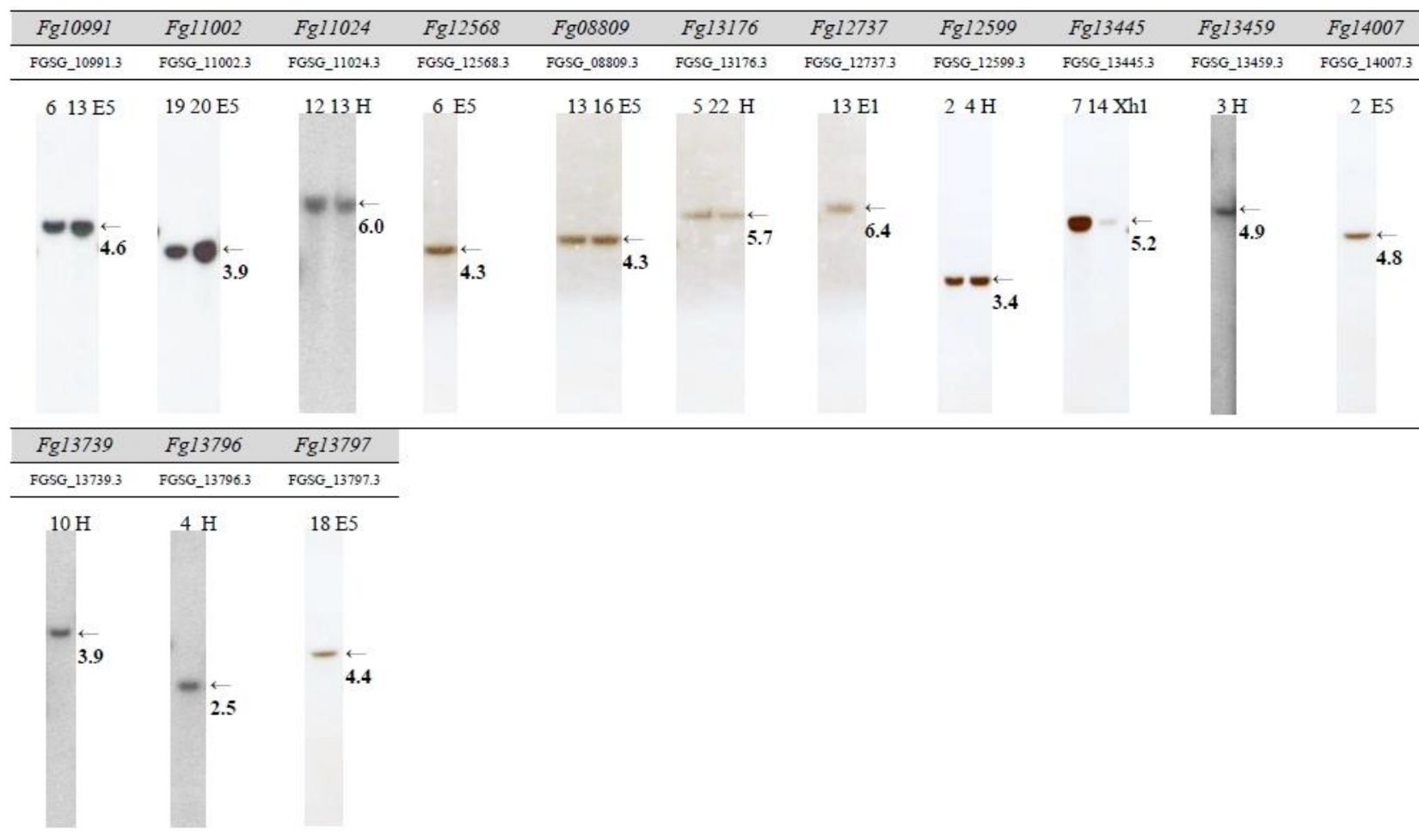


FIG 2. Confirmation of P450 deletion mutants by Southern blotting. The restriction enzymes used for each blot and the size of the DNA standards (kb) are indicated on the right of each blot. H, HindIII; E1, EcoRI, E5, EcoRV; B, BglII; S, SalI; Sc, SacI; Nd, NdeI; Nr, NruI; B1, BamHI; Xh, XhoI; Xb, XbaI; P1, PstI; C1, ClaI; SS1, SspI; K1, KpnI

III. Sexual development

As half of the P450s were upregulated during several steps of sexual reproduction (Fig. 2), I next sought to identify the P450s involved in these processes. However, most of the P450 deletion mutants appeared normal in the production of perithecia and sexual spores (ascospores); only two P450 mutants ($\Delta Fg08320$ and $\Delta Fg09671$) exhibited defects in sexual reproduction (Fig. 5). $\Delta Fg09671$ rarely produced perithecial initials, which did not mature further (Fig. 5A). While mycelial growth was mostly suppressed during sexual development in the wild-type strain, aerial mycelia grew upward and covered cultures of $\Delta Fg09671$. The transcript levels of *Fg09671* were significantly induced until 3 days after sexual induction and declined as the perithecia matured, suggesting that *Fg09671* is required for the initial stages of perithecial development in *F. graminearum* (Fig. 5B).

Deletion mutants of *Fg08320* produced smaller perithecia than the wild-type strain (Fig. 5A). Two days after sexual induction, both the wild-type and $\Delta Fg08320$ strains produced small perithecia with similar numbers (Fig. 5C and D). After 3 days of incubation, some of the small perithecia were selectively matured in the wild-type strain, and the others remained immature. However, $\Delta Fg08320$ produced a greater number of small perithecia than the wild type, and none of them matured (Fig. 5C and D). Finally, the average perithecial diameter produced by $\Delta Fg08320$ was approximately 110 μm , which was nearly 2-fold smaller than the perithecia produced by the wild-type strain ($p < 0.05$) (Fig. 5D). *Fg08320* transcripts accumulated dramatically 3 days after sexual induction and decreased as the

perithecia matured, similar to *Fg09671* (Fig. 5B). Thus, both P450 genes are specifically involved in perithecium development and/or ascospore formation in *F. graminearum*. In particular, *Fg08320* plays roles in the determination of perithecium numbers and selective perithecium maturation.

IV. Virulence and mycotoxin production

To evaluate the virulence of the P450 mutants, a conidial suspension of each strain was point-inoculated onto wheat spikelets at mid-anthesis. While the wild-type strains caused typical head blight symptoms 21 days after inoculation, five of the P450 deletion mutants ($\Delta Fg03700$, $\Delta Fg02111$, $\Delta Fg00012$, $\Delta Fg10451$, and $\Delta Fg12737$) showed significantly reduced virulence compared with the wild type (Fig. 6).

All of the P450 deletion mutants were inoculated into rice media to analyze the production of secondary metabolites, such as deoxynivalenol (DON) and zearalenone. The levels of DON and zearalenone biosynthesized by P450 mutant strains were not significantly different compared with the wild-type strain according to conventional thin-layer chromatography (TLC) analysis (Supporting information Fig. S2), which was further confirmed by ELISA (Supporting information Table S2).

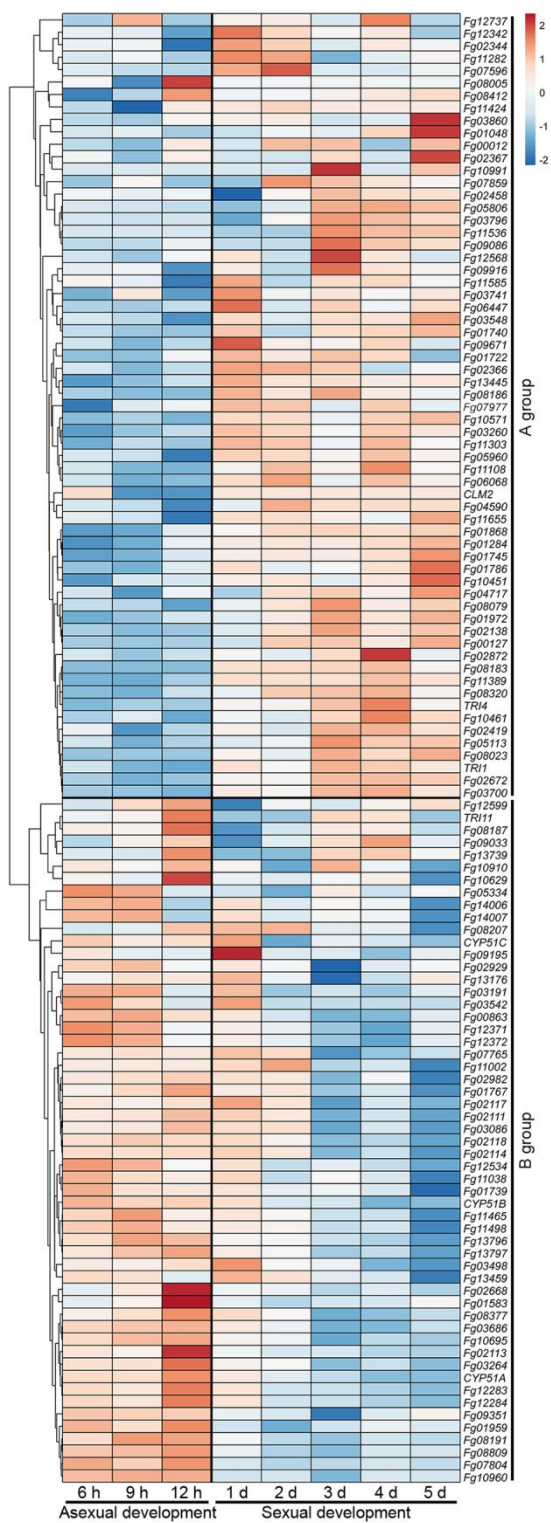


FIG 3. Heatmap visualization of P450 gene transcriptional profiles during asexual and sexual development. The heatmap depicts P450 gene transcript levels during asexual and sexual developmental time points based on Log2-based relative transcript abundances compared with 3 h (asexual development) and 2 h (sexual development). Red represents higher expression; blue represents lower expression; and the rows represent transcriptional units. RNA-seq results were obtained from previous studies (Sikhakolli et al., 2012; Son et al., 2013; Son et al., 2016) and visualized using ClustVis (Metsalu and Vilo, 2015).

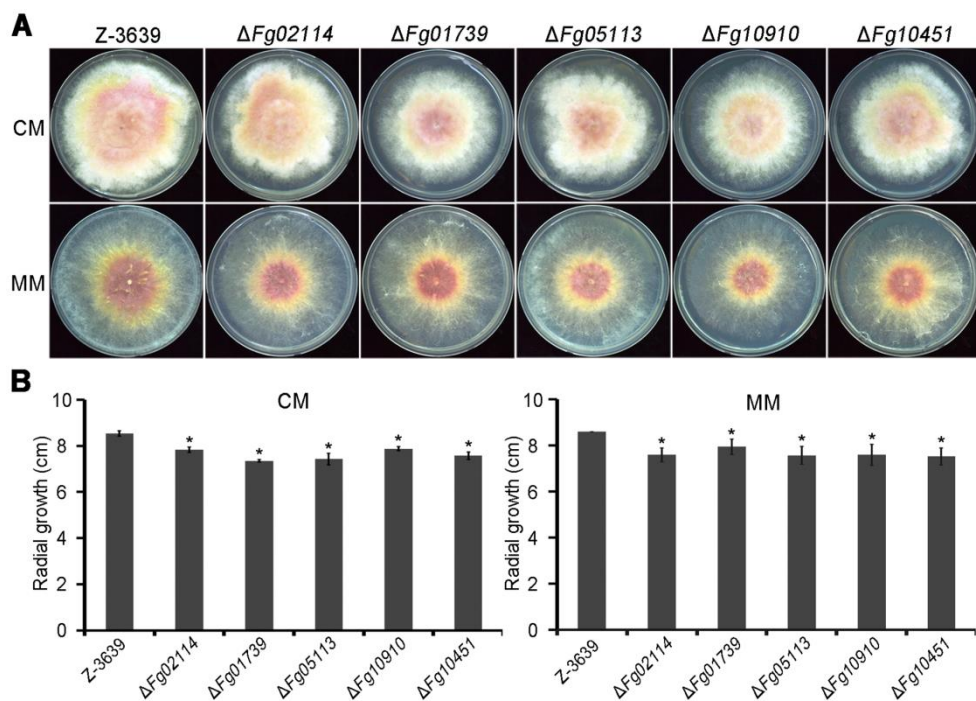


FIG 4. Vegetative growth of P450 mutants. A. Mycelial growth on complete medium (CM) and minimal medium (MM). Photographs were taken 5 days after inoculation. B. Radial growth on CM and MM. Radial growth was measured 5 days after inoculation. Significant differences ($p < 0.05$, t test) are indicated with an asterisk.

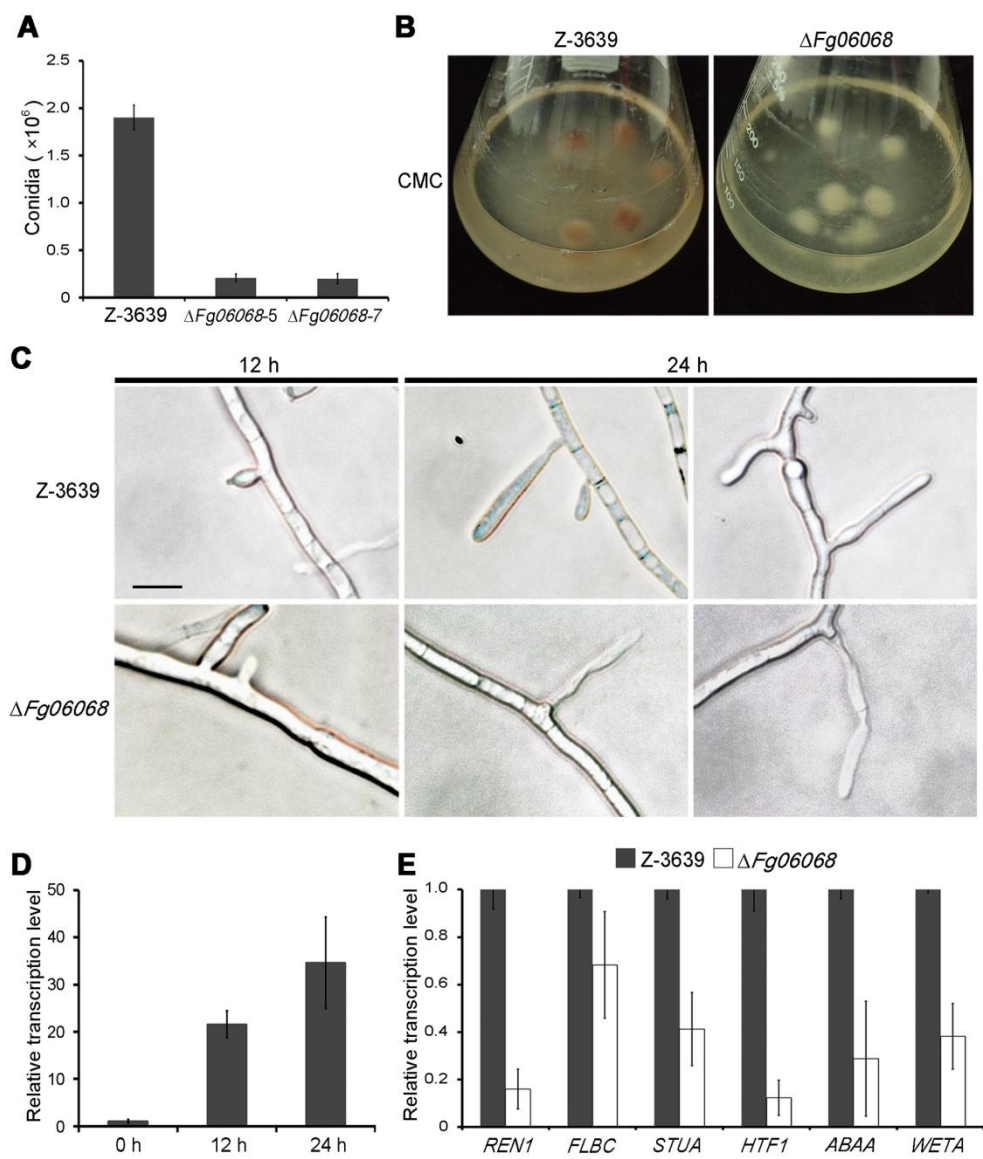


FIG 5. Defective conidiation of *Fg06068* deletion mutants. A. Conidium production. The number of conidia was counted at 5 days after inoculation in CMC. The values were generated based on three biological replicates. Significant differences ($p < 0.05$, t test) are indicated with an asterisk. B. Mycelial growth in CMC. The photographs were taken 3 days after inoculation. C. Morphology of conidiophores. The photographs were taken 1 to 2 days after conidium induction. Scale bar = 20 μm . D. Relative transcript levels of the *Fg06068* gene during asexual development. Total RNA was extracted from the wild type at 0, 6, 12, and 24 h after inoculation into CMC. E. Relative transcript levels of genes related to conidiation. Total RNA was extracted from the wild-type and $\Delta Fg06068$ strains at 18 h after inoculation into CMC. The relative transcript levels of each gene in the wild type were arbitrarily set to 1. Transcript levels were determined by qRT-PCR using two biological replicates with three technical repetitions each. Significant differences ($p < 0.05$, t test) are indicated with an asterisk.

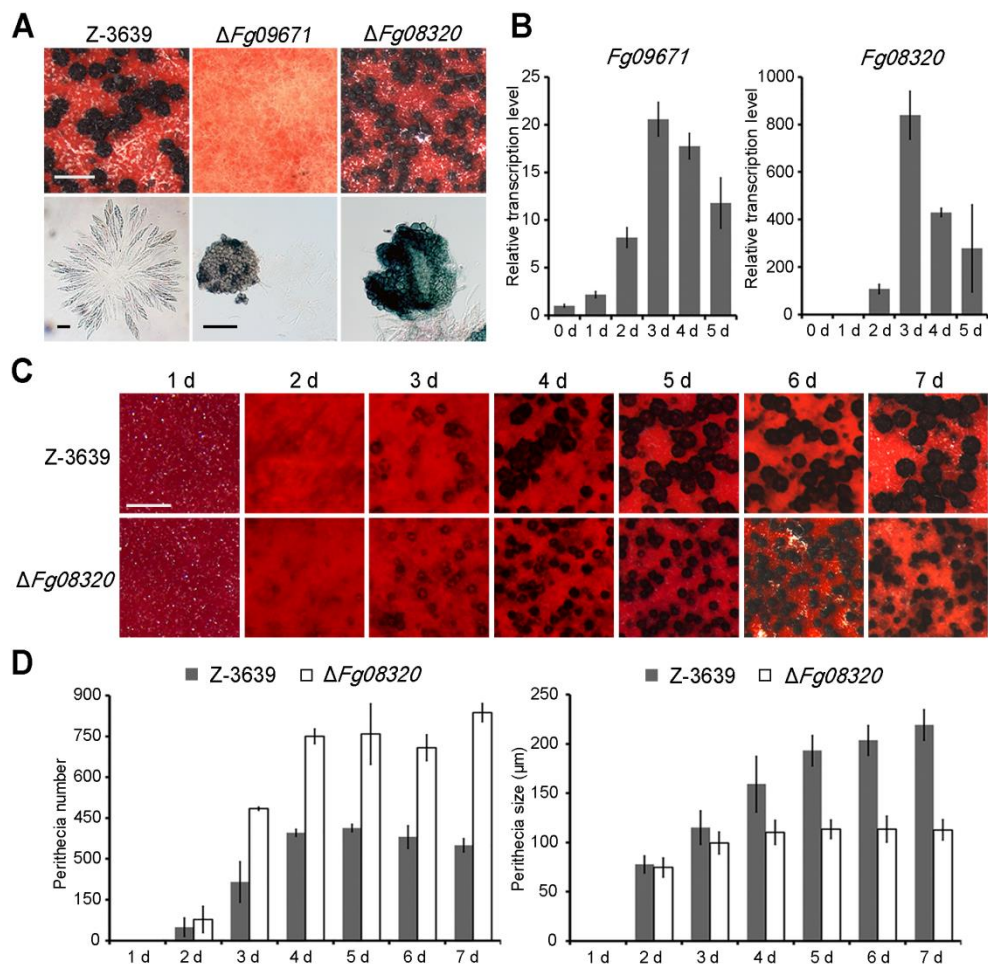
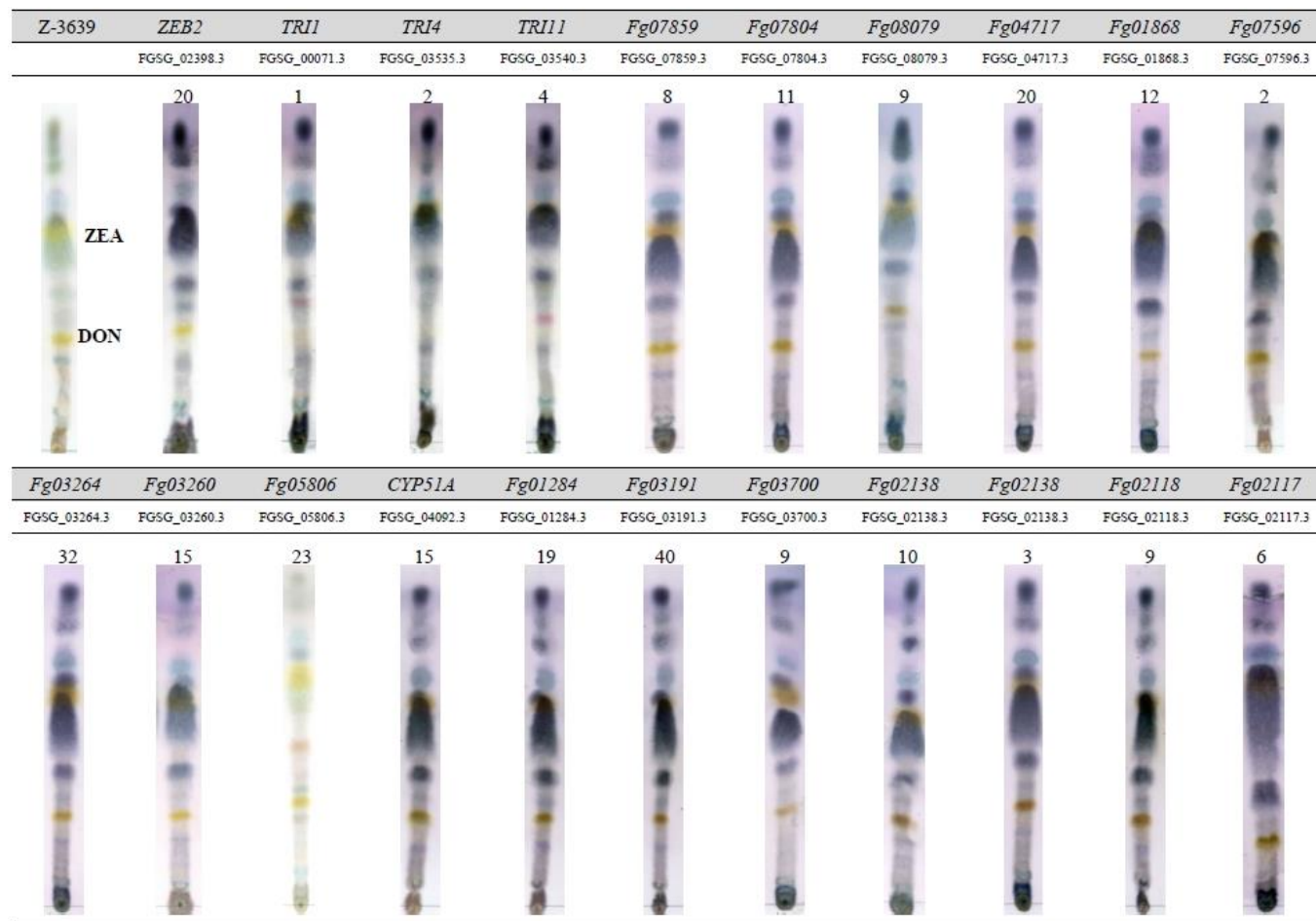
























FIG 6. Sexual development of P450 mutants. A. Perithecium formation and asci rosettes of Z-3639, $\Delta Fg09671$, and $\Delta Fg08320$. The photographs were taken 7 days after sexual induction on carrot agar. Scale bar = 200 μm (upper panel) or 20 μm (lower panel). B. Transcript profiles of *Fg08320* and *Fg09671* during perithecium development. Total RNA was prepared 0~5 days after sexual induction, and the qRT-PCR was performed to analyze the transcript levels of each gene. C. Time-course of perithecia production. The photographs were taken every 24 h after

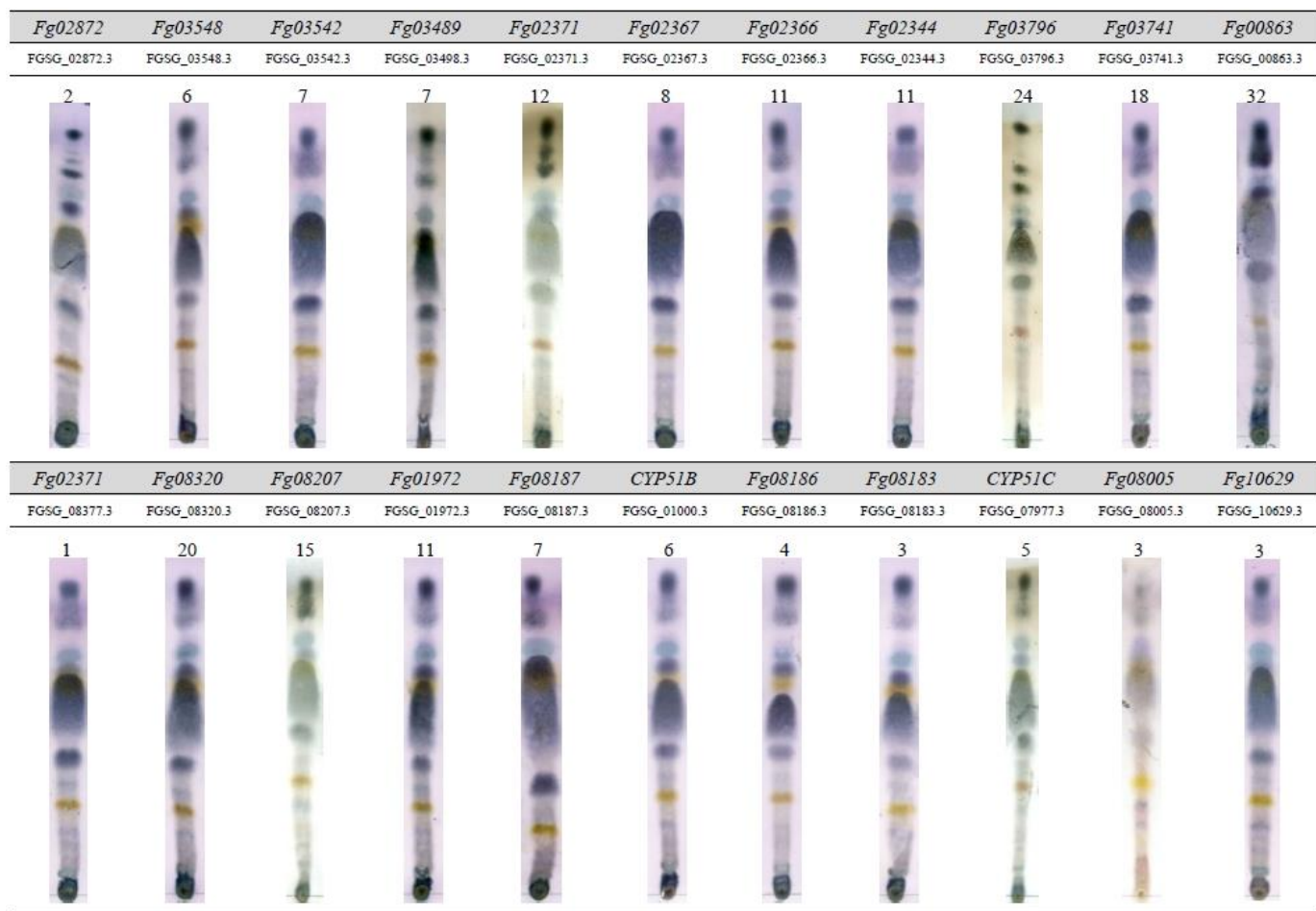
sexual induction. Scale bar = 200 μm . D. Time-course measurements of the number and size of perithecia. The diameters of the perithecia were measured for each strain using a dissecting microscope. The number of perithecia was counted from 30 mm^2 of carrot agar.

























FIG 7. Virulence of *F. graminearum* strains in wheat heads. A center spikelet of each wheat head was injected with 10 μl of a conidial suspension. The photographs were taken 21 days after inoculation. The number of infected spikelets with significant differences ($p < 0.05$, t test) was determined 21 days after inoculation. Mock inoculation was performed with 0.01% Tween 20.



<i>Fg05960</i>	<i>Fg03860</i>	<i>Fg07765</i>	<i>Fg02982</i>	<i>Fg02114</i>	<i>Fg02111</i>	<i>Fg02458</i>	<i>Fg05334</i>	<i>Fg02672</i>	<i>Fg02668</i>	<i>Fg02929</i>
FGSG_05960.3	FGSG_03860.3	FGSG_07765.3	FGSG_02982.3	FGSG_02114.3	FGSG_02111.3	FGSG_02458.3	FGSG_05334.3	FGSG_02672.3	FGSG_02668.3	FGSG_02929.3
5	12	4	1	30	10	6	2	36	1	28
										
<i>Fg06068</i>	<i>Fg01786</i>	<i>Fg01767</i>	<i>Fg01745</i>	<i>Fg01740</i>	<i>Fg01739</i>	<i>Fg01722</i>	<i>Fg05113</i>	<i>Fg01583</i>	<i>Fg01048</i>	<i>Fg02419</i>
FGSG_06068.3	FGSG_01786.3	FGSG_01767.3	FGSG_01745.3	FGSG_01740.3	FGSG_01739.3	FGSG_01722.3	FGSG_05113.3	FGSG_01583.3	FGSG_01048.3	FGSG_02419.3
5	8	14	16	6	24	9	18	11	13	31
										



<i>Fg00012</i>	<i>CLM2</i>	<i>Fg00127</i>	<i>Fg08412</i>	<i>Fg08377</i>	<i>Fg10910</i>	<i>Fg11389</i>	<i>Fg11424</i>	<i>Fg11282</i>	<i>Fg11303</i>	<i>Fg11038</i>
FGSG_00012.3	FGSG_00007.3	FGSG_00127.3	FGSG_08412.3	FGSG_10695.3	FGSG_10910.3	FGSG_11389.3	FGSG_11424.3	FGSG_11282.3	FGSG_11303.3	FGSG_11038.3
14	15	16	51	4	21	1	17	9	2	3
										
<i>Fg11024</i>	<i>Fg11465</i>	<i>Fg11498</i>	<i>Fg11536</i>	<i>Fg10451</i>	<i>Fg10461</i>	<i>Fg09351</i>	<i>Fg09195</i>	<i>Fg09916</i>	<i>Fg09671</i>	<i>Fg11655</i>
FGSG_11024.3	FGSG_11465.3	FGSG_11498.3	FGSG_11536.3	FGSG_10451.3	FGSG_10461.3	FGSG_09351.3	FGSG_09195.3	FGSG_09916.3	FGSG_09671.3	FGSG_11655.3
13	37	6	12	32	1	5	6	24	3	5
										

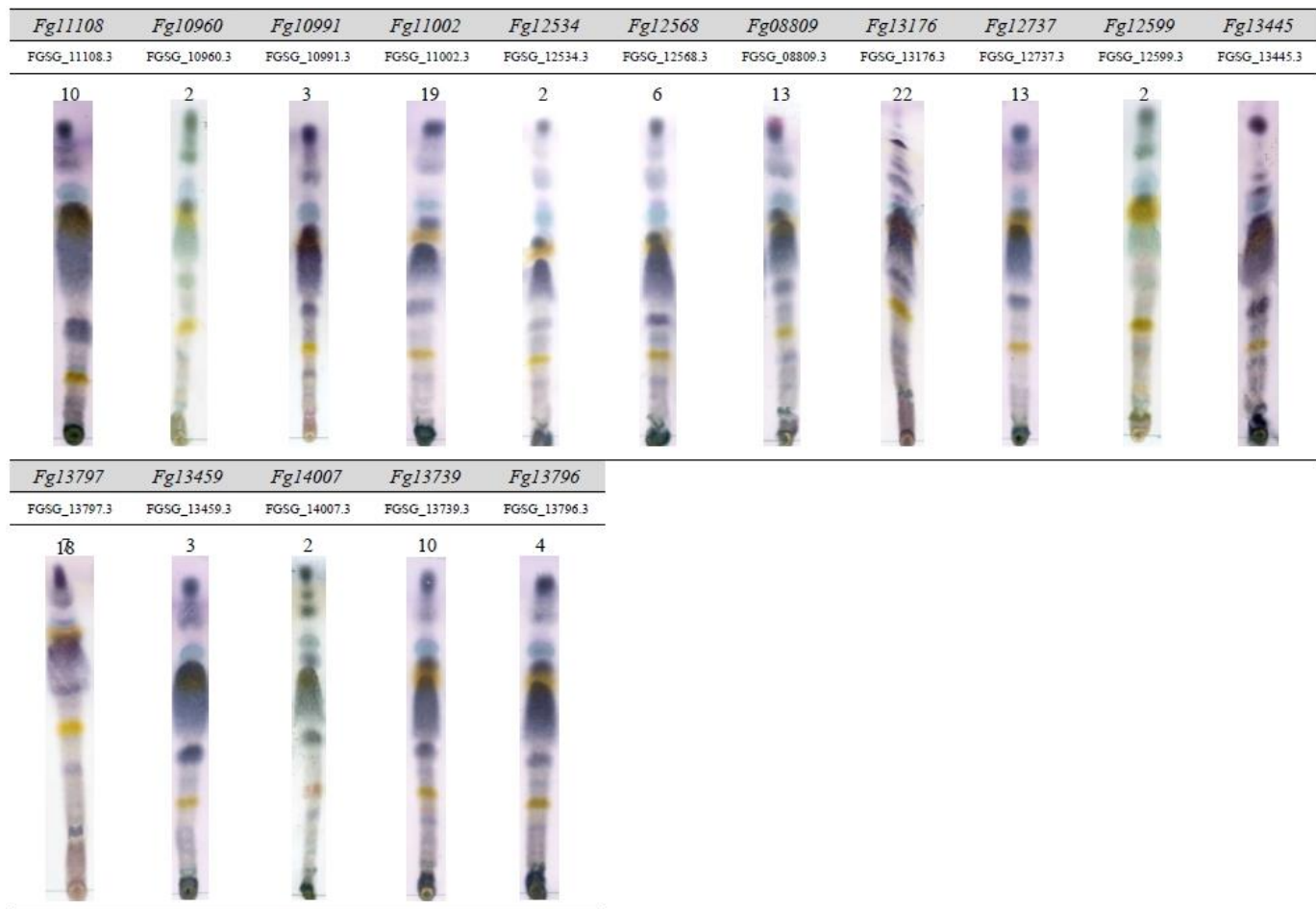


FIG 8. Mycotoxin analysis using thin-layer chromatography (TLC). Fungal strains were grown on a rice substrate for three weeks. Zearalenone (ZEA) and deoxynivalenol (DON) are indicated in a TLC plate image of the wild-type strain. The experiments were performed using two independent strains for each gene deletion with three biological replicates. *ZEB2* deleted mutant was used as a negative control for ZEA production. A representative TLC image of each gene deletion mutant is shown. WT, *F. graminearum* wild-type strain Z-3639. ZEA, zearalenone; DON, deoxynivalenol.

Table 2. Quantification of deoxynivalenol (DON) and zearalenone.

Gene name	Toxin production ^a	
	ZEA (µg/g)	DON (µg/g)
<i>Z-3639</i> ^b	1442.4 ± 159.1	477.1 ± 45.1
<i>ZEB2</i> ^c	0 ± 0	502.0 ± 13.8
<i>TRI1</i>	1455.7 ± 38.8	0 ± 0
<i>TRI4</i>	1260.5 ± 287.3	0 ± 0
<i>TRI11</i>	1299.7 ± 176.0	110.7 ± 17.8 ^d
<i>Fg02668</i>	1396.2 ± 86.6	484.5 ± 10.7
<i>Fg10960</i>	1511.5 ± 21.1	475.6 ± 8.9
<i>Z-3639</i>	1486.1 ± 383.3	463.5 ± 69.2
<i>Fg03264</i>	1035.4 ± 197.9	408.3 ± 61.5
<i>Fg03191</i>	1464.0 ± 25.4	415.2 ± 11.2
<i>Fg02118</i>	1186.9 ± 57.4	461.2 ± 8.4
<i>Fg11424</i>	1545.9 ± 80.8	433.7 ± 9.0
<i>Fg12568</i>	1125.0 ± 140.6	434.9 ± 16.0
<i>Fg07596</i>	1753.1 ± 75.8	551.0 ± 31.6
<i>CYP51A</i>	1557.5 ± 44.9	450.7 ± 7.1
<i>Fg01284</i>	1534.7 ± 59.7	566.3 ± 11.4
<i>Fg00127</i>	1699.7 ± 28.9	397.1 ± 10.5
<i>Fg10991</i>	1703.6 ± 38.9	447.6 ± 4.5
<i>Fg02982</i>	1327.0 ± 180.7	450.5 ± 13.1
<i>Fg03498</i>	1188.0 ± 87.9	419.3 ± 4.7
<i>Fg02419</i>	1247.7 ± 144.8	484.0 ± 3.9
<i>Fg10695</i>	1220.4 ± 91.1	479.3 ± 31.5
<i>Fg11465</i>	1493.4 ± 48.3	487.6 ± 13.4
<i>Fg11498</i>	1657.2 ± 68.6	510.4 ± 15.7
<i>Fg11536</i>	1422.3 ± 332.3	493.3 ± 17.7
<i>Fg10461</i>	1708.6 ± 51.7	551.1 ± 34.9
<i>Fg12534</i>	1089.8 ± 431.4	490.1 ± 15.6
<i>Fg00863</i>	1127.6 ± 397.4	433.8 ± 36.1
<i>Z-3639</i>	1686.5 ± 66.0	599.2 ± 16.0
<i>Fg02872</i>	1681.8 ± 99.3	577.0 ± 23.2

<i>Fg01767</i>	1726.6 ± 65.2	584.9 ± 7.9
<i>Fg01722</i>	1708.2 ± 34.5	604.7 ± 14.6
<i>Fg03686</i>	1588.4 ± 45.5	598.6 ± 21.2
<i>Fg02111</i>	1568.6 ± 78.0	594.8 ± 17.7
<i>Z-3639</i>	1181.9 ± 158.0	390.1 ± 17.2
<i>Fg11303</i>	1143.7 ± 160.6	368.7 ± 11.3
<i>Fg08005</i>	1397.1 ± 57.7	372.8 ± 15.1
<i>Z-3639</i>	1537.1 ± 250.1	470.9 ± 32.5
<i>Fg01048</i>	1342.1 ± 190.4	432.1 ± 19.9
<i>Fg08320</i>	1356.5 ± 90.5	481.2 ± 1.8
<i>Fg08186</i>	1379.8 ± 110.7	465.6 ± 10.2
<i>Fg05806</i>	1364.8 ± 135.6	477.8 ± 7.4
<i>Fg01786</i>	1396.5 ± 170.6	461.9 ± 43.6
<i>Fg11002</i>	1327.1 ± 120.8	474.2 ± 6.4
<i>Fg02138</i>	1323.7 ± 67.5	498.6 ± 16.5
<i>Fg13796</i>	1341.5 ± 89.1	476.9 ± 5.0
<i>Fg09671</i>	1230.9 ± 199.0	460.1 ± 8.2
<i>Fg09195</i>	1349.8 ± 87.7	398.0 ± 125.0
<i>Fg01972</i>	1589.8 ± 170.0	480.4 ± 5.0
<i>Fg08183</i>	1470.8 ± 77.1	475.5 ± 10.5
<i>CYP51B</i>	1417.9 ± 132.8	477.8 ± 24.7
<i>Fg04717</i>	1498.7 ± 47.1	479.0 ± 7.3
<i>Fg07804</i>	1537.1 ± 106.1	473.7 ± 32.7
<i>Fg03700</i>	1536.2 ± 91.7	403.4 ± 85.4
<i>Fg02366</i>	1479.9 ± 69.3	453.2 ± 14.6
<i>Fg01740</i>	1558.8 ± 41.5	484.6 ± 6.6
<i>Z-3639</i>	1561.6 ± 79.3	533.5 ± 22.3
<i>Fg02929</i>	1688.7 ± 146	562.5 ± 18.2
<i>Z-3639</i>	1664.8 ± 149.9	495.1 ± 25.1
<i>Fg03548</i>	1683.9 ± 88.9	515.0 ± 18.4
<i>Fg05334</i>	1702.6 ± 105.1	486.4 ± 13.5
<i>Fg02672</i>	1667.1 ± 109.7	535.6 ± 14.8
<i>Fg13739</i>	1679.3 ± 111.7	435.2 ± 42.5
<i>Fg05113</i>	1627.0 ± 21.8	492.8 ± 10.3

<i>Fg07859</i>	1570.2 ± 201.3	512.1 ± 21.5
<i>Fg05960</i>	1509.7 ± 275.7	497.0 ± 69.2
<i>Fg03860</i>	1558.6 ± 25.2	511.8 ± 19.7
<i>Z-3639</i>	1360.9 ± 265.3	373.9 ± 43.6
<i>Fg08079</i>	1351.5 ± 51.8	408.2 ± 48.0
<i>Fg02114</i>	1024.8 ± 299.2	330.2 ± 6.4
<i>Fg01745</i>	1019.5 ± 176.7	316.3 ± 14.7
<i>Fg01739</i>	1697.1 ± 205.0	354.7 ± 29.9
<i>Z-3639</i>	1466.8 ± 112.0	476.5 ± 32.3
<i>Fg09916</i>	1126.9 ± 259.9	451.5 ± 45.0
<i>Fg08187</i>	1566.4 ± 36.4	480.4 ± 8.8
<i>Fg02458</i>	1590.9 ± 53.4	485.4 ± 2.1
<i>Fg11024</i>	1249.5 ± 150.3	484.2 ± 0.2
<i>Fg02117</i>	1640.6 ± 53.9	470.9 ± 18.9
<i>Fg11282</i>	1694.0 ± 58.6	481.8 ± 37.2
<i>Fg10629</i>	1385.7 ± 181.0	479.5 ± 7.5
<i>Fg01583</i>	1619.2 ± 81.4	489.4 ± 23.9
<i>Fg13459</i>	1461.9 ± 230.0	480.6 ± 16.5
<i>CLM2</i>	1637.5 ± 33.5	477.2 ± 14.2
<i>Fg00012</i>	1487.0 ± 16.4	489.1 ± 12.9
<i>Fg02344</i>	1487.9 ± 39.9	481.0 ± 7.9
<i>Fg06068</i>	1347.4 ± 108.5	465.5 ± 30.5
<i>Fg03542</i>	1304.9 ± 166.7	469.3 ± 17.5
<i>Fg03260</i>	1574.9 ± 49.6	499.8 ± 18.6
<i>Fg12737</i>	1716.5 ± 100.4	416.7 ± 52.8
<i>Fg07765</i>	1727.1 ± 69.9	497.1 ± 11.9
<i>Fg11108</i>	1708.5 ± 103.3	438.0 ± 74.7
<i>Fg02367</i>	1348.5 ± 253.0	404.2 ± 71.7
<i>Z-3639</i>	1602.6 ± 83.1	481 ± 38.3
<i>Fg11655</i>	1531.0 ± 159.6	471.1 ± 9.8
<i>Fg13445</i>	1613.0 ± 21.8	470.9 ± 2.6
<i>Fg11038</i>	1624.6 ± 65.9	486.1 ± 28.0
<i>Fg12599</i>	1622.1 ± 52.9	472.4 ± 9.1
<i>Fg13176</i>	1545.3 ± 53.9	481.0 ± 12.6

<i>Fg08809</i>	1612.5 ± 23.0	485.0 ± 13.3
<i>Fg08377</i>	1638.4 ± 117.1	502.5 ± 34.9
<i>Fg11389</i>	1678.4 ± 118.7	454.5 ± 19.6
<i>Fg03741</i>	1626.9 ± 147.0	486.7 ± 60.2
<i>Fg09351</i>	1585.2 ± 17.3	488.6 ± 27.3
<i>Fg10451</i>	1527.9 ± 55.7	461.4 ± 16.9
<i>Fg13797</i>	1449.5 ± 149.8	441.5 ± 19.7
<i>Z-3639</i>	1449.6 ± 202.2	468.3 ± 21.4
<i>Fg02371</i>	1280.9 ± 93.7	479.9 ± 37.0
<i>Fg08412</i>	1089.8 ± 164.9	420.4 ± 99.0
<i>Fg08207</i>	1769.8 ± 191.4	452.2 ± 17.8
<i>Fg14007</i>	1573.0 ± 93.3	458.2 ± 89.9
<i>CYP51C</i>	1522.3 ± 73.3	490.7 ± 17.7
<i>Fg10910</i>	1488.7 ± 128.1	466.0 ± 84.5
<i>Fg01868</i>	1640.7 ± 95.3	487.2 ± 61.2
<i>Fg03796</i>	1382.4 ± 79.7	489.8 ± 18.3

^a Toxin production (ZEA, zearalenone; DON, deoxynivalenol) was quantified by Neogen Veratox kit and the experiments were performed using two independent strains with three biological repetitions/replicates for each gene deletion.

^b Mycotoxin production of each P450 mutant was not significantly different ($p < 0.05$, t test) compared to the wild type of corresponding experimental batch.

^c ZEB2 deletion mutant is used for the negative control strains for ZEA production.

^d Although ELISA signal for DON was detected in *tri11* mutants possibly due to accumulated DON intermediates, further GC-MS analysis confirmed that *tri11* deletion mutants did not produce DON.

V. Stress test

I performed in-depth phenotypic analyses under various environmental stresses. The tested conditions included osmotic (or ionic), reactive oxygen species (ROS), and cell wall stress; acidic (pH = 4) and basic (pH = 11) conditions; hot and cold conditions; and several fungicide treatments (Supporting information Table S1). None of the P450 mutants showed altered phenotypes under the evaluated stress conditions, with the exception of the *ΔFg05960* mutant strain, which produced red pigments 1 day earlier than the wild type in the cold stress condition (data not shown).

As secondary metabolite biosynthetic P450 enzymes are related to UV stress responses in higher eukaryotes, such as humans and plants (Katiyar et al., 2000; Winkel-Shirley, 2001), I additionally tested P450 mutants under UV stress. The *Fg01583* gene deletion mutant grew faster than the wild-type strain under the UV-B stress condition and accumulated more orange pigment in its mycelia (Fig. 7A). Moreover, *Fg01583* was upregulated in response to UV light in the wild-type strain (Fig. 7B). The results suggest that the *Fg01583* gene plays a role in the UV stress response, and pigment accumulation is possibly involved in this process.

VI. Sensitivity to azole fungicides

Azole antifungal agents are well-known fungicides that inhibit fungal sterol 14 α -demethylase (Cyp51) (Liu et al., 2011). To better understand the azole drug

resistance mechanism in *F. graminearum*, I screened P450 mutants on CM supplemented with five azole fungicides (tebuconazole, difenoconazole, epoxiconazole, propiconazole, and prochloraz). Experimental conditions were established in which the wild-type strain showed significant growth reduction compared with the control. In total, seventeen P450 mutants exhibited defective mycelial growth in at least one azole fungicide-complemented condition compared with the wild type (Supporting information Table S3). In particular, the $\Delta Fg02114$, $\Delta Fg02668$, $\Delta Fg05113$, and $\Delta Fg01048$ strains displayed markedly increased sensitivity to several azole fungicides, including tebuconazole, difenoconazole, and prochloraz (Fig. 8). Three *CYP51* genes (*CYP51A*, *CYP51B*, and *CYP51C*) have been reported as *CYP51* paralogs, and only the $\Delta cyp51A$ strains showed increased sensitivity to azole drugs, as previously reported (Liu et al., 2011; Fan et al., 2013) (Fig. 8A and B).

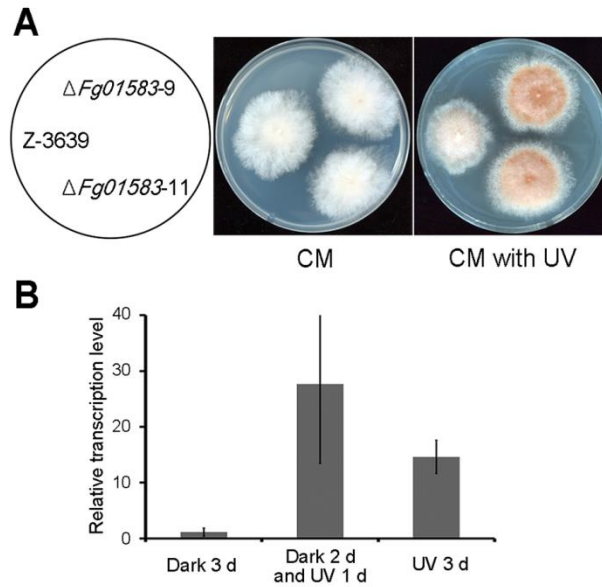


FIG 9. Altered UV stress response of *Fg01583* mutants. A. Mycelial growth in CM under UV light. The photographs were taken after 3 days of incubation under continuous UV-B light. B. Relative transcript levels of *Fg01583* in response to UV stress. Total RNA was extracted from the wild-type strain after 3 days of incubation in the dark (Dark 3 d); after 1 day of incubation under UV light after 2 days in the dark (Dark 2 d and UV 1 d); and after 3 days of incubation under UV light (UV 3 d) in CM.

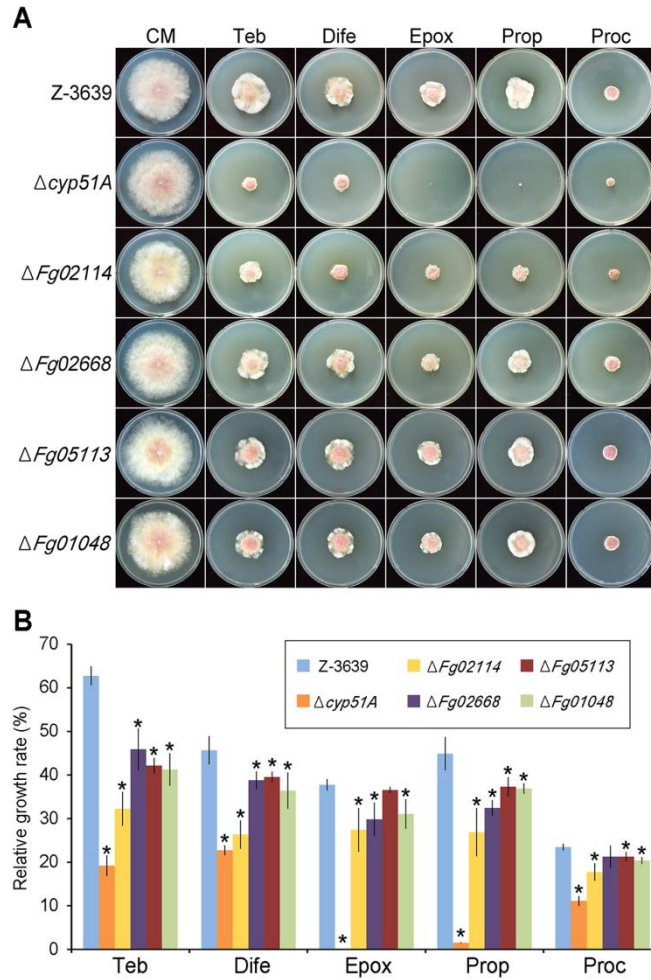


FIG 10. Azole fungicide sensitivity of *F. graminearum* strains. A. Mycelial growth of *F. graminearum* strains on CM with different azole fungicides. The photographs were taken 5 days after inoculation. B. Relative growth rate of *F. graminearum* strains on azole fungicide media compared with the control. Values with significant differences ($p < 0.05$, t test) compared with wild type are indicated by asterisks. Ten, tebuconazole; Dife, difenoconazole; Epox, epoxiconazole; Prop, propiconazole; Proc, prochloraz.

VII. Xenobiotic induction

Detoxification of xenobiotics (toxins, carcinogens, and pollutants) is one of the well-known functions of P450 enzymes (Werck-Reichhart and Feyereisen, 2000). To investigate the involvement of P450s in xenobiotic degradation, I screened my P450 mutant library with various xenobiotic compounds. I selected 15 xenobiotic compounds, including three aliphatics, four aromatics, five polyaromatics, two alkyl-substituted aromatics, and one alicyclic, for my analysis (Supporting information Table S4). Xenobiotic conditions were established in which the wild-type strain showed significant growth reduction compared with the control. Intriguingly, only one P450 gene deletion mutant was highly sensitive to CM containing 1-dodecanol compared with the wild type (Fig. 9A), and the *Fg01972* gene was markedly upregulated in response to 1-dodecanol in the wild-type strain (Fig. 9B).

Assuming that most P450s exhibit redundant functions in xenobiotic degradation, I compared the transcript abundance of P450 genes under 15 different xenobiotic stress conditions using quantitative real-time (qRT) PCR analysis (Supporting information Table S5 and Fig. 10). The expression levels of most P450 genes were significantly upregulated in at least one xenobiotic condition compared with the untreated control, and each xenobiotic condition induced many P450 genes at the same time (Fig. 10). While 1-dodecanol (DD), 4-octylphenol (OP), naphthalene (NPH) and 1-naphthol (NAP) induced most of the P450 genes, the majority of P450s were negatively regulated in response to treatments with

phenoxyacetic acid (PAA), resorcinol (RES), 3-methyl cholanthrene (MA), benzo(a)pyrene (BAP), sodium dodecylbenzenesulfonate (DBS), and n-dodecane (D). P450s included in the CYP505 (*Fg07596* and *Fg01972*) and CYP540 (*Fg02138* and *Fg03548*) families, which have been shown to be required for fatty acid hydroxylation (Nakayama et al., 1996), were highly induced in CM containing aliphatics (Supporting information Table S5 and Fig. 10).

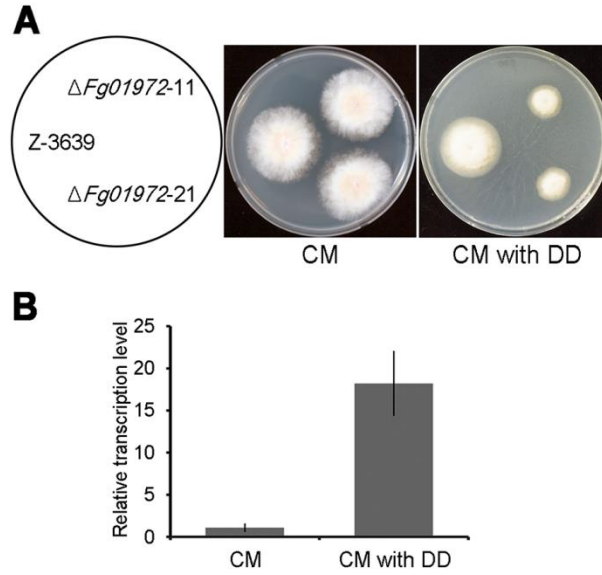


FIG 11. Altered xenobiotic stress response of the *Fg01972* mutants. A. Mycelial growth in $\Delta Fg01972$ on CM and CM supplemented with 1-dodecanol. The photographs were taken 3 days after inoculation. B. Relative transcript levels of the *Fg01972* gene in response to 1-dodecanol. Total RNA was extracted from the wild type 24 h after inoculation into CM with 1-dodecanol. DD, 1-dodecanol.

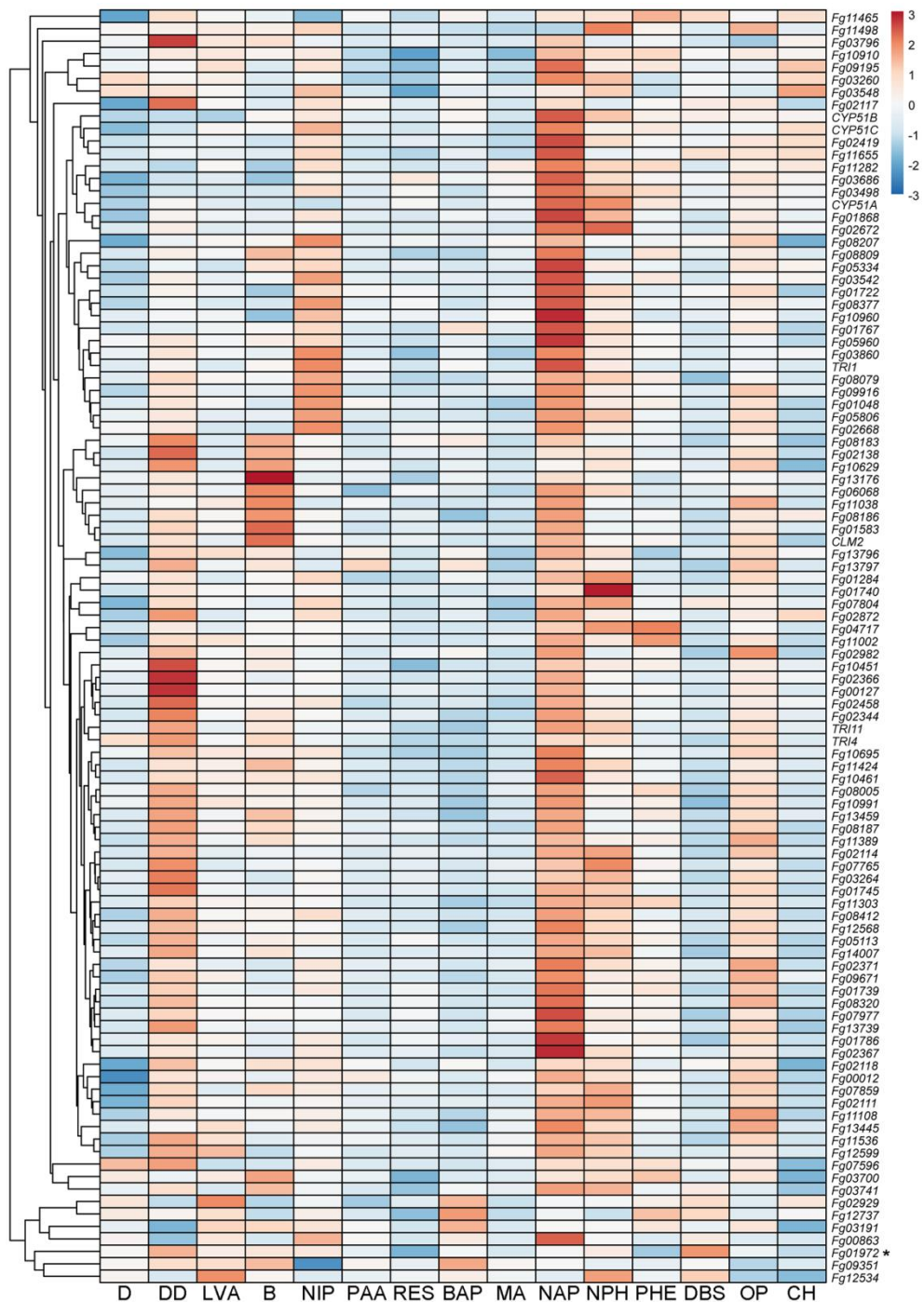


FIG 12. Heatmap visualization of P450 gene transcript levels under various xenobiotic conditions. The xenobiotic induction of P450 transcript levels was measured via qRT-PCR and visualized using ClustVis (Metsalu and Vilo, 2015) based on the Log2-based relative transcript abundance compared with the control. B, benzoate; DD, 1-dodecanol; OP, 4-octylphenol; NPH, naphthalene; PAA, phenoxy acetic acid; RES, resorcinol; MA, 3-methyl cholanthrene; BAP, benzo(a)pyrene; DBS, dodecyl benzene sulfonate; D, n-dodecane; LVA, levulinic acid; PHE, phenanthrene; CH, cyclohexanone; NIP, nitrophenol; NAP, 1-naphthol.

Table 3. Phenotypes of P450 mutants under various azole fungicide conditions.

Gene name	Azole fungicides				
	Tebuconazole	Difenoconazole	Epoxiconazole	Propiconazole	Prochloraz
<i>Fg08079</i>	3 ^a	4 ^b	4 ^c	4 ^d	4 ^e
<i>Fg05960</i>	4, cc ^f	4, cc	4, cc	4, cc	4, cc
<i>CYP51A</i>	1	2	0	1	2
<i>Fg02114</i>	2	2	3	2	3
<i>Fg02672</i>	4	4	4	4	3
<i>Fg02668</i>	3	3	3	3	4
<i>Fg02929</i>	3	4	4	3	3
<i>Fg03498</i>	3	4	4	4	4
<i>Fg01767</i>	3	4	4	4	4
<i>Fg01745</i>	3	4	3	4	4
<i>Fg01740</i>	3	4	4	3	4
<i>Fg01739</i>	3	4	4	4	4
<i>Fg01722</i>	3	4	4	4	4
<i>Fg05113</i>	3	3	4	3	4
<i>Fg01583</i>	3	4	4	4	4
<i>Fg01048</i>	3	3	3	3	4
<i>Fg02419</i>	3	4	4	4	4

^a Percentage of average radial growth on CM with tebuconazole of deletion mutants compared to Z-3639 was scored (5, more than Z-3639; 4, 91-100%; 3, 61-90%; 2, 31-60 %; 1, 0-30 %; 0, 0 %).

^b Percentage of average radial growth on CM with difenoconazole of deletion mutants compared to Z-3639 was scored (5, more than Z-3639; 4, 91-100%; 3, 61-90%; 2, 31-60 %; 1, 0-30 %; 0, 0 %).

^c Percentage of average radial growth on CM with epoxiconazole of deletion mutants compared to Z-3639 was scored (5, more than Z-3639; 4, 91-100%; 3, 61-90%; 2, 31-60 %; 1, 0-30 %; 0, 0 %).

^d Percentage of average radial growth on CM with propiconazole of deletion mutants compared to Z-3639 was scored (5, more than Z-3639; 4, 91-100%; 3, 61-90%; 2, 31-60 %; 1, 0-30 %; 0, 0 %).

^e Percentage of average radial growth on CM with prochloraz of deletion mutants compared to Z-3639 was scored (5, more than Z-3639; 4, 91-100%; 3, 61-90%; 2, 31-60 %; 1, 0-30 %; 0, 0 %).

^f Color change of pigment (cc)

Table 4. Transcription of P450 genes under xenobiotic induction.

Gene name	Locus number	Z-3639 DW	Z-3639 DMSO	Aliphatics			Aromatics					Polyaromatics				Alkyl-substituted aromatics		Alicyclics
				n-Dodecane	1-Dodecanol	Levulinic acid	Benzoate	Nitrophenol	Phenoxy acetic acid	Resorcinol	Benzo(a)pyrene	3-Methyl cholanthrene	1-Naphthol	Naphthalene	Phenanthrene	Dodecyl benzene sulfonate	Octylphenol	Cyclohexanone
<i>Fg07859</i>	FGSG_07859.3	1.11 ± 0.27	1.22 ± 0.37	0.26 ± 0.15	14.98 ± 0.95	1.69 ± 0.58	14.08 ± 5.32	7.09 ± 0.57	2.14 ± 0.15	2.25 ± 0.58	1.23 ± 0.23	1.51 ± 0.95	16.92 ± 0.58	31.95 ± 5.32	3.60 ± 0.95	1.22 ± 0.15	17.24 ± 0.23	0.79 ± 0.23
<i>Fg07804</i>	FGSG_07804.3	1.03 ± 0.28	1.28 ± 0.38	0.28 ± 0.36	4.16 ± 0.36	1.53 ± 1.40	1.13 ± 0.02	4.63 ± 2.85	0.88 ± 0.24	0.56 ± 0.40	1.06 ± 0.81	0.39 ± 0.17	7.58 ± 0.47	9.37 ± 3.84	1.42 ± 0.71	2.69 ± 0.79	2.41 ± 0.53	1.10 ± 0.17
<i>Fg08079</i>	FGSG_08079.3	1.28 ± 0.46	1.05 ± 0.25	1.3 ± 1.65	10.3 ± 2.99	3.2 ± 1.18	2.4 ± 0.10	23.5 ± 8.78	1.5 ± 0.53	0.5 ± 0.35	0.6 ± 0.20	1.57 ± 0.48	21.40 ± 7.49	12.40 ± 5.45	4.56 ± 2.91	0.34 ± 0.10	2.88 ± 1.38	0.82 ± 0.41
<i>Fg04717</i>	FGSG_04717.3	1.07 ± 0.46	1.05 ± 0.34	1.42 ± 0.50	18.51 ± 5.04	3.23 ± 0.98	9.52 ± 3.01	8.94 ± 3.28	5.58 ± 1.98	1.41 ± 0.78	1.61 ± 1.16	2.04 ± 0.31	68.02 ± 6.06	230.74 ± 84.42	355.92 ± 245.75	1.47 ± 0.45	8.88 ± 2.81	1.46 ± 1.30
<i>Fg01868</i>	FGSG_01868.3	1.02 ± 0.20	1.24 ± 0.45	0.54 ± 0.06	2.28 ± 0.59	1.58 ± 0.49	1.38 ± 0.40	3.33 ± 1.04	1.16 ± 0.57	1.00 ± 0.62	1.71 ± 0.66	1.23 ± 0.52	18.23 ± 0.47	6.85 ± 2.12	1.60 ± 0.61	0.85 ± 0.28	2.68 ± 1.02	1.43 ± 0.55
<i>Fg07596</i>	FGSG_07596.3	1.19 ± 0.41	1.27 ± 0.65	122.44 ± 3.76	255.92 ± 170.78	0.76 ± 0.30	1.08 ± 0.81	15.45 ± 1.99	1.59 ± 0.54	1.48 ± 0.70	0.99 ± 0.11	2.10 ± 1.27	43.72 ± 27.53	34.86 ± 0.93	35.96 ± 24.19	5.20 ± 3.31	15.53 ± 3.63	0.20 ± 0.06
<i>Fg05960</i>	FGSG_05960.3	1.00 ± 0.06	1.03 ± 0.28	1.04 ± 0.12	1.76 ± 1.55	1.00 ± 0.02	1.46 ± 0.16	2.40 ± 0.08	0.61 ± 0.04	0.45 ± 0.18	0.78 ± 0.52	0.46 ± 0.16	9.94 ± 0.40	1.78 ± 0.57	1.14 ± 0.06	0.58 ± 0.07	0.89 ± 0.31	0.42 ± 0.44
<i>Fg03860</i>	FGSG_03860.3	1.00 ± 0.06	1.03 ± 0.25	0.87 ± 0.39	1.83 ± 1.75	1.03 ± 0.05	0.81 ± 0.63	4.94 ± 0.14	0.66 ± 0.09	0.34 ± 0.19	0.92 ± 0.34	0.39 ± 0.11	5.16 ± 0.20	1.66 ± 1.09	1.21 ± 0.13	0.64 ± 0.04	0.96 ± 0.32	1.05 ± 0.04
<i>Fg07765</i>	FGSG_07765.3	1.12 ± 0.15	1.10 ± 0.25	0.69 ± 0.48	115.15 ± 72.10	0.95 ± 0.29	1.80 ± 0.70	2.48 ± 0.85	0.99 ± 0.24	1.23 ± 0.58	0.65 ± 0.23	1.45 ± 0.47	29.96 ± 7.06	117.08 ± 20.44	4.17 ± 2.58	0.53 ± 0.32	6.97 ± 0.28	0.36 ± 0.12
<i>Fg02982</i>	FGSG_02982.3	1.02 ± 0.21	1.03 ± 0.26	0.86 ± 0.12	3.83 ± 0.65	1.36 ± 0.40	1.91 ± 0.68	1.21 ± 0.29	0.80 ± 0.23	1.16 ± 0.44	1.48 ± 1.26	0.62 ± 0.31	5.20 ± 1.26	1.01 ± 0.31	1.55 ± 0.42	0.50 ± 0.15	5.85 ± 2.05	0.53 ± 0.09
<i>Fg03264</i>	FGSG_03264.3	1.04 ± 0.28	1.10 ± 0.41	1.26 ± 0.74	40.16 ± 30.27	1.81 ± 0.47	0.88 ± 0.40	2.76 ± 0.46	1.04 ± 0.24	1.45 ± 1.06	0.70 ± 0.19	1.28 ± 0.39	11.81 ± 3.27	17.18 ± 15.29	2.83 ± 1.08	0.55 ± 0.35	9.38 ± 6.25	0.67 ± 0.32
<i>Fg03260</i>	FGSG_03260.3	1.04 ± 0.30	1.04 ± 0.30	5.47 ± 1.04	2.45 ± 0.18	2.87 ± 0.19	1.18 ± 1.03	1.85 ± 0.63	0.61 ± 0.14	0.57 ± 0.17	2.01 ± 0.79	0.67 ± 0.03	15.53 ± 4.75	7.68 ± 2.81	0.82 ± 0.25	2.05 ± 0.57	1.69 ± 1.09	5.70 ± 0.57
<i>Fg05806</i>	FGSG_05806.3	1.05 ± 0.35	1.13 ± 0.49	2.39 ± 0.35	7.38 ± 1.60	1.79 ± 0.43	2.04 ± 1.61	42.57 ± 0.99	1.44 ± 0.24	1.20 ± 0.72	1.21 ± 0.5	0.77 ± 0.11	42.57 ± 6.43	21.35 ± 11.68	2.81 ± 0.14	1.54 ± 0.47	13.91 ± 9.86	0.70 ± 0.10
<i>CYP51A</i>	FGSG_04092.3	2.07 ± 1.99	1.05 ± 0.39	0.32 ± 0.11	4.46 ± 3.69	1.29 ± 0.79	0.80 ± 0.61	0.48 ± 0.07	1.21 ± 0.31	3.91 ± 4.14	0.85 ± 0.88	2.07 ± 0.39	243.88 ± 30.13	127.33 ± 20.91	10.39 ± 4.26	2.54 ± 0.84	5.58 ± 2.16	3.21 ± 1.33
<i>Fg01284</i>	FGSG_01284.3	1.03 ± 0.26	1.03 ± 0.22	1.86 ± 0.56	3.23 ± 1.12	1.22 ± 0.05	1.97 ± 0.60	3.97 ± 1.35	0.81 ± 0.20	0.85 ± 0.34	1.77 ± 0.74	0.98 ± 0.37	5.15 ± 0.34	7.29 ± 2.57	1.14 ± 0.04	0.87 ± 0.59	3.98 ± 1.79	1.59 ± 0.53
<i>Fg03191</i>	FGSG_03191.3	1.06 ± 0.38	1.01 ± 0.16	0.73 ± 0.03	0.38 ± 0.27	1.44 ± 0.06	1.43 ± 0.11	1.23 ± 0.05	0.90 ± 0.35	0.57 ± 0.07	1.80 ± 1.34	0.62 ± 0.01	0.91 ± 0.33	0.91 ± 0.31	1.16 ± 0.04	0.76 ± 0.48	1.20 ± 0.05	0.38 ± 0.03
<i>Fg03700</i>	FGSG_03700.3	1.24 ± 0.61	1.13 ± 0.43	4.28 ± 1.69	1.55 ± 0.16	3.85 ± 0.11	21.48 ± 6.76	1.91 ± 0.70	1.44 ± 0.86	0.20 ± 0.14	0.87 ± 0.63	0.98 ± 0.38	5.16 ± 0.39	7.52 ± 2.45	12.03 ± 8.05	0.95 ± 0.34	5.22 ± 0.31	0.19 ± 0.11
<i>Fg03686</i>	FGSG_03686.3	1.25 ± 0.83	1.15 ± 0.63	0.23 ± 0.03	0.49 ± 0.42	0.74 ± 0.28	0.30 ± 0.20	1.67 ± 0.07	0.86 ± 0.52	2.18 ± 0.95	1.06 ± 0.74	1.50 ± 0.11	12.93 ± 0.15	3.11 ± 0.19	1.47 ± 0.04	1.19 ± 0.38	2.39 ± 0.82	1.49 ± 0.54

<i>Fg02138</i>	FGSG_02138.3	1.03 ± 0.26	1.05 ± 0.33	1.23 ± 0.10	119.59 ± 12.18	1.76 ± 0.53	38.06 ± 4.36	5.70 ± 0.55	1.48 ± 0.27	2.61 ± 2.23	2.79 ± 2.53	1.25 ± 0.03	11.75 ± 2.08	11.24 ± 1.31	2.37 ± 0.20	1.89 ± 0.57	15.30 ± 5.66	0.92 ± 0.27
<i>Fg02118</i>	FGSG_02118.3	1.01 ± 0.15	1.05 ± 0.32	0.47 ± 0.17	2.98 ± 0.59	1.44 ± 0.46	2.17 ± 1.99	2.10 ± 0.28	1.41 ± 0.31	0.82 ± 0.44	1.45 ± 0.94	0.90 ± 0.23	2.53 ± 0.14	2.28 ± 0.54	1.09 ± 0.11	1.45 ± 0.48	2.31 ± 0.34	0.51 ± 0.08
<i>Fg02117</i>	FGSG_02117.3	1.24 ± 0.60	1.02 ± 0.23	0.26 ± 0.07	10.23 ± 7.85	1.50 ± 0.54	0.83 ± 0.62	2.85 ± 0.43	1.67 ± 0.15	0.82 ± 0.63	1.78 ± 1.70	0.66 ± 0.21	3.13 ± 3.37	0.86 ± 0.03	1.50 ± 0.20	2.06 ± 1.29	2.36 ± 1.06	0.52 ± 0.03
<i>Fg02114</i>	FGSG_02114.3	1.01 ± 0.17	1.05 ± 0.36	0.69 ± 0.51	15.87 ± 3.47	1.09 ± 0.10	1.43 ± 0.63	1.38 ± 0.57	0.76 ± 0.21	1.15 ± 0.73	1.04 ± 1.26	0.62 ± 0.43	17.13 ± 3.78	20.60 ± 18.56	1.70 ± 0.73	0.40 ± 0.13	11.67 ± 8.43	0.66 ± 0.14
<i>Fg02111</i>	FGSG_02111.3	1.04 ± 0.31	1.04 ± 0.29	0.48 ± 0.13	6.69 ± 2.15	2.56 ± 0.48	2.01 ± 0.28	2.72 ± 0.28	1.67 ± 0.59	1.55 ± 0.99	1.57 ± 1.00	1.44 ± 0.14	10.22 ± 3.46	12.86 ± 8.44	2.29 ± 0.68	1.01 ± 0.26	6.24 ± 1.01	0.91 ± 0.30
<i>Fg02458</i>	FGSG_02458.3	1.04 ± 0.31	1.04 ± 0.33	1.24 ± 0.51	14.36 ± 6.00	2.48 ± 0.59	2.74 ± 0.61	3.71 ± 0.21	0.95 ± 0.20	1.14 ± 0.75	1.28 ± 0.88	0.89 ± 0.03	8.05 ± 0.59	3.38 ± 0.63	1.78 ± 0.26	1.49 ± 0.42	3.29 ± 0.20	2.05 ± 0.64
<i>Fg05334</i>	FGSG_05334.3	1.28 ± 0.68	1.03 ± 0.24	0.40 ± 0.21	0.96 ± 0.09	0.50 ± 0.04	2.13 ± 1.70	2.78 ± 0.10	0.65 ± 0.06	0.84 ± 0.58	0.57 ± 0.28	0.70 ± 0.06	10.81 ± 0.77	1.65 ± 1.36	0.94 ± 0.35	0.52 ± 0.13	2.00 ± 0.76	1.38 ± 0.41
<i>Fg02672</i>	FGSG_02672.3	1.03 ± 0.26	1.07 ± 0.34	1.40 ± 0.61	18.56 ± 2.75	2.04 ± 0.84	2.24 ± 0.84	6.50 ± 2.98	1.56 ± 0.46	1.46 ± 0.93	1.90 ± 1.37	1.38 ± 0.40	13699.26 ± 5314.40	1757.45 ± 720.12	4.12 ± 0.70	1.17 ± 0.71	22.50 ± 15.26	6.41 ± 4.26
<i>Fg02668</i>	FGSG_02668.3	1.08 ± 0.37	1.04 ± 0.29	1.43 ± 0.31	2.52 ± 0.36	0.70 ± 0.16	0.59 ± 0.19	9.19 ± 0.26	0.59 ± 0.05	1.10 ± 0.19	1.21 ± 0.74	0.89 ± 0.53	8.97 ± 0.66	2.63 ± 2.13	1.24 ± 0.13	0.57 ± 0.13	3.58 ± 2.02	0.54 ± 0.16
<i>Fg02929</i>	FGSG_02929.3	1.00 ± 0.02	1.16 ± 0.63	1.48 ± 0.58	0.43 ± 0.29	3.86 ± 0.06	0.41 ± 0.16	0.88 ± 0.03	0.36 ± 0.17	0.64 ± 0.18	2.58 ± 0.96	0.46 ± 0.05	0.91 ± 0.04	0.90 ± 0.04	1.15 ± 0.38	1.74 ± 0.18	0.65 ± 0.22	1.44 ± 0.50
<i>Fg02872</i>	FGSG_02872.3	1.06 ± 0.38	1.02 ± 0.24	0.48 ± 0.16	8.86 ± 1.54	0.94 ± 0.31	1.34 ± 1.10	3.47 ± 0.18	0.82 ± 0.12	0.69 ± 0.36	1.07 ± 0.63	0.49 ± 0.04	7.50 ± 0.62	2.39 ± 1.58	1.43 ± 0.59	0.89 ± 0.10	2.43 ± 0.88	4.34 ± 1.59
<i>Fg03548</i>	FGSG_03548.3	1.01 ± 0.11	1.05 ± 0.33	14.04 ± 1.35	12.46 ± 3.23	5.49 ± 1.93	2.01 ± 0.20	26.83 ± 17.23	1.70 ± 1.11	0.42 ± 0.28	2.70 ± 0.13	2.90 ± 2.45	8.28 ± 2.85	21.72 ± 0.89	1.27 ± 0.20	4.23 ± 1.55	2.07 ± 0.89	42.59 ± 14.62
<i>Fg03542</i>	FGSG_03542.3	1.06 ± 0.37	1.00 ± 0.10	0.26 ± 0.11	2.87 ± 1.97	1.08 ± 0.42	1.61 ± 1.47	21.76 ± 9.16	0.97 ± 0.07	0.60 ± 0.41	0.91 ± 0.79	0.60 ± 0.19	63.25 ± 3.01	1.03 ± 0.55	3.93 ± 0.30	0.50 ± 0.04	1.88 ± 0.20	2.18 ± 0.74
<i>TRI11</i>	FGSG_03540.3	1.06 ± 0.39	1.01 ± 0.13	1.89 ± 0.44	22.36 ± 13.00	2.11 ± 0.74	6.442 ± 2.49	2.57 ± 0.80	2.49 ± 1.26	0.84 ± 0.15	0.66 ± 0.23	1.20 ± 0.40	24.80 ± 8.98	12.86 ± 9.39	1.53 ± 0.54	1.64 ± 0.34	8.95 ± 5.22	1.94 ± 0.65
<i>TRI4</i>	FGSG_03535.3	1.05 ± 0.35	1.02 ± 0.23	8.68 ± 3.80	37.62 ± 23.15	2.24 ± 0.78	11.84 ± 6.27	1.90 ± 0.55	0.91 ± 0.51	0.41 ± 0.02	0.47 ± 0.09	0.63 ± 0.26	10.17 ± 3.12	8.76 ± 6.54	1.40 ± 0.45	0.46 ± 0.25	10.19 ± 7.90	0.93 ± 0.31
<i>Fg03498</i>	FGSG_03498.3	1.26 ± 0.85	1.15 ± 0.62	0.29 ± 0.19	0.85 ± 0.87	0.90 ± 0.10	0.92 ± 0.96	14.99 ± 10.45	1.22 ± 0.86	3.91 ± 2.31	0.56 ± 0.39	2.63 ± 0.91	117.57 ± 3.59	29.92 ± 0.59	16.53 ± 9.28	1.86 ± 0.17	6.97 ± 0.66	2.79 ± 0.99
<i>Fg06068</i>	FGSG_06068.3	1.07 ± 0.32	1.14 ± 0.43	1.22 ± 0.45	3.14 ± 2.95	1.31 ± 0.52	13.96 ± 0.25	2.01 ± 0.67	0.33 ± 0.02	1.59 ± 1.13	0.97 ± 0.43	0.52 ± 0.17	10.75 ± 0.78	5.12 ± 0.22	0.95 ± 0.34	0.97 ± 0.26	2.46 ± 0.36	0.84 ± 0.05
<i>Fg01786</i>	FGSG_01786.3	1.06 ± 0.31	1.18 ± 0.51	0.90 ± 0.46	2.31 ± 0.34	1.65 ± 0.44	0.96 ± 0.85	1.98 ± 0.88	0.75 ± 0.32	1.28 ± 0.86	1.05 ± 0.72	0.77 ± 0.33	11.87 ± 1.25	1.41 ± 0.55	1.77 ± 0.58	0.46 ± 0.28	2.88 ± 0.16	0.63 ± 0.45
<i>Fg01767</i>	FGSG_01767.3	1.04 ± 0.32	1.47 ± 0.78	0.49 ± 0.29	0.78 ± 0.69	0.97 ± 0.29	1.23 ± 0.88	2.78 ± 0.30	0.63 ± 0.06	0.47 ± 0.20	2.39 ± 0.32	0.71 ± 0.42	10.46 ± 0.78	2.62 ± 0.11	1.19 ± 0.08	0.56 ± 0.12	1.95 ± 0.73	0.40 ± 0.03
<i>Fg01745</i>	FGSG_01745.3	1.04 ± 0.32	1.37 ± 0.76	1.64 ± 1.10	73.25 ± 58.11	3.14 ± 0.34	2.03 ± 0.76	3.99 ± 0.99	1.80 ± 0.47	1.97 ± 0.71	1.13 ± 0.57	1.18 ± 0.32	27.76 ± 8.61	18.33 ± 13.94	2.49 ± 0.15	1.05 ± 0.34	14.28 ± 9.88	0.82 ± 0.13
<i>Fg01740</i>	FGSG_01740.3	1.03 ± 0.25	1.23 ± 1.05	0.68 ± 0.35	14.13 ± 7.94	4.20 ± 1.14	4.59 ± 3.79	5.20 ± 3.22	1.65 ± 1.15	1.45 ± 1.05	0.93 ± 0.42	1.25 ± 0.37	18.95 ± 4.61	1059 ± 5.55	2.13 ± 0.89	1.18 ± 0.90	9.46 ± 6.72	0.82 ± 0.26

<i>Fg01739</i>	FGSG_01739.3	1.01 ± 0.11	1.20 ± 1.22	1.12 ± 0.89	6.93 ± 2.23	2.07 ± 0.31	1.58 ± 0.40	3.95 ± 2.16	1.29 ± 0.36	2.18 ± 1.43	1.18 ± 1.02	1.06 ± 0.21	17.69 ± 1.57	3.18 ± 0.95	4.46 ± 1.02	0.82 ± 0.28	7.72 ± 0.30	0.84 ± 0.12
<i>Fg01722</i>	FGSG_01722.3	1.05 ± 0.38	1.31 ± 0.79	1.07 ± 0.92	4.52 ± 0.03	2.64 ± 0.56	0.72 ± 0.13	9.40 ± 2.70	3.26 ± 1.88	3.09 ± 2.22	1.63 ± 0.42	1.98 ± 0.82	47.59 ± 6.62	10.23 ± 0.57	2.12 ± 0.42	3.44 ± 0.69	9.76 ± 0.64	0.78 ± 0.27
<i>Fg05113</i>	FGSG_05113.3	1.13 ± 0.49	1.11 ± 0.66	0.77 ± 0.41	16.65 ± 9.01	2.14 ± 1.58	4.35 ± 2.09	4.11 ± 1.55	1.30 ± 0.46	2.93 ± 2.67	1.11 ± 0.62	0.95 ± 0.12	18.35 ± 8.01	9.61 ± 6.07	4.11 ± 2.26	0.70 ± 0.31	9.67 ± 5.42	0.73 ± 0.25
<i>Fg01583</i>	FGSG_01583.3	1.03 ± 0.27	1.05 ± 0.64	0.90 ± 0.58	6.90 ± 1.63	2.36 ± 0.61	27.55 ± 2.40	1.78 ± 0.41	0.68 ± 0.28	1.03 ± 0.29	1.05 ± 0.28	0.73 ± 0.30	12.72 ± 3.76	1.64 ± 0.46	1.66 ± 0.29	0.72 ± 0.33	3.78 ± 0.99	0.96 ± 0.20
<i>Fg01048</i>	FGSG_01048.3	1.00 ± 0.03	1.16 ± 0.53	0.76 ± 0.56	1.46 ± 1.36	1.01 ± 0.06	0.71 ± 0.69	2.63 ± 0.75	1.11 ± 0.21	0.85 ± 0.39	0.64 ± 0.23	0.53 ± 0.12	2.64 ± 0.77	1.35 ± 0.97	1.23 ± 0.33	0.68 ± 0.19	1.62 ± 0.60	0.55 ± 0.02
<i>Fg02419</i>	FGSG_02419.3	1.03 ± 0.30	1.30 ± 0.78	0.82 ± 0.06	2.41 ± 0.66	2.55 ± 0.88	0.92 ± 0.79	10.74 ± 0.44	1.23 ± 0.18	1.90 ± 0.79	1.26 ± 0.24	0.73 ± 0.05	133.62 ± 47.33	13.82 ± 8.73	4.03 ± 1.47	2.94 ± 0.27	8.40 ± 3.22	13.21 ± 0.69
<i>Fg02371</i>	FGSG_02371.3	1.17 ± 0.51	1.21 ± 0.84	1.50 ± 1.17	15.96 ± 5.67	4.33 ± 0.65	2.41 ± 1.94	14.26 ± 3.93	3.83 ± 0.43	4.40 ± 3.13	2.33 ± 1.94	3.81 ± 1.33	97.42 ± 16.35	18.68 ± 6.77	8.79 ± 4.19	2.82 ± 3.09	53.47 ± 21.35	1.79 ± 0.73
<i>Fg02367</i>	FGSG_02367.3	1.04 ± 0.34	1.08 ± 0.52	1.28 ± 0.98	12.86 ± 2.06	3.29 ± 2.35	1.78 ± 1.04	10.65 ± 4.76	1.36 ± 0.21	2.00 ± 1.26	0.67 ± 0.37	1.08 ± 0.32	667.88 ± 14.56	19.48 ± 1.98	3.66 ± 1.19	1.28 ± 0.67	11.17 ± 1.14	0.78 ± 0.51
<i>Fg02366</i>	FGSG_02366.3	1.14 ± 0.64	1.16 ± 0.62	1.06 ± 0.56	35.77 ± 28.45	2.09 ± 0.82	1.19 ± 0.17	1.97 ± 0.48	0.95 ± 0.32	0.65 ± 0.42	1.01 ± 0.21	0.62 ± 0.15	9.62 ± 3.54	2.04 ± 0.51	1.10 ± 0.33	0.95 ± 0.11	2.81 ± 0.53	1.59 ± 0.49
<i>Fg02344</i>	FGSG_02344.3	1.05 ± 0.35	1.15 ± 0.66	1.16 ± 0.72	35.75 ± 27.52	2.91 ± 0.49	6.76 ± 5.21	3.44 ± 1.57	1.38 ± 0.46	1.77 ± 1.26	0.75 ± 0.42	0.82 ± 0.10	25.61 ± 2.59	6.74 ± 0.42	2.74 ± 0.60	1.01 ± 0.33	6.39 ± 1.23	1.76 ± 0.30
<i>Fg03796</i>	FGSG_03796.3	1.02 ± 0.20	1.21 ± 0.46	1.10 ± 0.87	39.58 ± 31.11	2.71 ± 0.72	3.44 ± 2.57	0.91 ± 0.35	0.61 ± 0.12	0.90 ± 0.65	0.50 ± 0.18	0.58 ± 0.06	5.99 ± 0.62	0.68 ± 0.05	1.09 ± 0.53	0.42 ± 0.18	0.21 ± 0.34	1.73 ± 0.15
<i>Fg03741</i>	FGSG_03741.3	1.00 ± 0.04	1.04 ± 0.29	1.12 ± 0.11	2.84 ± 1.79	0.80 ± 0.29	4.52 ± 2.03	1.32 ± 0.05	0.79 ± 0.50	0.40 ± 0.16	1.34 ± 0.02	1.05 ± 0.44	6.03 ± 0.53	4.98 ± 0.28	1.25 ± 0.08	1.99 ± 0.63	0.93 ± 0.38	0.43 ± 0.17
<i>Fg00863</i>	FGSG_00863.3	1.06 ± 0.39	1.19 ± 0.52	1.07 ± 0.37	0.33 ± 0.27	1.46 ± 0.03	0.56 ± 0.21	2.68 ± 0.11	0.94 ± 0.31	0.55 ± 0.11	1.34 ± 0.12	0.64 ± 0.02	4.91 ± 0.17	0.95 ± 0.36	0.63 ± 0.02	1.31 ± 0.09	0.64 ± 0.01	0.57 ± 0.21
<i>Fg00012</i>	FGSG_00012.3	1.14 ± 0.57	1.03 ± 0.72	0.18 ± 0.09	3.38 ± 2.92	2.07 ± 0.44	1.21 ± 0.93	2.88 ± 0.85	2.76 ± 1.25	1.49 ± 1.35	0.90 ± 0.10	1.60 ± 0.34	7.67 ± 0.79	3.51 ± 2.08	1.88 ± 0.46	0.75 ± 0.20	5.33 ± 2.27	0.52 ± 0.30
<i>CLM2</i>	FGSG_00007.3	1.02 ± 0.23	1.29 ± 0.53	1.30 ± 0.98	8.57 ± 0.90	2.23 ± 0.55	45.72 ± 18.56	3.58 ± 1.46	2.03 ± 0.43	2.36 ± 2.24	1.21 ± 0.60	1.31 ± 0.63	18.49 ± 7.41	3.64 ± 0.53	3.96 ± 1.64	1.00 ± 0.72	11.23 ± 2.04	0.75 ± 0.30
<i>TR11</i>	FGSG_00072.3	1.00 ± 0.04	1.00 ± 0.07	1.05 ± 0.10	2.22 ± 1.46	0.96 ± 0.04	2.14 ± 1.26	15.07 ± 1.69	1.52 ± 0.51	0.73 ± 0.29	1.93 ± 0.06	0.80 ± 0.32	24.32 ± 9.92	2.90 ± 1.02	1.05 ± 0.10	1.05 ± 0.06	1.52 ± 0.59	0.95 ± 0.06
<i>Fg00127</i>	FGSG_00127.3	1.04 ± 0.30	1.07 ± 0.38	0.89 ± 0.54	49.42 ± 38.87	1.61 ± 0.29	1.59 ± 1.51	1.45 ± 0.43	0.70 ± 0.31	1.00 ± 0.83	0.60 ± 0.31	0.54 ± 0.25	9.77 ± 3.94	1.38 ± 0.24	1.66 ± 1.11	0.54 ± 0.20	3.18 ± 0.17	0.72 ± 0.50
<i>Fg08412</i>	FGSG_08412.3	1.00 ± 0.07	1.48 ± 0.83	0.76 ± 0.34	30.75 ± 19.51	4.41 ± 2.19	5.27 ± 3.32	11.51 ± 1.63	1.86 ± 1.02	1.55 ± 1.23	1.11 ± 0.46	1.31 ± 0.40	40.75 ± 1.07	14.85 ± 10.71	2.73 ± 1.58	1.55 ± 0.46	12.46 ± 8.28	0.97 ± 0.21
<i>Fg08377</i>	FGSG_08377.3	1.08 ± 0.45	1.03 ± 0.27	0.35 ± 0.03	1.28 ± 1.08	0.68 ± 0.02	0.57 ± 0.17	12.34 ± 8.03	0.93 ± 0.38	1.48 ± 1.34	0.47 ± 0.21	0.95 ± 0.35	25.49 ± 16.41	2.70 ± 0.20	1.18 ± 0.08	0.97 ± 0.33	2.57 ± 0.09	1.11 ± 0.44
<i>Fg08320</i>	FGSG_08320.3	1.03 ± 0.26	1.17 ± 0.64	0.76 ± 0.42	6.73 ± 1.85	1.96 ± 0.55	2.05 ± 0.88	1.86 ± 0.44	1.08 ± 0.25	1.71 ± 1.42	0.85 ± 0.46	0.94 ± 0.50	14.13 ± 1.02	1.98 ± 1.01	2.10 ± 0.92	0.90 ± 0.50	7.27 ± 1.09	0.78 ± 0.42
<i>Fg08207</i>	FGSG_08207.3	1.40 ± 0.58	1.82 ± 1.02	0.27 ± 0.17	1.39 ± 0.74	2.01 ± 0.61	2.00 ± 1.79	13.22 ± 0.55	1.33 ± 0.52	1.65 ± 1.44	1.21 ± 0.63	1.97 ± 1.19	7.38 ± 1.16	1.82 ± 0.12	1.56 ± 0.40	2.03 ± 0.80	5.80 ± 3.65	0.28 ± 0.20

<i>Fg01972</i>	FGSG_01972.3	1.07 ± 0.43	1.40 ± 1.07	1.91 ± 1.35	18.21 ± 3.79	2.98 ± 0.29	4.18 ± 1.77	4.08 ± 1.45	1.00 ± 0.40	0.11 ± 0.09	1.60 ± 1.22	0.49 ± 0.19	1.87 ± 0.72	3.24 ± 1.00	0.20 ± 0.06	31.64 ± 10.35	1.33 ± 0.05	0.32 ± 0.05
<i>Fg08187</i>	FGSG_08187.3	1.00 ± 0.05	1.18 ± 0.62	0.67 ± 0.47	6.34 ± 0.15	1.53 ± 0.37	3.19 ± 0.16	1.79 ± 0.15	0.85 ± 0.17	1.04 ± 0.85	0.56 ± 0.40	0.49 ± 0.07	6.33 ± 1.48	1.13 ± 0.24	1.19 ± 0.51	0.48 ± 0.09	3.78 ± 0.48	0.47 ± 0.15
<i>CYP51B</i>	FGSG_01000.3	1.07 ± 0.42	1.17 ± 0.66	0.27 ± 0.09	0.33 ± .28	0.26 ± 0.88	2.02 ± 0.63	3.70 ± 0.11	0.86 ± 0.08	0.60 ± 0.34	2.01 ± 1.74	0.44 ± 0.02	57.49 ± 2.24	10.65 ± 3.62	2.48 ± 1.84	2.70 ± 0.86	2.65 ± 0.96	2.08 ± 0.70
<i>Fg08186</i>	FGSG_08186.3	1.21 ± 0.52	1.12 ± 0.67	1.10 ± 0.69	6.21 ± 0.64	2.81 ± 0.81	16.46 ± 3.58	2.71 ± 0.79	1.25 ± 0.41	1.63 ± 1.25	0.53 ± 0.39	0.81 ± 0.09	15.11 ± 2.94	1.77 ± 0.24	2.33 ± 1.10	0.83 ± 0.69	4.71 ± 1.15	3.84 ± 3.13
<i>Fg08183</i>	FGSG_08183.3	1.08 ± 0.47	1.27 ± 0.62	0.71 ± 0.70	10.93 ± 4.90	0.43 ± 0.16	6.35 ± 0.61	0.95 ± 0.30	0.34 ± 0.14	1.07 ± 0.50	1.56 ± 1.38	0.25 ± 0.04	4.17 ± 0.89	0.74 ± 0.35	0.65 ± 0.22	0.19 ± 0.11	1.32 ± 0.23	0.14 ± 0.05
<i>CYP51C</i>	FGSG_07977.3	1.16 ± 0.70	1.03 ± 0.80	0.95 ± 0.74	5.91 ± 0.25	2.00 ± 0.94	2.00 ± 1.41	1.92 ± 0.12	0.86 ± 0.28	1.72 ± 1.61	0.68 ± 0.30	1.46 ± 0.63	29.70 ± 3.02	3.23 ± 0.39	2.70 ± 1.61	0.44 ± 0.27	3.88 ± 0.44	0.48 ± 0.23
<i>Fg08005</i>	FGSG_08005.3	1.32 ± 0.72	1.21 ± 0.36	1.81 ± 1.70	11.13 ± 3.78	2.86 ± 0.64	3.09 ± 1.10	2.62 ± 0.81	0.70 ± 0.54	1.03 ± 0.63	0.71 ± 0.60	1.59 ± 1.13	12.52 ± 0.63	3.05 ± 0.48	5.96 ± 1.91	0.60 ± 0.56	6.54 ± 1.21	0.90 ± 0.47
<i>Fg10629</i>	FGSG_10629.3	1.13 ± 0.64	1.26 ± 0.87	1.06 ± 1.31	37.03 ± 7.59	1.07 ± 0.28	31.02 ± 21.62	1.82 ± 1.86	1.62 ± 2.03	0.65 ± 0.91	1.56 ± 2.19	0.67 ± 0.63	2.78 ± 1.75	6.15 ± 4.00	2.27 ± 3.42	1.12 ± 1.12	14.09 ± 10.89	0.22 ± 0.25
<i>Fg10695</i>	FGSG_10695.3	1.04 ± 0.28	1.11 ± 0.48	1.48 ± 0.94	4.52 ± 1.41	2.50 ± 0.40	2.33 ± 0.09	2.67 ± 0.84	0.79 ± 0.46	0.64 ± 0.23	0.63 ± 0.17	0.87 ± 0.26	7.87 ± 1.77	1.76 ± 0.21	1.40 ± 0.35	1.07 ± 0.46	3.83 ± 0.61	1.14 ± 0.17
<i>Fg10910</i>	FGSG_10910.3	1.07 ± 0.41	1.02 ± 0.26	1.44 ± 0.06	2.81 ± 2.30	2.91 ± 0.17	2.17 ± 0.97	2.79 ± 0.98	0.71 ± 0.30	0.35 ± 0.16	1.05 ± 0.88	0.48 ± 0.02	5.60 ± 1.96	3.92 ± 0.10	4.33 ± 3.79	1.92 ± 0.12	2.60 ± 0.84	2.17 ± 0.74
<i>Fg11389</i>	FGSG_11389.3	1.08 ± 0.39	1.12 ± 0.58	0.95 ± 0.56	22.69 ± 10.27	2.93 ± 0.71	9.51 ± 2.68	3.88 ± 0.69	2.09 ± 0.87	1.33 ± 0.54	0.90 ± 0.64	1.61 ± 0.84	17.18 ± 1.06	4.75 ± 0.41	5.09 ± 1.31	0.84 ± 0.61	22.54 ± 9.22	0.89 ± 0.49
<i>Fg11424</i>	FGSG_11424.3	1.02 ± 0.19	1.09 ± 0.47	0.83 ± 0.47	5.48 ± 1.84	3.27 ± 1.02	11.67 ± 5.23	3.07 ± 0.67	1.06 ± 0.68	0.72 ± 0.25	0.60 ± 0.20	0.77 ± 0.39	26.36 ± 1.58	7.80 ± 1.12	1.90 ± 0.16	0.68 ± 0.22	4.12 ± 0.44	1.12 ± 0.73
<i>Fg11282</i>	FGSG_11282.3	1.00 ± 0.06	1.16 ± 0.63	0.63 ± 0.40	0.54 ± 0.42	1.13 ± 0.13	0.48 ± 1.07	1.82 ± 0.06	0.72 ± 0.33	0.63 ± 0.19	0.69 ± 0.25	1.39 ± 0.56	3.83 ± 0.25	2.12 ± 1.46	1.93 ± 1.66	0.69 ± 0.18	1.32 ± 0.42	1.50 ± 0.54
<i>Fg11303</i>	FGSG_11303.3	1.04 ± 0.29	1.08 ± 0.42	1.06 ± 0.86	8.00 ± 0.35	2.77 ± 0.86	3.01 ± 1.66	2.26 ± 0.88	0.94 ± 0.50	0.96 ± 0.54	0.51 ± 0.13	0.67 ± 0.30	12.84 ± 0.67	10.28 ± 1.35	7.57 ± 5.24	0.78 ± 0.31	5.95 ± 1.81	0.95 ± 0.50
<i>Fg11038</i>	FGSG_11038.3	1.02 ± 0.22	1.14 ± 0.56	0.97 ± 0.63	1.49 ± 0.24	2.09 ± 0.86	8.08 ± 3.77	0.77 ± 0.10	1.32 ± 1.40	0.61 ± 0.23	0.62 ± 0.33	0.78 ± 0.50	6.38 ± 1.23	1.29 ± 0.37	0.88 ± 0.06	0.82 ± 0.19	5.39 ± 1.60	0.61 ± 0.23
<i>Fg11108</i>	FGSG_11108.3	1.05 ± 0.33	1.16 ± 0.66	0.72 ± 0.56	5.64 ± 0.92	2.78 ± 0.13	3.45 ± 0.90	4.23 ± 1.71	2.16 ± 1.52	1.20 ± 0.36	0.75 ± 0.55	2.20 ± 0.69	18.13 ± 7.61	15.93 ± 3.53	3.35 ± 0.32	1.39 ± 0.78	22.13 ± 7.77	0.74 ± 0.45
<i>Fg10960</i>	FGSG_10960.3	1.01 ± 0.14	1.00 ± 0.07	0.70 ± 0.27	1.14 ± 1.18	1.02 ± 0.13	0.25 ± 0.12	4.73 ± 0.50	0.97 ± 0.43	0.90 ± 0.38	0.73 ± 0.25	1.18 ± 0.14	22.39 ± 4.44	1.85 ± 0.69	0.57 ± 0.08	0.78 ± 0.24	1.32 ± 0.31	0.63 ± 0.21
<i>Fg10991</i>	FGSG_10991.3	1.16 ± 0.54	1.05 ± 0.34	1.52 ± 1.24	8.47 ± 0.78	3.22 ± 2.03	2.31 ± 0.32	2.32 ± 0.61	1.00 ± 0.49	1.04 ± 0.67	0.49 ± 0.13	0.86 ± 0.31	10.27 ± 1.56	2.26 ± 0.52	2.42 ± 0.52	0.43 ± 0.18	4.73 ± 0.77	0.91 ± 0.80
<i>Fg11002</i>	FGSG_11002.3	1.06 ± 0.39	1.03 ± 0.27	0.87 ± 0.18	14.27 ± 11.91	10.29 ± 3.47	4.95 ± 2.87	5.09 ± 0.84	2.49 ± 1.22	1.51 ± 0.61	1.71 ± 0.84	2.83 ± 0.98	31.73 ± 7.02	8.87 ± 1.59	40.97 ± 27.82	1.38 ± 0.53	10.45 ± 0.94	1.24 ± 0.92
<i>Fg11024</i>	FGSG_11024.3	1.07 ± 0.42	1.01 ± 0.16	0.14 ± 0.05	0.34 ± 0.30	1.46 ± 0.46	1.14 ± 0.34	8.94 ± 0.40	0.67 ± 0.45	0.37 ± 0.25	1.13 ± 1.17	0.29 ± 0.02	18.41 ± 0.47	1.76 ± 0.54	2.56 ± 2.22	0.90 ± 0.25	1.05 ± 0.11	3.94 ± 1.16
<i>Fg11465</i>	FGSG_11465.3	1.08 ± 0.75	1.03 ± 0.25	0.15 ± 0.06	1.67 ± 0.40	0.86 ± 0.14	0.64 ± 0.47	0.20 ± 0.06	0.82 ± 0.58	0.32 ± 0.05	0.95 ± 0.69	0.43 ± 0.13	1.45 ± 0.36	1.50 ± 1.29	2.95 ± 1.33	1.86 ± 0.66	0.85 ± 0.44	1.63 ± 0.10

<i>Fg11498</i>	FGSG_11498.3	1.02 ± 0.20	1.04 ± 0.32	1.56 ± 0.51	1.83 ± 1.99	2.03 ± 0.07	1.81 ± 1.34	3.04 ± 0.03	0.81 ± 0.71	1.02 ± 0.45	0.75 ± 0.63	0.69 ± 0.04	0.68 ± 0.39	5.70 ± 2.81	1.78 ± 0.79	0.99 ± 0.33	4.05 ± 1.84	1.09 ± 0.06
<i>Fg11536</i>	FGSG_11536.3	1.07 ± 0.27	1.04 ± 0.32	0.58 ± 0.46	19.87 ± 12.60	6.97 ± 2.75	1.43 ± 1.30	2.36 ± 0.39	2.38 ± 1.21	1.51 ± 1.21	0.98 ± 0.44	2.49 ± 1.22	16.75 ± 2.79	12.40 ± 5.20	1.69 ± 0.31	0.66 ± 0.25	9.90 ± 2.93	0.96 ± 1.07
<i>Fg10451</i>	FGSG_10451.3	1.07 ± 0.42	1.18 ± 0.68	1.34 ± 1.06	21.61 ± 9.30	1.81 ± 0.82	2.47 ± 2.23	1.44 ± 0.97	0.88 ± 0.81	0.30 ± 0.08	0.65 ± 0.32	0.63 ± 0.58	5.59 ± 1.51	2.48 ± 2.71	2.25 ± 1.96	0.75 ± 0.20	2.46 ± 1.11	1.08 ± 0.49
<i>Fg10461</i>	FGSG_10461.3	1.03 ± 0.28	1.07 ± 0.44	0.94 ± 0.67	6.43 ± 4.83	3.39 ± 1.54	4.69 ± 0.71	4.56 ± 0.63	0.85 ± 0.65	0.72 ± 0.30	0.52 ± 0.06	1.21 ± 0.42	35.94 ± 2.41	4.05 ± 0.67	1.96 ± 0.26	0.58 ± 0.25	5.10 ± 0.63	0.96 ± 0.36
<i>Fg09351</i>	FGSG_09351.3	1.18 ± 0.69	1.04 ± 0.29	0.73 ± 0.26	1.49 ± 1.41	1.44 ± 0.56	2.23 ± 1.35	0.08 ± 0.01	0.75 ± 0.66	0.46 ± 0.13	2.89 ± 2.84	0.71 ± 0.45	0.65 ± 0.38	0.48 ± 0.14	0.46 ± 0.17	0.66 ± 0.05	0.22 ± 0.10	0.36 ± 0.12
<i>Fg09195</i>	FGSG_09195.3	1.06 ± 0.40	1.42 ± 0.97	1.08 ± 0.47	1.33 ± 1.11	2.70 ± 0.12	0.69 ± 0.50	2.53 ± 0.06	0.47 ± 0.19	0.30 ± 0.02	0.68 ± 0.58	0.51 ± 0.19	10.32 ± 0.15	1.83 ± 0.60	2.13 ± 1.49	0.92 ± 0.24	0.92 ± 0.30	4.12 ± 1.47
<i>Fg09916</i>	FGSG_09916.3	1.00 ± 0.10	1.03 ± 0.25	0.39 ± 0.16	1.95 ± 0.90	0.79 ± 0.26	0.79 ± 0.48	5.37 ± 0.19	0.49 ± 0.19	0.45 ± 0.17	0.68 ± 0.57	0.61 ± 0.04	5.37 ± 0.35	1.61 ± 1.01	1.11 ± 0.11	0.63 ± 0.05	3.17 ± 2.09	0.77 ± 0.29
<i>Fg09671</i>	FGSG_09671.3	1.04 ± 0.28	1.14 ± 0.47	0.30 ± 0.03	4.02 ± 0.13	1.66 ± 0.52	0.59 ± 0.02	1.74 ± 0.26	0.67 ± 0.41	0.61 ± 0.18	0.38 ± 0.12	0.55 ± 0.24	8.46 ± 0.75	1.78 ± 0.19	1.80 ± 0.37	0.48 ± 0.03	5.28 ± 2.94	0.93 ± 0.46
<i>Fg11655</i>	FGSG_11655.3	1.00 ± 0.07	1.15 ± 0.61	1.24 ± 1.06	2.20 ± 2.05	2.01 ± 0.16	1.52 ± 1.22	7.02 ± 0.34	1.03 ± 0.23	0.79 ± 0.12	1.80 ± 0.17	0.89 ± 0.22	28.99 ± 1.60	1.90 ± 0.15	2.31 ± 1.34	5.79 ± 1.81	5.41 ± 2.02	6.36 ± 2.44
<i>Fg12534</i>	FGSG_12534.3	1.00 ± 0.07	1.04 ± 0.28	1.41 ± 0.55	0.83 ± 0.69	3.95 ± 0.12	1.36 ± 0.48	0.88 ± 0.30	1.05 ± 0.58	1.18 ± 0.11	1.58 ± 1.08	1.18 ± 0.06	0.96 ± 0.37	3.66 ± 1.05	1.13 ± 0.07	2.48 ± 0.15	0.60 ± 0.02	0.48 ± 0.03
<i>Fg12568</i>	FGSG_12568.3	1.21 ± 0.28	1.09 ± 0.45	0.94 ± 0.79	6.78 ± 1.62	2.53 ± 0.84	1.94 ± 1.29	1.90 ± 0.66	0.88 ± 0.52	0.87 ± 0.38	0.53 ± 0.12	0.94 ± 0.32	12.43 ± 1.75	4.81 ± 0.26	2.04 ± 0.36	0.80 ± 0.27	4.99 ± 0.46	0.87 ± 0.40
<i>Fg08809</i>	FGSG_08809.3	1.08 ± 0.44	1.02 ± 0.22	0.53 ± 0.20	1.48 ± 1.03	0.97 ± 0.37	3.97 ± 2.00	2.73 ± 0.89	0.58 ± 0.39	0.32 ± 0.13	0.33 ± 0.12	0.69 ± 0.23	7.75 ± 0.53	0.57 ± 0.35	2.07 ± 1.82	0.69 ± 0.19	1.70 ± 0.17	0.55 ± 0.21
<i>Fg13176</i>	FGSG_13176.3	1.00 ± 0.11	1.14 ± 0.58	0.99 ± 0.61	2.65 ± 0.57	1.02 ± 0.16	48.59 ± 17.39	0.83 ± 0.36	0.89 ± 0.76	0.29 ± 0.09	1.10 ± 0.72	0.66 ± 0.22	3.38 ± 1.66	1.82 ± 0.19	0.43 ± 0.04	1.25 ± 0.48	1.32 ± 0.60	0.88 ± 0.52
<i>Fg12737</i>	FGSG_12737.3	1.00 ± 0.05	1.17 ± 0.66	1.49 ± 0.53	1.19 ± 1.20	2.00 ± 0.14	0.46 ± 0.42	1.35 ± 0.44	0.60 ± 0.43	0.33 ± 0.12	4.02 ± 1.02	0.47 ± 0.03	0.97 ± 0.10	0.49 ± 0.05	2.82 ± 2.79	2.26 ± 1.39	0.69 ± 0.27	1.03 ± 0.06
<i>Fg12599</i>	FGSG_12599.3	1.29 ± 0.52	1.13 ± 0.57	0.95 ± 0.67	22.20 ± 15.79	18.20 ± 5.31	1.17 ± 1.15	3.57 ± 0.57	1.60 ± 0.39	2.14 ± 1.86	1.77 ± 0.59	4.21 ± 0.76	24.47 ± 14.06	14.61 ± 2.55	2.07 ± 0.20	1.84 ± 0.68	6.52 ± 1.88	1.84 ± 1.54
<i>Fg13445</i>	FGSG_13445.3	1.00 ± 0.06	1.12 ± 0.49	1.20 ± 0.90	2.32 ± 1.19	4.51 ± 1.56	1.77 ± 0.15	3.23 ± 1.42	1.68 ± 1.03	0.89 ± 0.32	0.57 ± 0.25	1.79 ± 1.25	12.53 ± 1.71	4.98 ± 0.89	1.68 ± 0.63	0.85 ± 0.36	8.92 ± 2.97	1.17 ± 0.71
<i>Fg13459</i>	FGSG_13459.3	1.00 ± 0.11	1.10 ± 0.25	1.09 ± 0.97	15.07 ± 1.11	1.93 ± 0.36	10.07 ± 6.05	2.77 ± 1.02	1.22 ± 0.53	0.78 ± 0.24	0.44 ± 0.15	1.01 ± 0.20	14.48 ± 1.73	1.82 ± 0.60	3.99 ± 0.65	0.66 ± 0.38	6.22 ± 1.45	1.33 ± 1.48
<i>Fg14007</i>	FGSG_14007.3	1.03 ± 0.25	1.15 ± 0.63	1.55 ± 1.06	26.58 ± 19.56	3.19 ± 1.42	8.12 ± 3.46	2.27 ± 0.93	1.14 ± 0.45	1.42 ± 0.84	1.58 ± 1.08	1.41 ± 0.40	27.55 ± 11.69	18.77 ± 10.99	2.64 ± 1.35	0.52 ± 0.28	8.42 ± 0.42	0.68 ± 0.31
<i>Fg13739</i>	FGSG_13739.3	1.03 ± 0.24	1.20 ± 0.84	1.10 ± 0.92	34.47 ± 3.60	2.81 ± 1.30	2.00 ± 0.65	2.58 ± 0.65	2.04 ± 0.98	2.22 ± 1.68	1.11 ± 0.44	2.35 ± 0.57	51.28 ± 16.15	5.70 ± 0.85	3.60 ± 2.32	0.93 ± 0.52	13.25 ± 4.80	0.51 ± 0.09
<i>Fg13796</i>	FGSG_13796.3	1.34 ± 0.75	1.59 ± 1.23	0.36 ± 0.15	3.80 ± 0.77	3.10 ± 2.5	2.55 ± 1.89	0.93 ± 0.35	1.95 ± 1.97	0.68 ± 0.08	1.74 ± 1.14	0.47 ± 0.33	5.68 ± 1.82	2.95 ± 1.05	0.47 ± 0.34	0.90 ± 0.41	3.46 ± 2.19	1.49 ± 0.97
<i>Fg13797</i>	FGSG_13797.3	1.14 ± 0.60	1.13 ± 0.67	0.30 ± 0.13	2.09 ± 1.34	0.58 ± 0.20	1.08 ± 0.76	0.42 ± 0.07	1.52 ± 1.49	0.37 ± 0.06	1.10 ± 0.39	0.24 ± 1.11	1.43 ± 0.32	0.81 ± 0.47	0.37 ± 0.21	0.26 ± 0.13	1.73 ± 0.93	0.39 ± 0.02

DISCUSSION

The construction of genome-wide knockout mutant libraries with corresponding phenotypic datasets in model organisms has led to great progresses in basic and applied biology. P450 enzymes play important roles in diverse types of primary and secondary metabolism and the degradation of xenobiotics in eukaryotic organisms (van den Brink et al., 1998). Plant pathogenic fungi possess a relatively large number of P450 genes compared with other eukaryotic organisms (Črešnar and Petrič, 2011), suggesting that P450-mediated molecular processes are particularly important for these fungal species. However, no systematic functional analysis of P450s has yet been attempted in filamentous fungi, including plant pathogens. In this study, I targeted each of 119 putative P450s for gene deletion and generated a comprehensive phenome of 102 P450s related to 38 traits in the plant pathogenic ascomycete *F. graminearum*. In most cases, the P450 mutants with mutant phenotypes showed a single phenotypic defect, demonstrating that my genetic resources are valuable for further genetic studies on specific fungal developmental processes, including sexual development and virulence. In particular, genes specifically involved in virulence (5), conidiation (1), and sexual development (2) have rarely been found in *F. graminearum*.

There has been considerable interest in identifying factors involved in reproduction in *F. graminearum* because this fungus utilizes sexual (ascospores) and asexual (conidia) spores for primary and secondary infection, respectively (Leslie and Summerell, 2006). Some P450 enzymes have been reported to regulate

development by producing phytohormones in plants (Kushiro et al., 2004). In *Aspergillus nidulans*, the hormone-like fatty acid-derived oxylipins serve as signal molecules that modulate reproductive processes by affecting the timing and balance of asexual and sexual development (Fischer and Keller, 2016). *CYP4* orthologs encoding elongationless 2 are involved in sexual development in the mushroom-forming fungi *Coprinopsis cinerea* and *Agaricus bisporus* (De Groot et al., 1997; Kamada, 2002). Based on expression profiles, P450s appear to have distinct functions in sexual and asexual reproduction processes in *F. graminearum* (Fig. 2).

I discovered that sex-induced *Fg08320* and *Fg09671* are required for selective perithecium maturation and initial perithecium formation, respectively, in *F. graminearum*. The $\Delta Fg09671$ strains failed to switch its developmental stage from mycelial growth to perithecium production, and the maturation of overproduced small perithecia was arrested in the $\Delta Fg08320$ strains; in contrast, in the wild-type strain, mycelial growth was completely inhibited and perithecia were selectively matured among the initial perithecia after sexual induction (Fig. 5). I suspect that *Fg08320* and *Fg09671*-mediated signaling molecules are involved in each step of perithecium production in *F. graminearum*. Likewise, *Fg06068* is required for phialide formation and function in this fungus (Fig. 4). Further studies are necessary to identify the P450-mediated signaling molecules to elucidate the molecular mechanisms underlying fungal reproduction processes.

Plants synthesize and accumulate plant antibiotics known as phytoalexins that

are toxic to most fungal pathogens (Paxton, 1980). However, fungi frequently possess the ability to detoxify plant toxins using P450s. Pisatin demethylase (*PDA*, CYP57A1) of *Nectria haematococca* is a well-known P450 enzyme that degrades pisatin, a fungistatic isoflavonoid pterocarpan produced by host plants (George et al., 1998). Introduction of the *PDA* (*FgPDA1*) ortholog into a *PDA*-deficient *F. oxysporum* strain was shown to promote pathogenicity in pea (Coleman et al., 2011); however, orthologs of *PDA* are unclear in *F. graminearum*. Five P450 mutants (*Fg03700*, *Fg02111*, *Fg00012*, *Fg10451*, and *Fg12737*) were revealed to be specifically defective in virulence (Supporting information Table S1 and Fig. 6), and two of these genes (*Fg03700* and *Fg12737*) are highly upregulated during plant infection (Harris et al., 2016). Moreover, all of these genes were markedly upregulated under at least one xenobiotic condition compared with the untreated control, suggesting that they function in the detoxification of plant toxins in *F. graminearum* (Fig. 10). Because few virulence-specific mutants of *F. graminearum* have been available, further studies addressing these mutants will be helpful for understanding host-pathogen interaction mechanisms.

Azole fungicides comprise a large number of commercially available fungicides, and more than 36 azole drugs have been used in agricultural applications. These compounds specifically bind to the sterol 14 α -demethylase encoded by the *CYP51* gene and therefore inhibit the biosynthesis of ergosterol, which is the principal sterol present in fungal membranes. In *F. graminearum*, single or double deletion of three *CYP51* paralogs (*CYP51A*, *CYP51B*, and

CYP51C) was not found to affect fungal development or ergosterol contents (Liu et al., 2011; Fan et al., 2013), suggesting that *CYP51* genes have redundant functions, and additional P450s are involved in ergosterol biosynthesis. Likewise, 17 P450 mutants showing altered sensitivity to azole drugs are expected to be closely related to ergosterol production in *F. graminearum* either directly or indirectly (Supporting information Table S3). Because the ergosterol biosynthetic pathway is still a good target for fungicides, elucidating the novel mechanisms of these P450s will provide genetic resources for the development or application of new fungicides.

Several filamentous fungi are able to detoxify environmental pollutants via substrate-inducible P450-mediated reactions (Cerniglia et al., 1978; Sutherland, 1992; Bezalel et al., 1997). For example, in the white rot fungus *Phanerochaete chrysosporium*, P450 enzymes show an extraordinary ability to degrade a broad spectrum of aromatic, alicyclic, and aliphatic chemicals (Reddy, 1995). In my study, the transcript levels of most P450 genes were markedly upregulated under at least one xenobiotic condition (Fig. 10). In particular, many P450 genes were highly induced in the presence of 1-naphthol (94 genes), naphthalene (68 genes), 1-dodecanol (75 genes), and 4-octylphenol (76 genes), but only one mutant ($\Delta Fg01972$) showed reduced growth in the 1-dodecanol-supplemented condition, demonstrating that most P450s exhibit overlapping biochemical functions in *F. graminearum*. However, P450 enzymes may not be the only source of tolerance to xenobiotics. Not only P450s but also ATP-binding cassette (ABC) transporters,

major facilitator superfamily (MFS) transporters and other enzymes such as peroxidase have been shown to play important roles in xenobiotic detoxification in *P. chrysosporium* and *C. albicans* (Bumpus and Aust, 1987; Vandeputte et al., 2012). Thus, the requirements for P450s in fungal development are not properly reflected by developmental phenotypes, likely because of their redundant functions among P450s and the existence of other mechanisms (Sanglard et al., 1998).

In conclusion, I constructed a *F. graminearum* P450 mutant library and a comprehensive phenotypic dataset. Most P450s have redundant functions in xenobiotic detoxification and fungal development, suggesting that other approaches, such as RNA silencing, are required for genetic studies on P450s in filamentous fungi. Nevertheless, I identified novel P450s that are specifically involved in fungal reproduction and virulence. This study is the first systemic functional analysis of P450s, and my results provide a platform for further metabolomic and biochemical studies on P450s in filamentous fungi, including plant pathogens.

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e45432.

APPENDIX

Table S1. Primers used in this study.

A. Primer for transformation

Gene name	Oligo name	Sequence
<i>Fg07859</i>	07859.3-5F	CATTGCGAATACCCAGGGAAGTC
	07859.3-5R	gcacaggtacactgttttagagTGACTTCAAAGGCTACTACCAGGGC
	07859.3-3F	ccttcaatcatcttctgtcgACAACCACCATCTGAGCGTGAGT
	07859.3-3R	TCTTGCTTAACCTCGCCACCCCT
	07859.3-5N	CACCTGCGCCTGTGAACCTTGAT
	07859.3-3N	GCTTTGCCAACGTTCTCTGAG
<i>Fg07804</i>	07804.3-5F	CATGAGCCTCTTGGAGTGGTCG
	07804.3-5R	gcacaggtacactgttttagagCCAGCCATGCGTGAGTAGGAGA
	07804.3-3F	ccttcaatcatcttctgtcgTATGGCGCGGCGAATAGAGAC
	07804.3-3R	CCAGCCATGCGTGAGTAGGAGA
	07804.3-5N	CTGGTAAAGTGCTGGCAACAACG
	07804.3-3N	AGCGGGTATAAGGATCAAAATGCG
<i>Fg08079</i>	08079.3-5F	GCTGTATGCCATCCGTCCTACCT
	08079.3-5R	gcacaggtacactgttttagagGCCAGATCAAGGGAAGGACAGTG
	08079.3-3F	ccttcaatcatcttctgtcgCGATGGAGATTTTGGAGGGCTT
	08079.3-3R	CCCCGACTCAGACGCCTACAC
	08079.3-5N	TGGGCACAGCGGGACACT
	08079.3-3N	ACCAAGTCCCCGAAATACCCAGTT
<i>Fg04717</i>	04717.3-5F	CCATCAGAACTGATCCCAAGACCA
	04717.3-5R	gcacaggtacactgttttagagTAACGGACCTTACAATGCGAGA
	04717.3-3F	ccttcaatcatcttctgtcgACTTTGTGGCCATACTTTCTCCCC
	04717.3-3R	CCCTAGATCCCTCGGTCTGACTTC
	04717.3-5N	CGAGAGTGTCATCCGAGCCGTA
	04717.3-3N	GGATCACGGCAGAATATGAACAGG
<i>Fg01868</i>	01868.3-5F	ACAAGAACGGGACGTAATGAGTG
	01868.3-5R	gcacaggtacactgttttagagAAGTTTTCGAATATGGAATGCGAT

B. Primer for qRT-PCR

Name	Oligo name	Sequence
<i>Fg07859</i> FGSG_07859.3	RT1-F	TGGCCCATCGGTCCAAGTCT
	RT1-R	TGTACCCTTTTGGTTGGAATGAT
<i>Fg07804</i> FGSG_07804.3	RT2-F	GTTTCGCTCGTGCTCTTCATCAG
	RT2-R	TCTGCGGGAATAGGCCCTTCTTT
<i>Fg08079</i> FGSG_08079.3	RT3-F	TCACACAGTTTTGCACCACAGTCC
	RT3-R	ATCGCCGATGATATCAAGGACAAG
<i>Fg04717</i> FGSG_04717.3	RT4-F	CCGACTCAAAAGAATGGCACTCA
	RT4-R	AGAAACCAATGGATATAAGGGCGAA
<i>Fg01868</i> FGSG_01868.3	RT6-F	TCAACCGCAGCAGAAGTGG
	RT6-R	CCATGTTTTGCAGTCGCTCCTT
<i>Fg07596</i> FGSG_07596.3	RT7-F	CGGTAACGGTATGCGAGGATGTAT
	RT7-R	ACCTTTCGGTCGTTGCGGAG
<i>Fg05960</i> FGSG_05960.3	RT8-F	AGCTTGCATGTCCATCCCTCTG
	RT8-R	GCTCCTGCTCGACTCGTTTGTA
<i>Fg03860</i> FGSG_03860.3	RT9-F	ACCAAAGATGAGGGAGGCAAAATG
	RT9-R	TCGCTGAGGATGGCTTGACTTC
<i>Fg07765</i> FGSG_07765.3	RT10-F	TGGAGATCTGGCCTTTGGTGAG
	RT10-R	TACCTTTTCCCTCGCCATTTC
<i>Fg02982</i> FGSG_02982.3	RT11-F	TCAAACCGACCCTGTGAAGCA
	RT11-R	CCAGCATTATGTCCAGTAAACCGC
<i>Fg03264</i> FGSG_03264.3	RT12-F	TGACTGGCTTGACAACAGAGTGG
	RT12-R	GGCGTCAGGGCTAGTGAGAAGAT
<i>Fg03260</i> FGSG_03260.3	RT13-F	AGAATCTCAAACATCCCCAAACC
	RT13-R	AGGTGGTAGAAAAGAGCAGAAAGGC
<i>Fg05806</i> FGSG_05806.3	RT14-F	ACGCTCTTGAATGCCGATAATGTT
	RT14-R	ATCCGATCCCGCAATAATAAGGTT

	01868.3-3F	ccttcaatatcatcttctgtcgACGAGACCAATACTTTGATACCCAG
	01868.3-3R	ATCCTGAGCAATCAACCAACTCCA
	01868.3-5N	TCACTCTTCCGAAAACCTCCA
	01868.3-3N	ACATTGCCATTGAAAGAATACCGTCT
Fg07596	07596.3-5F	CCGGTTTTATACCCCACTACTCAG
	07596.3-5R	gcacaggtacacttgttagagTGTGAGTTGTGTTGTGGAGTGTG
	07596.3-3F	ccttcaatatcatcttctgtcgCCAAGATGCGATGCCAGTCTT
	07596.3-3R	CTCGGCACACGTAGATTCACTCTTC
	07596.3-5N	CTCTCGTCGTCGCGGGTTTAG
	07596.3-3N	CGCACTCATGTTACTACGACTTACCG
Fg05960	05960.3-5F	GCCAGACCAAGCAGTATCAGCAAC
	05960.3-5R	gcacaggtacacttgttagagCAAGTTCAATTGGCGCATAGGC
	05960.3-3F	ccttcaatatcatcttctgtcgGAAAAGGTTCCGTGGGCTACTCTC
	05960.3-3R	GGCTGAATGCATCACATTGGAAA
	05960.3-5N	TTCATCCATCGCCACTTCTTCAA
	05960.3-3N	GGACGGGATAACTGCTTGTAGGTTT
Fg03860	03860.3-5F	AAGAAAGAAGAAGCATTGCGTGAGC
	03860.3-5R	gcacaggtacacttgttagagTGACGCGACGGCTGTTCTTTA
	03860.3-3F	ccttcaatatcatcttctgtcgCCTGGCCTAGTCCCTACATAGCG
	03860.3-3R	TCCAGAGGGCCAACAGAGACT
	03860.3-5N	TCACAGAGAGATGCAGCGACACA
	03860.3-3N	TCTCACACCAGTCAGGGACACAAA
Fg07765	07765.3-5F	CGGTCTGGGTCTCATATCTGGTCTC
	07765.3-5R	gcacaggtacacttgttagagGATCCGGATCTTGGACATCTTGC
	07765.3-3F	ccttcaatatcatcttctgtcgTCTTTTGTTAAACGAGGGACCAAGTGT
	07765.3-3R	ACGGTGAATTTGGCAACAGTGTAAAG
	07765.3-5N	AGATGGCTTCGTATGGTCTTGGTG
	07765.3-3N	TGACTCTCTCGGGATGAGCAGGT
Fg02982	02982.3-5F	CAAGCCAGGGTGACTTACACAAGAA
	02982.3-5R	gcacaggtacacttgttagagCGAGATGGGATGTGCGTGTGTTT
	02982.3-3F	ccttcaatatcatcttctgtcgGTTGGTCAGAGAACATCACAGAGCCT

CYP51A FGSG_04092.3	RT15-F	CCACTGCCTCACAAACCGAAAAAC
	RT15-R	GCTGGCCTGCCATGAGTAGAGTAA
Fg01284 FGSG_01284.3	RT16-F	CCTGGGATGGGATGTCAAGTCTC
	RT16-R	CAACGCTCGCTCGATGTAAAAGT
Fg03191 FGSG_03191.3	RT17-F	GGAGGGCAAGGGAATTACTGTCA
	RT17-R	GCGAGACGTTTCAATGTGATGTC
Fg03700 FGSG_03700.3	RT19-F	CGAAGCTGCCGCCATTATTCT
	RT19-R	GCTAGTACCGGGGAACCAAGTCAG
Fg03686 FGSG_03686.3	RT20-F	ACCGAATACCGCCGAGGACTTA
	RT20-R	AAGAAGGTTTGACAAGAGAGTGCG
Fg02138 FGSG_02138.3	RT21-F	GAGCGTATCATGGAACCCCTTGT
	RT21-R	GCGGTGATCTGGGTGTTTTCTG
Fg02118 FGSG_02118.3	RT22-F	CCGCCACATCAAGGAAGTCTACC
	RT22-R	GTCCTCGATGCTGGGTCTGA
Fg02117 FGSG_02117.3	RT23-F	ACAAAGCCTCTGTGTCGAAAATGG
	RT23-R	CAGGGTGCTCGTAGATGGTTGG
Fg02114 FGSG_02114.3	RT24-F	CTCACGAGTGTGTTGTTGGTTTG
	RT24-R	CTCCAAAAGACATCGGCAAAAT
Fg02111 FGSG_02111.3	RT26-F	AACCCGTCCGTGATTTCCACA
	RT26-R	CGACGGATGCCAAGATTAGATAGC
Fg02458 FGSG_02458.3	RT27-F	AGGTCGAACTCAATGAATGGATGC
	RT27-R	AGCTTGGCAGCCAGAGGAAATAGT
Fg05334 FGSG_05334.3	RT28-F	TGCCGAGAAGGACCCAGTAA
	RT28-R	GGACAACATCGTCTGTGAAGCGT
Fg02672 FGSG_02672.3	RT29-F	GTCGATCGAATACACCGTCCAAC
	RT29-R	GCGATGGTCGCTCTTACAGCA
Fg02668 FGSG_02668.3	RT30-F	GCCTGATCCTGGTTTGCTCTTTG
	RT30-R	ACAAAGGCGCAAGATCAAGGTATG
Fg02929 FGSG_02929.3	RT31-F	TTGGCTACAAGGCGCAGTATTTT
	RT31-R	TCCTTTTCCGACGGTAGCATAGAT
Fg02872 FGSG_02872.3	RT32-F	CGTGCACTTCTTGGCCTGTATCT

	02982.3-3R	TCCTCGGATGTCTAGTATGCGGTATC		RT32-R	TGGAACCAAAGGGATCAGTGTCA
	02982.3-5N	CACTTCTCGACAACAATCCGTAAACAG	<i>Fg03548</i> FGSG_03548.3	RT33-F	GTTCCCGGCCATCCTTTACATC
	02982.3-3N	TGGCTAGGAGTACAGAAATGAGAGTCG		RT33-R	TAGTGGGTGCATAGTGCCTGACAG
<i>Fg03264</i>	03264.3-5F	AGACTTGAAACCAGCACTGGAACATT	<i>Fg03542</i> FGSG_03542.3	RT34-F	AGCTACTTTGCGCGAATCCTCC
	03264.3-5R	gcacaggtacactgttttagagCTGCCATGTCAGCACATTCTTT		RT34-R	AAACACTAGGTCCATGGGCAAGAA
	03264.3-3F	ccttcaatcatcttctgtcgCTTGCTGGATCAAATTGTCTGTGG	<i>TRI11</i> FGSG_03540.3	RT35-F	GCCCAGGCCATGCTTGAAC
	03264.3-3R	CAGTTTGTATGTCAGTCGTAGTAGCCG		RT35-R	GACTCTCAAAGGCCAGAGCAACC
	03264.3-5N	AAAGGAATTGGAAGGGATGTTACGA	<i>TRI4</i> FGSG_03535.3	RT36-F	GTTCTCGTCGCGGTACCTGA
	03264.3-3N	TCTAGGAAGAGGCTCGCATCATTATG		RT36-R	TCGTTGTGTTGCCATAGAATCG
<i>Fg03260</i>	03260.3-5F	AATTTGTCAAGATGCTTTCAAGTCGC	<i>Fg03498</i> FGSG_03498.3	RT37-F	TAGAGTGGGTTAGGTACGCAGTCG
	03260.3-5R	gcacaggtacactgttttagagCAGACGTTTTTCGGAATGGATACAGA		RT37-R	CGTCGGTAGGGATATTCGTGTTGT
	03260.3-3F	ccttcaatcatcttctgtcgTTCGTTTGCCCGGGAACATTA	<i>Fg06068</i> FGSG_06068.3	RT38-F	CAACAAGGCCCTCAAGGACATCTA
	03260.3-3R	GCCTGCCTGCATCTTGTCTCT		RT38-R	GGATGTATCGCTGCATCTCTTTCA
	03260.3-5N	TCTGAAGACCAAGGAGGAGCTAAAAAT	<i>Fg01786</i> FGSG_01786.3	RT39-F	TTCACGCAATCAAAGCCAATCTC
	03260.3-3N	CACCTACCAGGTCCCATCTCATTC		RT39-R	GTCGAAGCAAGCCAGACATAACC
<i>Fg05806</i>	05806.3-5F	CTATCATGCCATCAGATCCCGAAC	<i>Fg01767</i> FGSG_01767.3	RT40-F	AGCACGCTACAGCCAAGAAGATG
	05806.3-5R	gcacaggtacactgttttagagCCACATCCCGATTACCCCAT		RT40-R	TTTGATAGACGCGCGGTGATG
	05806.3-3F	ccttcaatcatcttctgtcgGTTCCCAATCCTCTCAAGCATCT	<i>Fg01745</i> FGSG_01745.3	RT41-F	TTGCCAAAAAGTCCGAGGTCTG
	05806.3-3R	GTTACAGGGCCAATCACAAGGTCA		RT41-R	CCCCACTGAGAAAGCTTGAGGTAG
	05806.3-5N	GTTTGTGCTTTCTGTAAGTTGTAA	<i>Fg01740</i> FGSG_01740.3	RT42-F	AATCGTTCGTACGCACTACC
	05806.3-3N	GGCTGCCTACTCAGAGTTTCCG		RT42-R	CTCATAAAATCAGACGACCTGGG
<i>CYP51A</i>	04092.3-5F	ACCATCAAATGAACGCAGGGG	<i>Fg01739</i> FGSG_01739.3	RT43-F	GTTTCATCCCGAGAGTGTATCAT
	04092.3-5R	gcacaggtacactgttttagagGGAAGGCAACTACTCAAGCAGGG		RT43-R	ACCACTCTCGTGACAAGGAAGC
	04092.3-3F	ccttcaatcatcttctgtcgGAGTCAAGTTGTGTTAGCATCCCGAT	<i>Fg01722</i> FGSG_01722.3	RT44-F	GACGACGACGCTGAATCCAT
	04092.3-3R	GGGCTTTGCAGGGAGGTGTATT		RT44-R	CGAGTTGACCGAGTGGCTTCC
	04092.3-5N	TGAAGTTGTGCAGGATGTGGGTA	<i>Fg05113</i> FGSG_05113.3	RT45-F	GTACCCTTTGCCGCTGCTCA
	04092.3-3N	ATGCTTCCATGTCATGACTCCAC		RT45-R	AGCTCGTCTTGTGCCTTCTTTGT
<i>Fg01284</i>	01284.3-5F	TCTGGGCAAAGACGTTGTGGTC	<i>Fg01583</i> FGSG_01583.3	RT46-F	CACAAATCTCGCTCAACTCAAA
	01284.3-5R	gcacaggtacactgttttagagCTATGAACCCACGGACGTCAAGAG		RT46-R	CTACGGCTAGTGTGTACGCTGTGG
	01284.3-3F	ccttcaatcatcttctgtcgGATGCTGCAGTCTCGTGGTGTCT	<i>Fg01048</i> FGSG_01048.3	RT48-F	TTGCTATGCTACCGTGGCTTCTT
	01284.3-3R	CTCGTCCAGCACACTATGACACCTC		RT48-R	ACGTTGCATCTTGTGCTCATCG

	01284.3-5N	GAGTGCTTCGCCGGTTCAGAT	<i>Fg02419</i> FGSG_02419.3	RT49-F	CATTCGGCTTTTGGAGACAGACT
	01284.3-3N	ACACTTAAACAAGCAGCTCGCCATA		RT49-R	CAGGACCAAAGCGTTCAGCAGTA
<i>Fg03191</i>	03191.3-5F	CGACATTTCAGTTCGGCTACG	<i>Fg02371</i> FGSG_02371.3	RT50-F	TCTCTGACGCATCTCGCAACTG
	03191.3-5R	gcacaggtacactgttttagagAACACGGATATTGCAGTGAAGGG		RT50-R	TTCCGCAATAAGTTCATCGTCAGT
	03191.3-3F	ccttcaatcatcttctgtcgATGCATGTTACGTGGACTGGTAA	<i>Fg02367</i> FGSG_02367.3	RT51-F	CACAAATCTGCGCTCAACTCAAA
	03191.3-3R	TCCGCATACACTAGTCGAGCACATTG		RT51-R	CTACGGCTAGTGTGTACGCTGTGG
	03191.3-5N	ACAACGACACTGGTGATACCGAAAG	<i>Fg02366</i> FGSG_02366.3	RT52-F	AGGTGCCATTGCGCTTCTTGT
	03191.3-3N	GACCCGGCGTTAGTCAGAGACAT		RT52-R	CGGTCTTGGGCCCTTGAATG
<i>Fg03700</i>	03700.3-5F	CGACGCCATCTTCGCCTAAATA	<i>Fg02344</i> FGSG_02344.3	RT53-F	TCCGGGACTTTTCGCAGATG
	03700.3-5R	gcacaggtacactgttttagagCACAGCCGGCAGAAGGAGTAA		RT53-R	CCCAAGTAGCCAGGTTAGGTTCC
	03700.3-3F	ccttcaatcatcttctgtcgCTTCGTTGCCCAAGGATTCTCT	<i>Fg03796</i> FGSG_03796.3	RT54-F	GAACGCAAATGCCCTCAACTACA
	03700.3-3R	TGGTCCGTCATCCACATTTTCATC		RT54-R	CGGGGAAGATGTTAACCAGTAGCA
	03700.3-5N	ATAAGCGCGGGCAATAAATGACT	<i>Fg03741</i> FGSG_03741.3	RT55-F	TGATGACTGCGAGAAAGGGACAA
	03700.3-3N	GACCACGCTTATCAACGCTATCG		RT55-R	GGGTACGTAATCCTGCCAGTTGTT
<i>Fg03686</i>	03686.3-5F	GGCTTCCAATCCTTTCATCGTGT	<i>Fg00863</i> FGSG_00863.3	RT56-F	TGCTCTCGTTGCTCAGGCTATTG
	03686.3-5R	gcacaggtacactgttttagagAAAGATGGATGACCCCGAGAGAAG		RT56-R	GAGGTCATCAACTCGGTCAGGCT
	03686.3-3F	ccttcaatcatcttctgtcgAAGCTGAGCTACAAACTCCGCCTAT	<i>Fg00012</i> FGSG_00012.3	RT57-F	CCGAGACGACTGATGGCACAC
	03686.3-3R	CCCCTCTCTCCCAAGTGTTTC		RT57-R	TCAGTCTGTTTTTGGCTTGGTGGT
	03686.3-5N	TGCAGTGGCTCTTGTATCATCAT	<i>CLM2</i> FGSG_00007.3	RT58-F	CCTCCACAACCCAGAAACCTATCA
	03686.3-3N	TGCTGCCTCCATCAACTATCGC		RT58-R	ATCCTTTTCGATGTCAAATGCTGC
<i>Fg02138</i>	02138.3-5F	TGGCGTAGACGAGGCTGATTTG	<i>TRI1</i> FGSG_00071.3	RT59-F	CAACCTGTGCTACCCAATATT
	02138.3-5R	gcacaggtacactgttttagagAGTCGCTGGCTGGTCTACTGGTC		RT59-R	CGCTGTCGAGAAGGAACATCTTG
	02138.3-3F	ccttcaatcatcttctgtcgGATGAGCTGTGATTGGCTGAGTGA	<i>Fg00127</i> FGSG_00127.3	RT60-F	GGCCAGAAACTACCACCAATGT
	02138.3-3R	TCGAACCTACGGAAACGCTCAAAG		RT60-R	CGTTGAGCTTCATCCCGCATAG
	02138.3-5N	GGGCATCTTCTTGTGAGGAGCATA	<i>Fg08412</i> FGSG_08412.3	RT61-F	TTCGACTACCGCGCTTTCACA
	02138.3-3N	CTATCTCAGCTCGACCTGACCC		RT61-R	GCAAGGATAGACTGGGCACGAG
<i>Fg02118</i>	02118.3-5F	TCAAGAAACGCAACATGGAGACA	<i>Fg08377</i> FGSG_08377.3	RT62-F	CAGGCTGTGGTCCATGAGGTTAC
	02118.3-5R	gcacaggtacactgttttagagTGTGGATGAGGAGGGAATGGAAT		RT62-R	CGCATTCGAGCTCCCGTAAA
	02118.3-3F	ccttcaatcatcttctgtcgGAGATGCATGAAGCAAGGGTAGAGAG	<i>Fg08320</i> FGSG_08320.3	RT63-F	AGGTGCCATTGCGCTTCTTGT
	02118.3-3R	TGACGTTACACATCTTACCCTG		RT63-R	CGGTCTTGGGCCCTTGAATG
	02118.3-5N	TCCTGGAGATACTCGCCTCTGATG	<i>Fg08207</i> FGSG_08207.3	RT65-F	CCTCAAGCACGTTTACAGCAC

	02118.3-3N	TCCCTCAGTCTTGAGCAAGTGTCC		RT65-R	GGTGCACCATTTTCCGAGAGC
Fg02117	02117.3-5F	AATATCCAAGTGAGCCAAGCGATG	Fg01972 FGSG_01972.3	RT67-F	GCATTGCGACCCAACTTCTCTA
	02117.3-5R	gcacaggtacactgtttagagGCGTTTCTTGATGCTGGGAATAGA		RT67-R	GTCGGATAGCTTCTCGCTGTTT
	02117.3-3F	ccttcaatcatcttctgtcgTTAGCCTGTAGCTTGCCACTTGAA	Fg08187 FGSG_08187.3	RT68-F	TTGGAATGTCTATGGCTGCTGTTG
	02117.3-3R	GGCTTAGCAACAATCCCATCAGTG		RT68-R	ATCGAGCAATCGCATATTTGTAAGC
	02117.3-5N	TGATAAAACAATGCGCATAAGAGGC	CYP51B FGSG_01000.3	RT69-F	CTTTACTGCTTCCCATGCTCTCCA
	02117.3-3N	GGTAGTGTCCCTGAGAGAATCCCG		RT69-R	CGCGGCGCTCCTTGATAGT
Fg02114	02114.3-5F	GCGTGTGCCAGGTAAAAAGTTC	Fg08186 FGSG_08186.3	RT70-F	GGGCGCCTTCAATTGCTACAC
	02114.3-5R	gcacaggtacactgtttagagAGAGGTTTCGAGCCAGTGATTCA		RT70-R	ATCTGGTGCATGATCCTCTTGACA
	02114.3-3F	ccttcaatcatcttctgtcgAGGGATATGGTTGTTGCGTTATGG	Fg08183 FGSG_08183.3	RT72-F	TTATGGAGAACGGGTGGCAAAA
	02114.3-3R	AAACCTCCGTACTTCTGGCAAAGA		RT72-R	ATTTTGGCGTTGCGTCTCCTTA
	02114.3-5N	AGGGAGAGGTGCTATTGTGGAGTG	CYP51C FGSG_07977.3	RT73-F	ATCGCTCCATCAAAACACCATTCA
	02114.3-3N	CACTACAACCATGACGGCTCAGAC		RT73-R	TCGAATGCATAGCACTGGAGCC
Fg02111	02111.3-5F	GCCCTGGTGCCAAAATGTTCTAC	Fg08005 FGSG_08005.3	RT74-F	AAATATTTCGACGGTGACGCAGAC
	02111.3-5R	gcacaggtacactgtttagagCGTTCTTGACATATGGTGGTTCC		RT74-R	GGTGGCTGATGCGAGGAAATACT
	02111.3-3F	ccttcaatcatcttctgtcgCAAGACGCAGTGTTGGGAGA	Fg10629 FGSG_10629.3	RT75-F	ATTTGATGACGCCGACTTTTCTGT
	02111.3-3R	TTGCTACGAAAAGGAATGTTGGTG		RT75-R	TTGAGTGTCAATCCCAAAAGAGGC
	02111.3-5N	GGGACATAGCTTGATCACGAGAG	Fg10695 FGSG_10695.3	RT76-F	TGCGAAAGCGACATATCAAAAACA
	02111.3-3N	TGTGGAACGTCTCAGGGTATCA		RT76-R	GGTGAGACATGTAGCGGTGTTTC
Fg02458	02458.3-5F	CCTATGACGGCCAGATTTACTTC	Fg10910 FGSG_10910.3	RT77-F	CAATGCAGAGCTTTTGGAGGACA
	02458.3-5R	gcacaggtacactgtttagagGAGTCATCATCCAGGCAAAGAAAA		RT77-R	GGAAAGGGGCCAGACAGG
	02458.3-3F	ccttcaatcatcttctgtcgCAAGACGGACAAAACGAGCAC	Fg11389 FGSG_11389.3	RT78-F	TTTGATGGGTTTGCCGATTTTG
	02458.3-3R	GTCCAAACGATGACCCATACGAA		RT78-R	AGCCAAGCCGACACAAAACAT
	02458.3-5N	AGACCTGCGTATGTCGTGTGCTT	Fg11424 FGSG_11424.3	RT79-F	GATGAACCGGTATGCCAATGACA
	02458.3-3N	GCGGATCCTTCTGCGTCGTTAT		RT79-R	AACTGCGACGTAAGCATTGAGAGC
Fg05334	05334.3-5F	GCTTTGGCTTGCTTGCTCAGTG	Fg11282 FGSG_11282.3	RT80-F	TTCTCAAGCCCTTCCCCAACTT
	05334.3-5R	gcacaggtacactgtttagagAAGATATCGCCCAAGACGGTCATA		RT80-R	TGCAGGTAGTCGTTCCGCTCAG
	05334.3-3F	ccttcaatcatcttctgtcgGGAGATGAAATCTGCCGAAGAGA	Fg11303 FGSG_11303.3	RT81-F	AATCAGCGGTTTCTTGCGTCAC
	05334.3-3R	CCTCATAGGTGCGGGTTGATAG		RT81-R	CGACTGGTGATCTGGATGCTGTTA
	05334.3-5N	TCTTTGGTGCTTCATCATCGTC	Fg11038 FGSG_11038.3	RT82-F	GCTACTGTTTCGGTGAGGCTTTTG
	05334.3-3N	GACTTTCGCTCCGATGTTGTCAC		RT82-R	GTACGGACAAGCATCGCAGCAT

Fg02672	02672.3-5F	GCCGATTATGCTTGCTCTCTTAT
	02672.3-5R	gcacaggtacactgttttagagGCCTCCATCATGCATGTCTTGT
	02672.3-3F	ccttcaatcatcttctgtcgGGGAGTGTGAAGGAATGGA
	02672.3-3R	GCTACTCTCCTGGCCCTGTC
	02672.3-5N	CTTTGCGGTCAATCCTTGCCCT
	02672.3-3N	CCCAATACCCTCATCCACCTTA
Fg02668	02668.3-5F	TTCATCGCATGATTCTGGTTAG
	02668.3-5R	gcacaggtacactgttttagagGGCTTCGAACACCAGCAAAATC
	02668.3-3F	ccttcaatcatcttctgtcgCCCATCCAGGAATACTACTCGTTG
	02668.3-3R	TTACCAGCTGCACCAAGCCTATC
	02668.3-5N	TCTTGGTACATACGCGGCAGAAT
	02668.3-3N	CAAAGGTAGCGCGAAAGTATCCG
Fg02929	02929.3-5F	AATGATGGGCGTGGTTGACTTACT
	02929.3-5R	gcacaggtacactgttttagagACAAGGGCGAAACCTCTGGAAT
	02929.3-3F	ccttcaatcatcttctgtcgCAATGACCTTCTCCCATGTAACCA
	02929.3-3R	TCCATTGGCGACCGAAGTTTT
	02929.3-5N	GATAGGACAGCTTTCGGGACCAC
	02929.3-3N	CCAACAGCGCCTGCCTTAGAC
Fg02872	02872.3-5F	CTGATGTTCCCTCAGATGGTTCTCT
	02872.3-5R	gcacaggtacactgttttagagTCGCTATAGCCGTGTAGTCCAC
	02872.3-3F	ccttcaatcatcttctgtcgGGTGCGCAGATCAAGAGTAGACAA
	02872.3-3R	TCGTCAGCCATGGCAAGAACTA
	02872.3-5N	CACAGTACCGAGGCGACGAGAC
	02872.3-3N	CAGGGTTCTCTCCAGCTTTTGACA
Fg03548	03548.3-5F	AGGCGTGTATGAACGAGCGACA
	03548.3-5R	gcacaggtacactgttttagagGCAAAATGGGAGGCAACAATACT
	03548.3-3F	ccttcaatcatcttctgtcgCAGGATGGACCGATTGAGTATGAC
	03548.3-3R	CCTACTGACACGGCCACTGAGAA
	03548.3-5N	GAGTGGCTTCAGTTTGGGCTTCT
	03548.3-3N	GCACCTTCGAGCTGCATAAGTGT
TR111	03540-5F	GGTCCCGCCTGGCCTCTTTA

Fg11108 FGSG_11108.3	RT83-F	CCACTAACCACCGGAGACTACG
	RT83-R	ACCCTTCGCGTTGTGAGCAGT
Fg10960 FGSG_10960.3	RT86-F	GGAAGTCGATTGATACCCACCTA
	RT86-R	CGCAAAAACAGACTCGAAAATGG
Fg10991 FGSG_10991.3	RT87-F	GGATAGAGTGTGGCTGCGATGAT
	RT87-R	CTTGCTAGAAGGTTGCGGTCATTT
Fg11002 FGSG_11002.3	RT88-F	GCTGCTGCCGTAAGGAAGTC
	RT88-R	TAAATCCGTGGTCGTGTGAATGG
Fg11024 FGSG_11024.3	RT89-F	GAAGTCATGGCCGAAATCACAATC
	RT89-R	AGCATGGTTTCGACGGAAGTTTT
Fg11465 FGSG_11465.3	RT90-F	CAAATGGTGGCGATGTTTAGCAG
	RT90-R	TGGAGAAAGCTGGGTAGCATCACT
Fg11498 FGSG_11498.3	RT91-F	GAGCTTTTTTCCATTACCCGTCA
	RT91-R	TTCTGGCTGAGTCCATTTTTGAT
Fg11536 FGSG_11536.3	RT92-F	GTCAACATCCATCAGACCTCCTT
	RT92-R	TCTCTTGACCTCAGGCTGGCTCT
Fg10451 FGSG_10451.3	RT93-F	GGCCAAGAACGGTAAGCGAGA
	RT93-R	CTGTGGGTGTGTCTCTCACTTCA
Fg10461 FGSG_10461.3	RT94-F	GTTAAGGAGCTGACCCCCATTTTC
	RT94-R	TCTCGTGTCTGGGGTTGTGC
Fg09351 FGSG_09351.3	RT95-F	CAAGCGCAAGCCTGTGGACT
	RT95-R	AGCTCAGCGACCATCTTTTCACAA
Fg09195 FGSG_09195.3	RT98-F	CTGCCAGCCGAAACAAGGAGT
	RT98-R	GTCGACGCGCTGCTTCAAGA
Fg09916 FGSG_09916.3	RT100-F	AGCGAAGGCCAATCTGAAAATG
	RT100-R	GAATCTGCTGCAAGGCGTGTG
Fg09671 FGSG_09671.3	RT101-F	CAGCTGGTGGGGTTTTGTCT
	RT101-R	CTGCATCATGGTGGTGAAGGTCT
Fg11655 FGSG_11655.3	RT103-F	ACCGGTCAGGGGTCTACTAAAAAT
	RT103-R	TTCAAGGCCCGACCCAACCTT
Fg12534 FGSG_12534.3	RT104-F	TTTGACTTTGCGCCATTCTCG

	03540-5R	gcacaggtacacttgtttagagTCACGTCTTAGGTTCAACGCAGTA
	03540-3F	ccttcaatatcatctctgtcgCCTCAAAGCCAGCGATGATAGAT
	03540-3R	TCGGCGCAGAGACTACGGATAC
	03540-5N	AGGCATATCCGCTGGCAAATACT
	03540-3N	TCCATCTATCCAACCAATCGTGCT
TRI4	03535-5F	GTGAAGGTGGGAAGGGCGATAA
	03535-5R	gcacaggtacacttgtttagagTTGTTGAGTAGAAGCCGCCAGATA
	03535-3F	ccttcaatatcatctctgtcgCACTGTATCGGCATCGAATCCTG
	03535-3R	GAGTTCGACGACAAGTGACAGAAT
	03535-5N	GTCAAAGTCGAAGCAAACAAGTGGT
	03535-3N	AGGTATGAAATTTCCAGCAGGGCT
Fg03542	03542.3-5F	ATGGAACCGAGATGGCTCAAAATA
	03542.3-5R	gcacaggtacacttgtttagagGAGTCTTTGTGCTTACCTCTCCTT
	03542.3-3F	ccttcaatatcatctctgtcgGTTGTTTTGGCAAAATACTGGGC
	03542.3-3R	CCCAGGAGACACCAAGGAGAAAG
	03542.3-5N	GCCACTGCGTGTCTCAATAAACC
	03542.3-3N	AATGGTCGTCGCCCAATCTATC
Fg03498	03498.3-5F	TGGACAAGGAACCCATCACAG
	03498.3-5R	gcacaggtacacttgtttagagCACGAAGCATGATGATGACAACA
	03498.3-3F	ccttcaatatcatctctgtcgGCATCTGGGTAGAGTGATGACAGG
	03498.3-3R	AAGTCTCCGACACTCGTCAAAAGG
	03498.3-5N	GGCAGAGATCTACGGGTTCATT
	03498.3-3N	GTTTCTGTCCAGGCGGCTAATG
Fg06068	06068.3-5F	TTATGGCGTTGATGTGTGGCTTT
	06068.3-5R	gcacaggtacacttgtttagagACCCAGGTACCCACAGATG
	06068.3-3F	ccttcaatatcatctctgtcgGGACAAACAAGCATATGACGGA
	06068.3-3R	TGGCATTTCTGGTTGTTTCGCTGTA
	06068.3-5N	AACGAGCATCACATGGACTTTTGA
	06068.3-3N	GTTGTGATGGTTGACCAAAAACGC
Fg01786	01786.3-5F	GCGAGCAGCCCTACACCTTTACT
	01786.3-5R	gcacaggtacacttgtttagagGCAAGGTGAAGTTCTGCGAAAGTC

Fg12568 FGSG_12568.3	RT104-R	TTTGCCAGGAAAGTGCTCTTG
	RT105-F	GTACGTTGCCGGTTTGGTTGAA
	RT105-R	ACAACGCGGTCGATCTCTTCC
Fg08809 FGSG_08809.3	RT106-F	TCTTGTCACGCTTTACCCGAGTC
	RT106-R	GTAGATCGGGCCCTGGTAGACAG
Fg13176 FGSG_13176.3	RT107-F	GTGCTTAAACCGCGACTGCT
	RT107-R	ATGAGGTCGCGATGAATCTTGG
Fg12737 FGSG_12737.3	RT108-F	CACCAAGTTGCCGCTGACG
	RT108-R	GCTCGCTGCTCCATTCTGTTG
Fg12599 FGSG_12599.3	RT109-F	TCGGAATTCTTTTGGCAGTTG
	RT109-R	CAACCAATTTTTCGGTCTGCCTC
Fg13445 FGSG_13445.3	RT113-F	TGGCTATGGACGCGTTGACTG
	RT113-R	TTGGCGTAGGGCTTGAGTTGG
Fg13459 FGSG_13459.3	RT114-F	AGGAGCGTTCAGCCTCACAAAC
	RT114-R	AGCTTTTGCGCGAAGTTGACC
Fg14007 FGSG_14007.3	RT116-F	GGTTGGAGCCCGATAAGGATGT
	RT116-R	ACATCAAGCCCCAGTCCAACG
Fg13739 FGSG_13739.3	RT117-F	TTCGTCACCACTGGAGCAAAGATT
	RT117-R	CCGTTCTTGTGCAATAGCCAGTC
Fg13796 FGSG_13796.3	RT118-F	AAGGTCTCGGCTCCAAAGTTTC
	RT118-R	CCTCGGCGAGATGAAGAAGTTG
Fg13797 FGSG_13797.3	RT119-F	ACCCTCACTTTGGATCACGACTGT
	RT119-R	GCCTCGAAAAGAACGGGGTC

	01786.3-3F	ccttcaatcatcttctgtcgCGATCCCGATTACGAGATTTTGTT
	01786.3-3R	TCACGACGGGCTGAAAAAGAAAC
	01786.3-5N	CAAGGATTGGGCTGGATGCTC
	01786.3-3N	TGGATTGTTTGTGTCCCCGTGTA
Fg01767	01767.3-5F	TGACCACCGGCTACTCCACTTCTA
	01767.3-5R	gcacaggtacacttgtagagCAAAGGCCAACAGAACCACCAC
	01767.3-3F	ccttcaatcatcttctgtcgTCTCTCGGTGAGGTTACATCG
	01767.3-3R	CAAAAGAAGCGCCTCGGTGTAG
	01767.3-5N	CCTACCGTGCCCACTTACA
	01767.3-3N	TCATCAGCTTCATAGGCCATCTCA
Fg01745	01745.3-5F	ATGTTGCCCATGGTACTGGTTAGC
	01745.3-5R	gcacaggtacacttgtagagTGCTGGGCGAGGACCTTAGA
	01745.3-3F	ccttcaatcatcttctgtcgCGCTGCTCTTGATGATAGTTCCA
	01745.3-3R	CGGCCCTTCAAACAACCTCACCT
	01745.3-5N	CGCAACCAAGTGCACAAGTTTACA
	01745.3-3N	TTCGGGCACCAACACCTCTTC
Fg01740	01740.3-5F	CGTCTGCCTGTTTGTGGTTGACT
	01740.3-5R	gcacaggtacacttgtagagTCGGTAGTTCATGGGTGTGATAGG
	01740.3-3F	ccttcaatcatcttctgtcgGCACCTTGAGAGGATGATGGAAC
	01740.3-3R	AATTGTTCCACAAGTCCACGAGC
	01740.3-5N	TTCTTAGGCGCCTTTGTTATGACG
	01740.3-3N	TCGCGACGAGCTCAAGGACA
Fg01739	01739.3-5F	TGTCTTTGGGCGGAGAATCAT
	01739.3-5R	gcacaggtacacttgtagagGATGTTGAGGCTGTCCATCTTGAG
	01739.3-3F	ccttcaatcatcttctgtcgAAGGGTGTTCGCGTCACTGTC
	01739.3-3R	CTGGCGATGGCTTGAGTATCCT
	01739.3-5N	GCTCACAAGCCGGACATGGTAT
	01739.3-3N	GTCAAGGTAATGCCGTGAATCG
Fg01722	01722.3-5F	TCACATAGGGTCGGCAGACTTTTG
	01722.3-5R	gcacaggtacacttgtagagCCCAGAGACTATTTCCGGCTGTGA
	01722.3-3F	ccttcaatcatcttctgtcgCGGTAACCAAGGGGGACAAGT
	01722.3-3R	GGCTTCCAGTAAACCAACGTTTAT

	01722.3-5N	CGGTTGGTTAAAAAGCAGTGTGGT
	01722.3-3N	CGTTTTCCGCCAGAGATGAGTTC
<i>Fg05113</i>	05113.3-5F	TTGAGAGAACCCAGGACGTAATC
	05113.3-5R	gcacaggtacacttgtttagagCCATCTTGAGTAAACCTCAGCAGC
	05113.3-3F	ccttcaatatcatcttctgtcgCCCAGAGCACTTCGATTGTGTCA
	05113.3-3R	TTGTGCGCCATCATTCCATTG
	05113.3-5N	TGAGTCGCTATTGTGGATGGTGC
	05113.3-3N	CCTCGAGCGGCTGTTCATAGAT
<i>TRII</i>	00071-5F	TGATGACAGGTGAAAGAGGCTGGT
	00071-5R	gcacaggtacacttgtttagagGGTCTGTCAAGCCTTGAAGAGAT
	00071-3F	ccttcaatatcatcttctgtcgGCGATTGGAATTGGTACAGGATG
	00071-3R	ATGTATGAGCAGCGAGGTTAGCC
	00071-5N	ATAGATCATGGGGGAAACAGTAGGC
	00071-3N	CGACTTTACCGTTGGCTCTTTTCA
<i>Fg01583</i>	01583.3-5F	CCTTGCCAAGACCATTGAACATC
	01583.3-5R	gcacaggtacacttgtttagagGCTACCAAGAAGAATATGAGGGTT
	01583.3-3F	ccttcaatatcatcttctgtcgGATATCACGTTCCATCCCTTGAGA
	01583.3-3R	CGTGCCAAGAGAACGTATGAATGA
	01583.3-5N	TGACAATGACTCGGCACTGGTGT
	01583.3-3N	ACAGGATTCTTGCATCGGACCAT
<i>Fg01048</i>	01048.3-5F	GACAACGCAGGTCAACCTCATCA
	01048.3-5R	gcacaggtacacttgtttagagGCGTCTTAACCAACCCAGACCAAGG
	01048.3-3F	ccttcaatatcatcttctgtcgTTGGGCGTTGGTATTGGTTGA
	01048.3-3R	CTTGGACAGACGTGCCAGATTAGAG
	01048.3-5N	CCGGTAGCTCTAGTAGGGCATTTCAT
	01048.3-3N	CTGATACTGCTGTGGTCGTGTGG
<i>Fg02419</i>	02419.3-5F	GGGTGGGTGAGTGAGTAGCATACAA
	02419.3-5R	gcacaggtacacttgtttagagTAATTGGTTGGATGTGGACCTGAAG
	02419.3-3F	ccttcaatatcatcttctgtcgTTCCATCTGTGACCCTACAAAGCC
	02419.3-3R	GGAACAAGCCAATTGTCCGTAGTG
	02419.3-5N	CTACTTGCAGCGTCGTCCTATT
	02419.3-3N	TGCACCAACCAACAAATCCATA

Fg02371	02371.3-5F	GACAATAAGGCACCATACTGAAACCC
	02371.3-5R	gcacaggtacactgttttagagCGACGTAGATTGCAGATATGACCCA
	02371.3-3F	ccttcaatatcatcttctgctgTGCATCAGGATTTGGTACGGAGTC
	02371.3-3R	CACCAGCTCCCCTCATCATCCT
	02371.3-5N	GGGGTCGGAATGTGTGGATTTC
	02371.3-3N	CGACATTTACATCCACAGTCTTCC
Fg02367	02367.3-5F	TGCTCGCATGCTCCATTGAC
	02367.3-5R	gcacaggtacactgttttagagGTATCGTGATGGAAGTGTTCGGA
	02367.3-3F	ccttcaatatcatcttctgctgCGGTTGGAGGTTAGAATTTCGTAC
	02367.3-3R	AAAGGAAGACATCTCGAAATCTGGTG
	02367.3-5N	GGCATCGCTAACCTTTTTTGTCTG
	02367.3-3N	AGAGATCTGGAGATTGTTCTGCGTA
Fg02366	02366.3-5F	TCCCAGAAACACCACCGCATAG
	02366.3-5R	gcacaggtacactgttttagagGCAAAGCTCAAATAAGAGTCCAAAGG
	02366.3-3F	ccttcaatatcatcttctgctgGCAGGCGGATACTTTTACACA
	02366.3-3R	GCCTGTCTTCCGAAGTTTGATGTTT
	02366.3-5N	TAGCTTCTGTCCCAGTCCGTGTTT
	02366.3-3N	GGAGCTGGTTGACAAGTCGGATAA
Fg02344	02344.3-5F	CGCACCGCAGTGTTCAAAATG
	02344.3-5R	gcacaggtacactgttttagagAGAACAAGGACTCAGAGCGACCA
	02344.3-3F	ccttcaatatcatcttctgctgGACATTTACAGGCTCAACGTGCTCTA
	02344.3-3R	GCCGTGGGGAATGCTGTTTAC
	02344.3-5N	CTGGAAGGTGGGACACTAACATCAA
	02344.3-3N	CTTAGCTAGAACGGGCCAAGTCAATA
Fg03796	03796.3-5F	GACATGCATCCAGCCACAGTAATCT
	03796.3-5R	gcacaggtacactgttttagagAAACACCCAACTTCTGGCGTAAAT
	03796.3-3F	ccttcaatatcatcttctgctgGAGCCGCAGCATGGTATCAAAGT
	03796.3-3R	GCTTTACCCAGCCACGGAATA
	03796.3-5N	CCTGGCAATAAAGATTCTACTGGC
	03796.3-3N	GGAAAGTTCCTCTGGGCTGGTG
Fg03741	03741.3-5F	CTGAGCTTGTGCTGCCATCTTG
	03741.3-5R	gcacaggtacactgttttagagTCTGGCTCGGCTAGAAGCTTTACTG

	03741.3-3F	ccttcaatcatcttctgtcgCGGTGGATAAACAATTAGCAATGGA
	03741.3-3R	TAGCAGCCCAGGTCTCGTAGTCAG
	03741.3-5N	CAGGTGAATGCTTCGACTTCTTTCTC
	03741.3-3N	GCTCTTGTCGACGAACGCTGG
<i>Fg00863</i>	00863.3-5F	CGACGGTCAGCATCTTCTCTGTAA
	00863.3-5R	gcacaggtacacttgtagagTCTGGCTCGGCTAGAAGCTTACTG
	00863.3-3F	ccttcaatcatcttctgtcgCGTAGCACTTCGGAGGCTTTGAT
	00863.3-3R	GAGGAGGCGAAGCAAAGACAATC
	00863.3-5N	TCTCATCAGAAAGCCATCATTCC
	00863.3-3N	GCTGGGGTCTCCTCTGTCTTGATT
<i>Fg00012</i>	00012.3-5F	AAGGCAACTCCAACGGCGATA
	00012.3-5R	gcacaggtacacttgtagagTTTGAGACGAGGAGGGGATTG
	00012.3-3F	ccttcaatcatcttctgtcgGCAGGGTACGCCTTTTAAAGACTACA
	00012.3-3R	CTGAGAAGCAGTCAAGGAGAGAATGG
	00012.3-5N	GCCTATTAAATACCATTGCAAGCGTT
	00012.3-3N	ATGATAATGAACGGTCTACTCGCCC
<i>CLM2</i>	00007.3-5F	GTTGCAGATGCCAGCACACTTG
	00007.3-5R	gcacaggtacacttgtagagTGATAGGCAAGGGTTGGGTCC
	00007.3-3F	ccttcaatcatcttctgtcgCTTGCTAAGAGAAAATGGGGTTGA
	00007.3-3R	TGTCATTGCACCTGATCTTGAAACA
	00007.3-5N	AGATGGGCTGGAAATGACCGTAA
	00007.3-3N	GGCACAGGTAGATACCAGCGTCAT
<i>Fg00127</i>	00127.3-5F	TCCGAGATCCGAATTAATACCACATC
	00127.3-5R	gcacaggtacacttgtagagTCCACGTGATCACATTAACCATTCTC
	00127.3-3F	ccttcaatcatcttctgtcgCTCGCGCTCTGGGTTCTGAAG
	00127.3-3R	CAAATTTGGATTGCCATCTGTAGACA
	00127.3-5N	TACAAATGCAACAGCGGTGGTAGAG
	00127.3-3N	TTAGCGATACCCAACCTCATCCTG
<i>Fg08412</i>	08412.3-5F	TGATGTGATTGCTGCCAATGTAGG
	08412.3-5R	gcacaggtacacttgtagagGATGGTTGTGTTAAGAGTGGAAGGGA
	08412.3-3F	ccttcaatcatcttctgtcgCTTGGGCTTCAACGACAAACTGAT
	08412.3-3R	TGAATCCTAGGCAAGATACTCCCA

	08412.3-5N	ATATGGGATATCGCGCTCTTGGTAA
	08412.3-3N	CTCCAAGATCCACAAGAGGCTATGTT
Fg08377	08377.3-5F	CAGCCGAGCAGATCATAGACAGGT
	08377.3-5R	gcacaggtacacttgtttagagAGAAGTTGTGTCGGTTTCACGGAC
	08377.3-3F	ccttcaatcatcttctgtcgTTGACCCCTGGTATCGCTATGCT
	08377.3-3R	CAAGGCATGACGGAGACACAGAA
	08377.3-5N	GCGAGTCAATCCGGTGTATCTGT
	08377.3-3N	TCGCCAACCAACATAAAACAGACC
Fg08320	08320.3-5F	ACGAAGGAGCCGACGTATTTT
	08320.3-5R	gcacaggtacacttgtttagagGGAAGGAAGGAGGCTTGTGGAG
	08320.3-3F	ccttcaatcatcttctgtcgGCTGCCACATATCATGATTCAAGAA
	08320.3-3R	GGTACTCGGAAAAGCGACAAACAG
	08320.3-5N	ATTCAAACCGTCGAGCTTAGACCC
	08320.3-3N	TGAGAGTCCTTGGGGCTTTTGTGA
Fg08207	08207.3-5F	GATTCAAGAAACCATGCCTCCGAT
	08207.3-5R	gcacaggtacacttgtttagagCAGTGTGTGATTGGGTCGTGTTTG
	08207.3-3F	ccttcaatcatcttctgtcgCTGAGATTGGCTTTGTTGGAATGAA
	08207.3-3R	GCTCTGGTTTTAGCGTCGGGTAG
	08207.3-5N	GGAAATGAGCGAGGTCTTGGAGA
	08207.3-3N	GATATTGCGTCTTACCCCTGAGAG
Fg01972	01972.3-5F	CGATGAACTTTGGGAGATGTGGAC
	01972.3-5R	gcacaggtacacttgtttagagAGGTGAAGATGCAGGATGAAGGAGT
	01972.3-3F	ccttcaatcatcttctgtcgATCTGTTCAGGAATCCTCCGTTGAT
	01972.3-3R	ACACACCAACCAAGGCTTTGTATTTC
	01972.3-5N	GAGGGAGAACTGTCGCAAATCAA
	01972.3-3N	TGGGAGCTAACCAACCTTCCAGTATA
Fg08187	08187.3-5F	GCAGTGCTCTCGTTGAAGTGGTT
	08187.3-5R	gcacaggtacacttgtttagagAGTCCCCACACCTAGCGTAGTG
	08187.3-3F	ccttcaatcatcttctgtcgGGAGGTCAAGCGCAGGTCATAG
	08187.3-3R	GATGACGTCCAGGTGCCAAAGT
	08187.3-5N	TGCGCTACCAGACACGAAAAGT
	08187.3-3N	ACTTCGACCTGGCTTTCCTGTGTA

<i>CYP51B</i>	01000.3-5F	CGGTATCGTAGCCTCGACTCCAT
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	01000.3-3F	ccttcaatcatctctgtcgGGGAGCGACGCCAGTAAACTTA
	01000.3-3R	ACAGGCACACAAAACACTCCAGC
	01000.3-5N	ATGGGCCATCAACACCTCAAAA
	01000.3-3N	CTTAGATAGCGGGCCAGGGAGAT
<i>Fg08186</i>	08186.3-5F	AATGGTTCCTCCCGCTTGTA
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	08186.3-3F	ccttcaatcatctctgtcgTTTGCCTTGGTAAAGGTGAGTC
	08186.3-3R	GGCTGCTGGCACATCACTTACAT
	08186.3-5N	CCTGAGGTGTACGAGAACCAGAC
	08186.3-3N	CCCAGACATTTGCGTCTCAGTAT
<i>Fg08183</i>	08183.3-5F	CCTGCCACTCATTCTCAACACC
	08183.3-5R	gcacaggtacacttgtagagATAACGCCGATGAGGATGATAGGA
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	08183.3-3R	CGGCTAAAGCTCCCTCAACACTG
	08183.3-5N	GTGCTACTCCGTTGTGACTCCCTC
	08183.3-3N	GGACTTGGCCCAGGAGTAAATGA
<i>CYP51C</i>	07977.3-5F	CCCAGGCAATGGTGACAGGCTA
	07977.3-5R	gcacaggtacacttgtagagCCAGTTAAGGAACGGCATCTTTGT
	07977.3-3F	ccttcaatcatctctgtcgTAATTTCTTTGTCCGGTTCGTTT
	07977.3-3R	GTA CTGCAACCAACTGGTCCGAG
	07977.3-5N	CAGAGTAAGCATAAACGCCGCAC
	07977.3-3N	GACCATGGCCAACACAATACGAG
<i>Fg08005</i>	08005.3-5F	ATGGATGCGCCCAACAGAAAT
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	08005.3-3R	ACAGTCATCAGCATCGCAAGACAT
	08005.3-5N	ATTCGGGCTGTATCTATCTCACGG
	08005.3-3N	CCAAACCTGGTTTTCGGACTAT
<i>Fg10629</i>	10629.3-5F	AAATCGAGCAGGACGCTAAGAACA
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	10629.3-3F	ccttcaatcatcttctgtcgCACCCGGATCGAGTCTTTATCAA
	10629.3-3R	TAGAGGGGGTTGCAGCGTGAA
	10629.3-5N	ATTCAGTCCACTCACCGAGAACCA
	10629.3-3N	GACTCATTGCGCAACCAACATA
<i>Fg10695</i>	10695.3-5F	GATGTCCATGAAGACGGGTTGTG
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	10695.3-3F	ccttcaatcatcttctgtcgGTGGAAAAGCCTCGTTTGATGG
	10695.3-3R	CTGTTCTGGACATGGCGGTGAG
	10695.3-5N	GGGCAGAATGCGTTCTATGACAG
	10695.3-3N	TATCATGTCGTCAGGGTTGGGCT
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	10910.3-3N	TAGACGTTCCGTTTTCACAGCAA
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	11389.3-5N	AGAAACTTCCGAAGGTTCCGAGG
	11389.3-3N	CGTTTTCACTCGCGGTCCTATG
<i>Fg11424</i>	11424.3-5F	ATTCAATCATGGTTTCTGGTCTCGG
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	11424.3-5N	AGGCGGACGGTATCTTAGGCAA
	11424.3-3N	TGAGTTGAGGATGAAAGGAAACG
<i>Fg11282</i>	11282.3-5F	CGCATGCGCTGTCTCAGTAAGTAG
	11282.3-5R	gcacaggtacacttgtttagagACCAAATCTGTAGCCGCCTGTATC
	11282.3-3F	ccttcaatcatcttctgtcgGTCGAACGATTTGTGCGGAGA
	11282.3-3R	GACGTCGATACCCAGTGCTTGTG

	11282.3-5N	TGCTTCCATATGCTACCGTCCCT
	11282.3-3N	CAGGCTGACTCCAAGAGCAATGTT
<i>Fg11303</i>	11303.3-5F	AGAGGAGGAGTTGGATGGTGAGGT
	11303.3-5R	gcacaggtacacttgtttagagCATGTTGACGGTGGTTGGTATCA
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<i>Fg11038</i>	11038.3-5F	ATCTGCATGCTCCAAGAGACGACT
	11038.3-5R	gcacaggtacacttgtttagagTCGCACCCTGAACAGAGTCGTA
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<i>Fg11108</i>	11108.3-5F	ATCGCTGCAGTCGTTTCTTTGA
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	10991.3-5R	gcacaggtacacttgtttagagGCCAAAGCGACAATAGCTCTCATT
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<i>Fg11002</i>	11002.3-5F	TGAATTTGTCGACATCCCGCTTAT
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	11002.3-3R	GGGAGCAACTAGACACTTATCGCAAA
	11002.3-5N	GTTTCGATCATTGGGGCAAAGGTAG
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	11024.3-5R	gcacaggtacacttgtttagagCTCAGGCCGTAGACTGATAGTGGTG
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	11024.3-5N	AACATAGCAGGCAATATAGCGAAACC
	11024.3-3N	CAC TTCCTCTTGATAGGTCTTTCGTC
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	11465.3-5R	gcacaggtacacttgtttagagCCGACGCCACATGAGGTATGA
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	11465.3-3R	TCACTGACTGATCGTCTGGTTCCTAA
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<i>Fg11498</i>	11498.3-5F	CCCTGAATACTATAAGCTTTCGCCCT
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	11498.3-3F	ccttcaatcatctctgtcgTCATGACCACTATTATTCACAGCCTTTG
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	11498.3-5N	GGTGGTGGTGCTCAAGATTTACTCC
	11498.3-3N	CTTAGCTCCCAGCAAGCACGAT
<i>Fg11536</i>	11536.3-5F	CGACTCGTTTGGTCCCTATTTCCTC
	11536.3-5R	gcacaggtacacttgtttagagCTGGTCGACTACGCATGGCTAAAT
	11536.3-3F	ccttcaatcatctctgtcgCTAGCGAGGAGGACAGCGACTCT
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	11536.3-5N	TCGACTCGCATGGGTTTCCTC
	11536.3-3N	GTGGAACAAGGCATCGTTCAGTCT
<i>Fg10451</i>	10451.3-5F	AAGGAAAGAAAAGAGAGGGCGAAA
	10451.3-5R	gcacaggtacacttgtttagagTCATTTCAACGCCGCATTCT

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	10451.3-5N	AGTGGTCGCGTGTTTGGTGTTT
	10451.3-3N	CAAAACCTTCCCACGACAGCAC
Fg10461	10461.3-5F	GAGTCATGGCAGAACTAGAAAGGGA
	10461.3-5R	gcacaggtacacttgtttagagCGTGATGCGATGAAGCTGATTTC
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	09351.3-5N	GTGAGCTGAAGGAAAAAGAAAGTTGC
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Fg09195	09195.3-5F	AGCATCCCAGCAAGCCCACTA
	09195.3-5R	gcacaggtacacttgtttagagTGACAGGAAACCAAGTCTTGCTTACG
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	09195.3-5N	TCTATAGAGCTTACAAACCAACCAACCC
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	09916.3-3N	ACAATGCTGAAAGTTAGGGGGTCC
Fg09671	09671.3-5F	GCTTCATGTGGATGATACCCGAGT
	09671.3-5R	gcacaggtacacttgtttagagGGGTGGAACCACAAGCGATAAA
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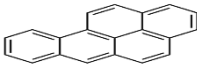
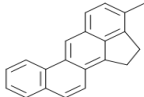
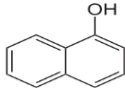
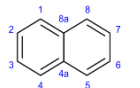
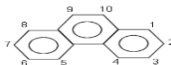
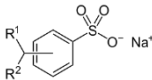
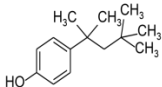

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	11655.3-3R	TCGTTGTTCCTGAGCCCTACCTC
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	11655.3-3N	TCCATACCCTCTGAGAAGCAATCG
<i>Fg12534</i>	12534.3-5F	CGAGGAAGCTAATAACCGCAAAGA
	12534.3-5R	gcacaggtacacttgtttagagTTGAGCCAATTGTGAGAAACGAGA
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	12534.3-5N	GCACAGACGTCAACTGGTAAGCAT
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<i>Fg12568</i>	12568.3-5F	ATGGAGCTACAGGACGGTGATGC
	12568.3-5R	gcacaggtacacttgtttagagCAAACAATTGCCCCAGAACAGAA
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	08809.3-5R	gcacaggtacacttgtttagagTTACGGGCAGCAATGGTCTCTT
	08809.3-3F	ccttcaatatcatcttctgtcgACGCGAGTCCGAGATTGTGAG
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	12737.3-3R	AAGATAGTTGAGGATGGGCGGAG
	12737.3-5N	GCCGCCAGTTTCTATCACTTACGA
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	13445.3-3F	ccttcaatcatcttctgctgGGGATTGGCTCGACGATAAATGTA
	13445.3-3R	TTATCGTGGGAACGGCATTATTG
	13445.3-5N	TGTTCAACGCGCCTTTTGGTATT
	13445.3-3N	GCTTCCACCATGCCGTCAG
<i>Fg13459</i>	13459.3-5F	AAGGAGTGTAAGTGGGTCGTGTGG
	13459.3-5R	gcacaggtacacttgtttagagGCGACTATCTTGCAAATGGAGGTT
	13459.3-3F	ccttcaatcatcttctgctgTCGCCTGAGGTAGACATCTGGAAT
	13459.3-3R	CCATCCACAATGTCCGCAAAAG
	13459.3-5N	GTCGGATCACCCATCAAAGTCAAG
	13459.3-3N	ATCCATGCAGAGGAGACGGTGA
<i>Fg14007</i>	14007.3-5F	AAGGGGTTGCCATTTCGTGAC
	14007.3-5R	gcacaggtacacttgtttagagTTCTTCTGCTTCCTTGTGCTCG
	14007.3-3F	ccttcaatcatcttctgctgGAGGGTCTCCCGATCATATAAACATT
	14007.3-3R	AACCGACAAATGTAACACCGCAT
	14007.3-5N	AGGCTCGAGAGAGAATATGGTGGC
	14007.3-3N	CGGCCCTATATCACGACCAAACT
<i>Fg13739</i>	13739.3-5F	CAGGGGGAGCTCTGGGATAAGA
	13739.3-5R	gcacaggtacacttgtttagagTGATGAGGTGTCGCATAGAAGGAA

	13739.3-3F	ccttcaatatcatcttctgtcgGGACAGCATGTTGCAGTCTTTGAA
	13739.3-3R	CCCAGTTTAGGCGCTGACTTCAA
	13739.3-5N	CAGCTGCTTTGGGAATTCTTTTCA
	13739.3-3N	GTGTGAGCCATTCCCTGTAGCCT
<i>Fg13796</i>	13796.3-5F	TCGTGAGAGGGCCAGACAATGTA
	13796.3-5R	gcacaggtacacttgtttagagTCCAGGGATTTTCAGGACTTTGAG
	13796.3-3F	ccttcaatatcatcttctgtcgGATGAGAGCCTCCGAAGCAAGTC
	13796.3-3R	GGGCACCGATCACGATAGTTTTC
	13796.3-5N	GCGACGATCTGTTGCCACTAAAG
	13796.3-3N	TGGA AACATGCTTGGTGGTCGTA
<i>Fg13797</i>	13797.3-5F	CACATACGTGCGGGATTGAACTC
	13797.3-5R	gcacaggtacacttgtttagagTTCTTACATTGTCTGGCCCTCTCA
	13797.3-3F	ccttcaatatcatcttctgtcgAAGCGTGGCTGATGGTTGAAGTAT
	13797.3-3R	ACAAGCCATCGTTCGGGGTAA
	13797.3-5N	CGGTCTTAGTCAGACGGTCCAGTG
	13797.3-3N	GATACGTCTCGCGAAGACTGGAAC

Table 4. List of 15 xenobiotics.

Group	Compound name	Concentration n (mM)	Structure	Description ^a
Aliphatics	n-Dodecane	176.12		Dodecane's production and use as a solvent, in organic synthesis, distillation chaser, in jet fuel research, standardized hydrocarbon, manufacture of paraffin products; rubber industry, paper processing industry, and as a component of gasoline may result in its release to the environment through various waste streams.
	1-Dodecanol	22.35		1-Dodecanol's production and use in detergents and soaps, in fragrances, and to a lesser extent in wetting, emulsifying, and foaming agents may result in its release to the environment through various waste streams.
	Levulinic acid	9.77		Compounds or agents that combine with an enzyme in such a manner as to prevent the normal substrate-enzyme combination and the catalytic reaction.
Aromatics	Methyl benzoate	14.69		Methyl benzoate's production and use as an additive in perfumes, disinfectants, soy sauce, and pesticides, an additive or preservative in cosmetics and personal care products, and as a solvent or dye carrier may result in its release to the environment through various waste streams.
	4-Nitrophenol	0.14		4-Nitrophenol is the most toxic of the mononitrophenols.
	Phenoxyacetic acid	0.53		Phenoxy acetic acid (an acid C ₈ H ₈ O ₃) used as a fungicide and in plasters and pads to soften callused skin surfaces.
	Resorcinol	0.72		Resorcinol's production and use as chemical intermediate, in the production of sunscreen preparations for the skin, production of dyes, manufacture of special pharmaceuticals, manufacturing adhesives, explosives, tires and rubber products may result in its release to the environment through various waste streams.

Polyaromatics	Benzo(a)pyrene	0.32		There is no commercial production or known use for benzo(a)pyrene; it is released to the environment as a product of incomplete combustion. Benzo(a)pyrene is found in fossil fuels, crude oils, shale oils, and coal tars, and is emitted with gases and fly ash from active volcanoes.
	3-Methylcholanthrene	0.15		3-Methylcholanthrene's production and use as an experimental cancer research chemical may result in its release to the environment through various waste streams.
	1-Naphthol	0.14		1-Naphthol's production and use in the production of agrochemicals, such as carbaryl, drugs, rubber oxidants and dye intermediates, may result in its release to the environment through various waste streams.
	Naphthalene	0.62		Naphthalene's production and use in the manufacture of phthalic anhydride, which is used as an intermediate in the production of phthalate plasticizers, resins, dyes, pharmaceuticals, insect repellents, and other materials may result in its release to the environment through various waste streams.
	Phenanthrene	0.11		Phenanthrene's production as a result of incomplete combustion of organic materials, will result in its release to the environment through various waste streams.
Alkyl-substituted aromatics	Sodium dodecylbenzenesulfonate	0.06		Sodium dodecylbenzenesulfonate's use as an anionic detergent results in its release to the environment through various waste streams.
	4-Octylphenol	0.1		This substance may be hazardous to the environment. Special attention should be given to fish.
Alicyclics	Cyclohexanone	48.2		Cyclohexanone may be released to the environment as a result of its manufacture, industrial use, transport, and disposal. Its major uses are as an intermediate in the production of adipic acid and caprolactam, which are used to make nylon 66 and nylon 6, respectively. It is also used as a solvent for materials such as synthetic resins, polymers, lacquers, inks, paint, and spot removers and a chemical intermediate in the production of plasticizers, insecticides, and other chemicals.

^a Description of each chemical came from The Pubchem Project (<https://pubchem.ncbi.nlm.nih.gov/>).

요약 (국문초록)

*Fusarium graminearum*의 cytochrome P450 효소들의 기능연구

신지영

붉은곰팡이(*Fusarium graminearum*)는 전 세계적으로 주곡작물인
벼, 보리, 밀, 옥수수 등에 큰 피해를 주는 식물병원균이다. 붉은곰팡이
는 수확량의 감소 뿐 아니라 Trichothecene과 Zearalenone과 같은 곰
팡이 독소를 생성하여 인축에 직접적인 피해를 유발한다. Cytochrome
P450은 자연계의 거의 모든 생물 중에 존재하며, 약물, 발암물질, 독성
물질, 스테로이드 등의 대사에 관여하는 물질이다. 현재 제약산업, 암 발
생, 살충제 개발, 미생물을 이용한 생물산업, 환경산업 등의 다양한 분야
에서 그 중요성이 대두되고 있다. 특히 cytochrome P450은 물질의 독
성을 증가시키는 기능으로 인해 병 발생 원인으로 주목 받고 있다. 따라
서 곰팡이 cytochrome P450에 대한 게놈수준에서의 접근이 필요하
는 인식이 대두되고 있지만, 식물병원균에서 전반적인 cytochrome
P450의 기능 연구는 미흡한 실정이다. 붉은곰팡이에서 cytochrome

P450은 병원성에 관련되어 있는 Trichothecene 독소를 생합성 하는데
관여한다고 알려져 있고 곰팡이의 생존에 필요한 막 지질인 Ergosterol
생합성에 관여한다고 알려져 있다. 이번 연구에서 붉은곰팡이의
cytochrome P450의 기능적 분석을 위하여 생물정보학적 분석을 통해
119개의 P450 유전자를 동정하였고 이 중 102개의 유전자를 삭제한
돌연변이체를 확보하였다. 102개의 유전자 삭제 돌연변이체를 병원성 실험,
외래물질 분해 등 38개의 조건에서 관찰 하여 phenotypic dataset
를 구축하였다. 이번 연구에서 구축된 붉은곰팡이의 cytochrome P450
mutant library는 붉은곰팡이의 대사 물질 연구(metabolomics) 분야 개
척의 발판이 될 수 있을 것이다.

주요어: 붉은곰팡이, cytochrome P450 (사이토크롬 피450), 병원성, 외
래물질 분해, phenotypic dataset (표현형 데이터 모음), mutant library

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